## Lecture 2 <br> Dynamical models for single neurons. Measuring experimental neural data. Ana Amador

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## The Hodgkin - Huxley model

Using pioneering experimental techniques of that time, Hodgkin and Huxley (1952) determined not only the equations but measured all the parameters values

$$
\begin{aligned}
C \dot{V} & =I-\overbrace{\bar{g}_{\mathrm{K}} n^{4}\left(V-E_{\mathrm{K}}\right)}^{I_{\mathrm{K}}}-\overbrace{\bar{g}_{\mathrm{Na}} m^{3} h\left(V-E_{\mathrm{Na}}\right)}^{I_{\mathrm{Na}}}-\overbrace{g_{\mathrm{L}}\left(V-E_{\mathrm{L}}\right)}^{I_{\mathrm{L}}} \\
\dot{n} & =\alpha_{n}(V)(1-n)-\beta_{n}(V) n \\
\dot{m} & =\alpha_{m}(V)(1-m)-\beta_{m}(V) m \\
\dot{h} & =\alpha_{h}(V)(1-h)-\beta_{h}(V) h
\end{aligned}
$$

Values of shifted Nernst equilibrium potentials (so that $V_{\text {rest }}=0$ ) :
$E_{\mathrm{K}}=-12 \mathrm{mV}, \quad E_{\mathrm{Na}}=120 \mathrm{mV}, \quad E_{\mathrm{L}}=10.6 \mathrm{mV} ;$
Values of maximal conductances:
$\bar{g}_{\mathrm{K}}=36 \mathrm{mS} / \mathrm{cm}^{2}, \quad \bar{g}_{\mathrm{Na}}=120 \mathrm{mS} / \mathrm{cm}^{2}$,

$$
g_{\mathrm{L}}=0.3 \mathrm{mS} / \mathrm{cm}^{2} .
$$

$$
\begin{aligned}
& \alpha_{n}(V)=0.01 \frac{10-V}{\exp \left(\frac{10-V}{10}\right)-1} \\
& \beta_{n}(V)=0.125 \exp \left(\frac{-V}{80}\right) \\
& \alpha_{m}(V)=0.1 \frac{25-V}{\exp \left(\frac{25-V}{10}\right)-1} \\
& \beta_{m}(V)=4 \exp \left(\frac{-V}{18}\right) \\
& \alpha_{h}(V)=0.07 \exp \left(\frac{-V}{20}\right) \\
& \beta_{h}(V)=\frac{1}{\exp \left(\frac{30-V}{10}\right)+1} .
\end{aligned}
$$

Value of membrane capacitance:
4-dimensional system: difficult to study the dynamics
$C=1 \mu \mathrm{~F} / \mathrm{cm}^{2}$

## The Hodgkin - Huxley model, simplified

Many interesting features of single neuron dynamics can be illustrated using two-dimensional systems

$$
\begin{aligned}
& C \dot{V}=I-\overbrace{g_{\mathrm{L}}\left(V-E_{\mathrm{L}}\right)}^{\text {leak } I_{\mathrm{L}}}-\overbrace{g_{\mathrm{Na}} m_{\infty}(V)\left(V-E_{\mathrm{Na}}\right)}^{\text {instantaneous } I_{\mathrm{Na}, \mathrm{p}}}-\overbrace{g_{\mathrm{K}(n)\left(V-E_{\mathrm{K}}\right)}}^{I_{\mathrm{K}}}, \\
& \dot{n}=\left(n_{\infty}(V)-n\right) / \tau(V), \\
& \text { Definition: } x \text {-Nullcline: } \\
& \text { curve such that } \frac{d x}{d t}=\dot{x}=0
\end{aligned}
$$

The $V$-nullcline is given by the equation $I-g_{\mathrm{L}}\left(V-E_{\mathrm{L}}\right)-g_{\mathrm{Na}} m_{\infty}(V)\left(V-E_{\mathrm{Na}}\right)-g_{\mathrm{K}} n\left(V-E_{\mathrm{K}}\right)=0$
which has the solution

$$
n=\frac{I-g_{\mathrm{L}}\left(V-E_{\mathrm{L}}\right)-g_{\mathrm{Na}} m_{\infty}(V)\left(V-E_{\mathrm{Na}}\right)}{g_{\mathrm{K}}\left(V-E_{\mathrm{K}}\right)}
$$

The equation $n_{\infty}(V)-n=0 \quad$ defines the $n$-nullcline $n=n_{\infty}(V)$

## The Hodgkin - Huxley model, simplified

Nullclines:

$$
n=\frac{I-g_{\mathrm{L}}\left(V-E_{\mathrm{L}}\right)-g_{\mathrm{Na}} m_{\infty}(V)\left(V-E_{\mathrm{Na}}\right)}{g_{\mathrm{K}}\left(V-E_{\mathrm{K}}\right)}
$$

$$
n=n_{\infty}(V)
$$



## FitzHugh-Nagumo model

The system

$$
\begin{aligned}
\dot{V} & =V(a-V)(V-1)-w+I, \\
\dot{w} & =b V-c w,
\end{aligned}
$$

imitates generation of action potentials by Hodgkin-Huxley-type models having cubic (N-shaped) nullclines (similar to the Figure in the previous slide)

- Parameter $I$ mimics the injected current. For the sake of simplicity, we set $I=0$
- Parameter $a$ describes the shape of the cubic parabola $V(a-V)(V-1)$

The nullclines of the FitzHugh-Nagumo are:

$$
\begin{array}{ll}
w=V(a-V)(V-1)+I & \\
w=b / c V & \\
w \text {-nullcline }) \\
(w \text {-nullcline })
\end{array}
$$

## FitzHugh-Nagumo model

The nullclines of the FitzHugh-Nagumo are:

$$
\begin{array}{ll}
w=V(a-V)(V-1)+I & (V \text {-nullcline }) \\
w=b / c V & (w \text {-nullcline })
\end{array}
$$

Spike and bi-stability


## Recording neural activity

Now that we have a model for spikes, we want to record real spikes!

Or is it the other way around?

We first make recording of spikes and propose to use dynamical models of spikes to make sense of our recordings
(we are just measuring few spikes out of many, many more!)

## Measuring electrical activity in the brain



## Measuring electrical activity in the brain


es NeuroNexus
Micromachined silicon probes


Value: around USD 1000



## Measuring electrical activity in the brain

In-house manufactured tetrodes


- Diameter 0.0005" (12.7 $\mu \mathrm{m}$ )
- Tungsten, HML coating
- Impedances: 500 k to 3 MOhm



## Measuring electrical activity in the brain

- ultra-lightweight - customizable
- compact - low-cost

Connector


- Tetrodes manufactured in-house
- Diameter 0.0005" (12.7 $\mu \mathrm{m}$ )
- Tungsten, HML coating
- Impedances:

500k to 3 MOhm

## Lightweight recording device

- ultra-lightweight
- compact
customizable
- low-cost

Connector


- Pitch 90 TPI = $282 \mu \mathrm{~m} / \mathrm{turn}$
(manual step $\approx 25 \mu \mathrm{~m}$ )
- Mass <1g

- 15 mm tall, $13 \mathrm{~mm}^{2}$ footprint
- Tetrode geometry easy to modify.


- Tetrodes manufactured in-house
- Diameter 0.0005" (12.7 $\mu \mathrm{m}$ )
- Tungsten, HML coating
- Impedances:

500k to 3 MOhm
$\approx 100 \mu \mathrm{~m}$

- 4-tetrode array (16ch) + Ref
- $2 x 2$ geometry


## Neural recordings

Raw data

Low-pass filter

High-pass filter


## Neural recordings

Multi-unit recordings


## Neural recordings



Different neurons may present different spike shapes


## Neural recordings



Spike sorting: PCA analysis

## Neural recordings

## Neural activity in response to the Bird's Own Song (BOS)





## Neural recordings: LFP and MUA

MUA (Multi-Unit Activity)



LFP
(Local Field Potential)

## Neural recordings: LFP and MUA

MUA

|  |
| :--- | :--- | :--- |



## Neural recordings: LFP and MUA



Rhythms in the Brain?

## Rhythms in the Brain

## nature <br> neuroscience

## Speech perception exhibits rhythmicity

When presented with an acoustic stimulus
(speech or modulated white noise) the auditory cortex tracks the amplitude modulation of the input.

## The speech rhythm enhances perception

Spontaneous synchronization to speech reveals neural mechanisms facilitating language learning
M. Florencia Assaneo ${ }^{\text {© }}$ 1,7*, Pablo Ripollés ${ }^{1,7}$, Joan Orpella ${ }^{2,3,4,7}$, Wy Ming Lin ${ }^{1}$,

Ruth de Diego-Balaguer ${ }^{2,3,4,5,8}$ and David Poeppel ${ }^{1,6,8}$

 b

$\times 10^{-3}$


## Rhythms in the Speech

## Speech production exhibits rhythmicity



Speech rhythms and their neural foundations

David Poeppel $\mathbb{C}^{1,2 凶}$ and M. Florencia Assaneo $\odot^{2,3}$

## Rhythms in the Brain of a Songbird?

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nature
neuroscience
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ARTICLES https://doi.org/10.1038/s41593-019-0353-z

Spontaneous synchronization to speech reveals neural mechanisms facilitating language learning
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a
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 Q IWH Whithandman b


## Rhythms in the Brain of a Songbird !



LFP oscillations are phase-locked (synchronization)

Spontaneous synchronization to speech reveals neural mechanisms facilitating language learning
M. Florencia Assaneo ${ }^{\text {© }}{ }^{1,7 \star}$, Pablo Ripollés ${ }^{17,7}$, Joan Orpella ${ }^{2,3,4,7}$, Wy Ming Lin ${ }^{1}$, Ruth de Diego-Balaguer ${ }^{2,3,4,5,8}$ and David Poeppel ${ }^{1,6,8}$
a

 b


## Rhythms in the Brain of a Songbird !



LFP oscillations are phase-locked (synchronization)




## LFP and coding of behavior (birdsong)



LFP oscillations occur at the song syllabic rate


## LFP vs. MUA vs. SUA

## Extracellular

 recordings in canaries while hearing song (BOS)

## LFP vs. MUA vs. SUA

## Extracellular recordings in canaries while hearing song (BOS)




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## Collaborators

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