Georgia Quantitative Biosciences APPLY BY DECEMBER 1ST

QBIOS.GATECH.EDU

QBioS Research and Training: Spanning Molecules to Ecosystems

An Interdisciplinary Graduate Program

The Ph.D. in Quantitative Biosciences (QBioS) is a new interdisciplinary program at Georgia Tech, founded in 2015 and supported by a consortium of more than 50 program faculty from six home schools in the College of Sciences.

A Novel and Flexible Training Program

Featuring a flexible training program, including:

- Foundational courses in Quantitative Blosciences

- Rotations in modeling and/or experimental groups

- Selection of thesis advisor from all program faculty
- Rigorous and personalized quantitative training
 Five-year program of study from entrance to
- defense

Quantitative Biosciences integrates the physical, mathematical and biological sciences, enabling the discovery of scientific priociples underlying the dynamics, structure, and function of living systems.

Home Schools

Biological Sciences, Chemistry & Biochemistry, Earth & Atmospheric Sciences, Mathematics, Physics, & Psychology



Sciences Physics

Physiology

A Growing QBioS Student Community

 25 students in first three cohorts, in total.
 Feature diverse backgrounds and scientific training.
 Multiple award winners (including 2 NSF GRFP awardes).
 Students come from the USA (12), Chins (4), Mexico (3), Bosnia, France, India, Iran, Israel, and Romania.



JOY PUTNEY Inaugural cohort & NSF GRFP awardee (2017)

When I was considering graduate schools, the Quantitative Biosciences program at Georgia Tech stood out because of its interdisciplinary approach to the biological sciences. My campus visit impressed me with the quality of the faculty, both in terms of their world-class research and care for students. Both before and after I accepted the admission offer, it was clear that program faculty were invested in each individual student and wanted us all to succeed.



Bacterial cell wall structures in atom-scale simulations



Simulating high energy shocks in a fibrillating heart

> Biomechanic principles of subsurface "swimming"

viruses

Clusters and waves of

infections of microbes by



Biogeochemical dynamics resulting from the coupling of physical and biological processes

Contact Information: Director of OBioS

Joshua S. Weitz, Professor School of Biological Sciences School of Physics director@gbios.gatech.edu

Admissions

Lisa Redding Academic Program Coordinator QBioS Program admissions@qbios.gatech.edu QBioS is now accepting applications from students interested in integrating quatitative methods with bioscience research.

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Cohorts, from 2016-2018 from bottom to top.











ICTP-SAIFR SUMMER WORKSHOP MATHEMATICAL MODELS OF EVOLUTION SAO PAULO JAN 21-26, 2019 SUPPORT: ARO; NSF – DIMBIO; NSF – BIO OC; NSF POLS SIMONS FOUNDATION - SCOPE, GEORGIA TECH

VIRUS ECOLOGY AND EVOLUTION:

PRINCIPLES & APPLICATIONS OF VIRUS-MICROBE DYNAMICS

Joshua S. Weitz School of Biological Sciences and School of Physics Graduate Program in Quantitative Biosciences Georgia Institute of Technology



Joshua S. Weitz, Georgia Tech, School of Biological Sciences & Physics Email: jsweitz@gatech.edu, Twitter: @joshuasweitz Web: http://ecotheory.biology.gatech.edu



	Replica																			
Experiment	1	2	3	4	5	6	7	8	9	10	11	12	13	14	15	16	17	18	19	20
1	10	18	125	10	14	27	3	17	17											
2	29	41	17	20	31	30	7	17												
3	30	10	40	45	183	12	173	23	57	51										
4	6	5	10	8	24	13	165	15	6	10										
5	1	0	3	0	0	5	0	5	0	6	107	0	0	0	1	0	0	64	0	35
6	1	0	0	7	0	303	0	0	3	48	1	4								
7	0	0	0	0	8	1	0	1	0	15	0	0	19	0	0	17	11	0	0	
8	38	28	35	107	13															

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MUTATIONS OF BACTERIA FROM VIRUS SENSITIVITY TO VIRUS RESISTANCE^{1,2}

S. E. LURIA³ AND M. DELBRÜCK

Indiana University, Bloomington, Indiana, and Vanderbilt University, Nashville, Tennessee

Received May 29, 1943

GENETICS 28: 491 November 1943

TABLE 2

The number of resistant bacteria in series of similar cultures.

					· · · · · · · · · · · · · · · · · · ·							
EXPERIMENT NO.	t	10	tτ	15	16	17	31A	21b				
Jumber of cultures	9	8	10	10	20	12	19	5				
olume of cultures, cc	10.0	10.0	10.0	10.0	, 2 [*]	. 2*	. 2	10.0				
folume of samples, cc	.05	.05	.05	.05	.08	.08	.05	.05				
Culture No.												
I	10	29	30	6	I	I	0	38				
2	18	41	10	5	•	0	0	28				
3	125	17	40	10	3	0	0	35				
4	10	20	45	8	•	7	0	107				
5	14	31	183	24	0	•	8	13				
6	27	30	E 2	13	5	303	I					
7	3	7	173	rós	•	•	•					
8	17	17	23	15	5	0	1					
9	17		57	6	0	3	0					
IO			51	10	6	48	15					
II	•				107	I	0					
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19					0		0					
20					35							



S.E. Luria



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10			51	10	6	48	15					
II					107	I	0					
L 2					•	4	Ð					
13					•		19					
I 4					0		0					
15 .					r		0					
сó					o		17					
17					0		11					
r8					64 ⁻		0					
19					o		0					
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M. Delbruck

Nobel Prize, 1969 (w/H. Chase)







A2. Exposure of population to viruses

A3. Subpopulation of bacteria acquire resistance and survive viral infection.





B1. Growth of a bacterial population from a single ancestor



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B2. Exposure of population to viruses

··· § § § § § § § § §

B3. Subset of resistant bacteria already present and survive viral infection.





	Replica																			
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Viruses impose a strong selection pressure.

Host mutations that confer resistance are beneficial.

Hence, viruses induce host evolution.

But, what about the viruses?

MUTATIONS OF BACTERIAL VIRUSES AFFECTING THEIR HOST RANGE¹



For decades, this dogma persisted...

"the coevolutionary potential of virulent phage is less than that of their bacterial hosts"

-Richard Lenski & Bruce Levin, Am. Nat. (1985)

until....

1989 - Numbers



Bergh et al., Nature 1989

"We have found up to 2.5×10^8 virus particles per ml in natural waters... 10^3 - 10^7 times higher than previous reports."

1999 - Functioning



"Viruses divert the flow of carbon and nutrients... by destroying host cells and releasing the contents of these cells into the pool of DOM in the ocean."



"We report a genomic analysis of two uncultured marine viral communities. Over 65% of the sequences were not significantly similar to previously reported sequences, suggesting that much of the diversity is previously uncharacterized."

What We Talk About When We Talk About Viruses



Ebola Virus Image source: CDC



John Moore, Getty Images (Nature, 2014)



Zika virus core Sirohi et al. Science, 2016



Source: CNN



Influenza virus virology.ws



Source: CDC

And viruses infect organisms across the diversity of life



And viruses infect organisms across the diversity of life, sometimes strangely









Michael Grove, NPR, 2011



Viral Ecology and Evolution Lectures at the Interface

From Ecology to Evolution (Lectures 1-2)

Principles of eco-evolutionary dynamics: Monday Jan 20

Dynamics in complex communities: Wednesday Jan 22



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Friday Jan 25



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Principles of eco-evolutionary dynamics: Monday Jan 20

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From Lysis to Latency (Lecture 3)

Friday Jan 25

From Theory to Therapy (Lecture 4)

Saturday Jan 26

Throughout: theory and modeling motivated by fundamental eco-evolutionary challenges & real world applications.



Problems in Quantitative Viral Ecology From Structure to Dynamics

• How does viral infection change microbial population dynamics?

3

- How does (co)evolutionary change alter viral-host population dynamics?
- What is the relationship between infection networks and host-viral dynamics in complex communities?



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Part 1:

How does viral infection change microbial population dynamics?

Over 80 years ago, Volterra was convinced by his son-in-law, Umberto d'Ancona, to examine the fluctuations of the Adriatic fisheries





Fluctuations in the Abundance of a Species considered Mathematically.¹ By Prof. VITO VOLTERRA, For. Mem. R.S., President of the R. Accademia dei Lincei.

The first case I have considered is that of two associated species, of which one, finding sufficient food in its environment, would multiply indefinitely when left to itself, while the other would perish for lack of nourishment if left alone; but the second feeds upon the first, and so the two species can co-exist together.

NATURE

[October 16, 1926



In turn, Volterra & Lotka proposed a coupled pair of ODEs to describe predator-prey dynamics

Model

$$\dot{N} = aN - bNP$$
$$\dot{P} = cNP - dP$$

N: prey abundance P: predator abundance

Interactions:

Prey birth/death Predation Predator death



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Conservative system (not true limit cycles)

 $a \log P - bP + d \log N - cN = \text{const}$



Later models included limit cycles, consistent with long-term observations

Features of limit cycles in predator-prey models

- I. Prey peaks before predator peaks.
- 2. Predator oscillations are quarter-phase lagged behind prey oscillations
- Hence, oscillations appear counter-clockwise in preypredator phase plane

[Models include "handling time" of prey by predators and limited prey growth]





"One Resource, One Prey, One Predator"

Vol. 111, No. 977

The American Naturalist January-February 1977

RESOURCE-LIMITED GROWTH, COMPETITION, AND PREDATION: A MODEL AND EXPERIMENTAL STUDIES WITH BACTERIA AND BACTERIOPHAGE

BRUCE R. LEVIN, FRANK M. STEWART, AND LIN CHAO

Zoology Department, University of Massachusetts, Amherst, Massachusetts 01002; and Mathematics Department, Brown University, Providence, Rhode Island 02912 3. One Resource, One Prey, One Predator

With one population at each trophic level the equilibrium conditions are:

$$\hat{r} + (\hat{n} + \hat{m})\phi(\hat{r})/\rho = C, \qquad (9)$$

$$\phi(\hat{r})|e - \gamma \hat{p} = \rho, \qquad (10)$$

$$\hat{m} = \gamma (1 - e^{-\rho t}) \hat{n} \hat{p} / \rho, \qquad (11)$$

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Idea:

Phage = Predators, Bacteria = Prey

Attributed to:

Allan Campbell 1961 (Evolution):

"<u>The simple predator.</u> If a virulent phage and a susceptible bacterium are mixed in an open growth system, such as a chemostat..."

CONDITIONS FOR THE EXISTENCE OF BACTERIOPHAGE¹

ALLAN CAMPBELL Department of Biology, University of Rochester

Received June 21, 1960

"One Resource, One Prey, One Predator"

3. One Resource, One Prey, One Predator

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The Lotka-Volterra model is the basis for models of **viral-host** population dynamics

Dynamic model

$$\frac{dR}{dt} = -\omega(R - R_0) - \gamma RN$$
$$\frac{dN}{dt} = \epsilon \gamma RN - \phi NV - \omega N$$
$$\frac{dV}{dt} = \beta \phi NV - \phi NV - \omega V$$

Interactions:

Resource inflow/outflow Host growth and outflow Viral lysis and outflow

(note: original model included time delays)

Similar model proposed by Campbell (1961) Evolution 15: 153 & adapted to phage-bacteria chemostats by Levin et al. (1977) Am. Nat. 111:3



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Again, counter-clockwise cycles













Counter-clockwise cycles have appeared robust given alternative viral-host models

I. Models with an infected class

$$\begin{aligned} \frac{dN}{dt} &= rN\left(1 - \frac{N+I}{K}\right) - \phi NV - \omega N\\ \frac{dI}{dt} &= \phi NV - \eta I - \omega I\\ \frac{dV}{dt} &= \beta \eta I - \phi NV - \omega V \end{aligned}$$



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2. Models with an explicit delay between infection and lysis

$$\frac{dN}{dt} = rN\left(1 - \frac{N+I}{K}\right) - \phi NV - \omega N$$
$$\frac{dI}{dt} = \phi NV - \frac{\phi N_{\tau}V_{\tau}e^{-\omega\tau}}{-\omega I} - \omega I$$
$$\frac{dV}{dt} = \beta \phi N_{\tau}V_{\tau}e^{-\omega\tau} - \phi NV - \omega V$$



Counter-clockwise cycles have appeared robust given alternative viral-host models



Mathematical Check-Point:

 10^{7}

What life history traits enable viral invasion and persistence with their microbial hosts?

The same types of cycles can be observed in virus-host population dynamics

"Lotka-Volterra" like cycles between T4 and *E. coli* B

Data: Bohannan & Lenski, Ecology (1997)



Summary of Part 1

Take-home message:

Original models of viral-host dynamics presuppose a "simple" one virus, one host relationship.

In these models, viruses act like a predator, leading to cyclical dynamics in which viral peaks follow host peaks (leading to counter-clockwise cycles).

Invasion and persistence depends on both life history traits and environmental conditions - it is not inevitable.

However...

Evolution can rapidly change the number/relative abundance of viruses and hosts strains...

