

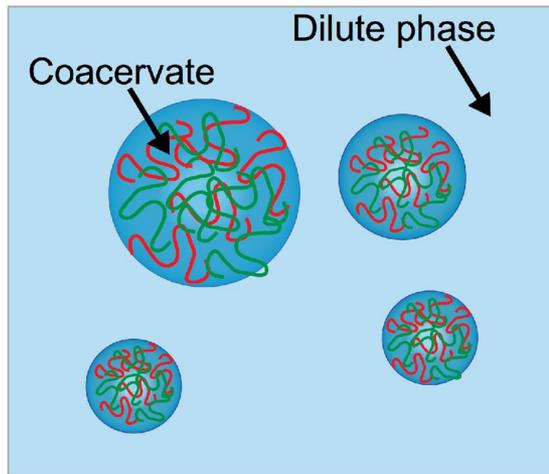
# Liquid Liquid Phase Separation and Fibrillization of Intrinsically Disordered Peptides



Joan Shea, Department of Chemistry, UC Santa Barbara

# Proteins can assemble in different ways

**“Liquid”**

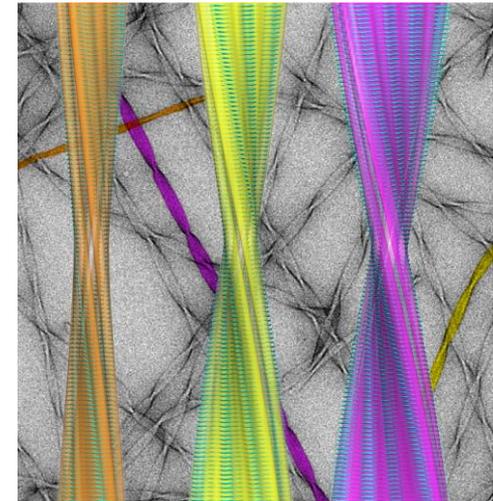


Droplets  
Biomolecular condensates  
Coacervates

**“Solid”**

Aging

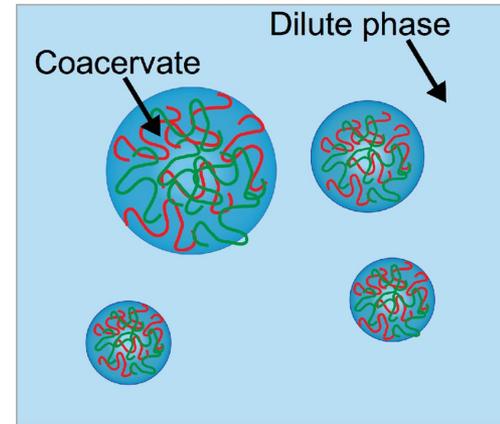
A black arrow pointing from the liquid phase diagram to the solid phase image.



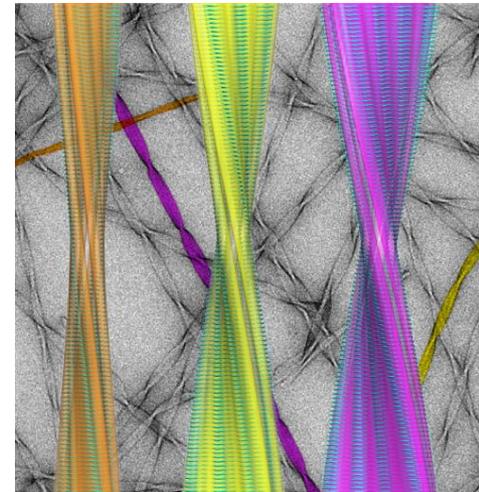
Amyloid Fibrils

# Outline

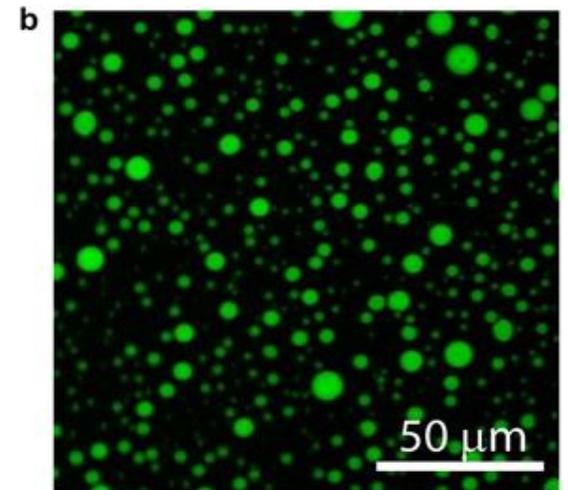
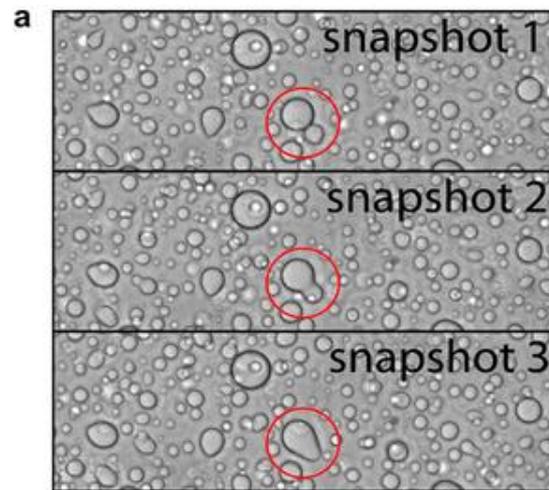
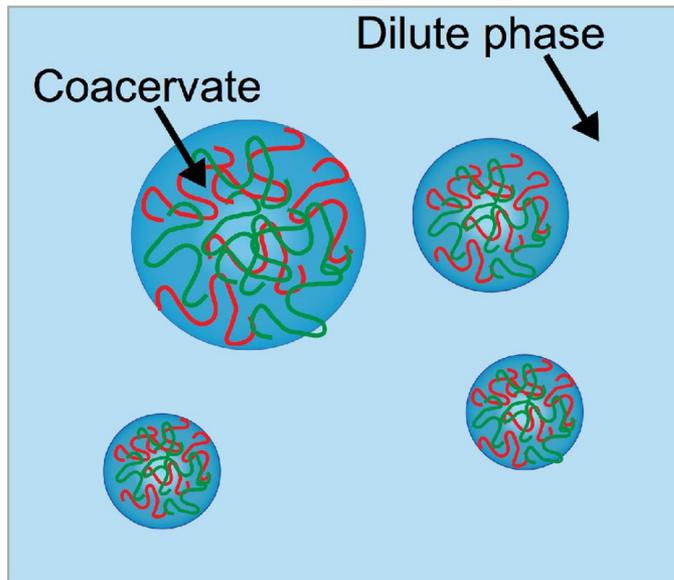
## Part 1: Liquid-Liquid Phase Separation: From Model Systems to Tau



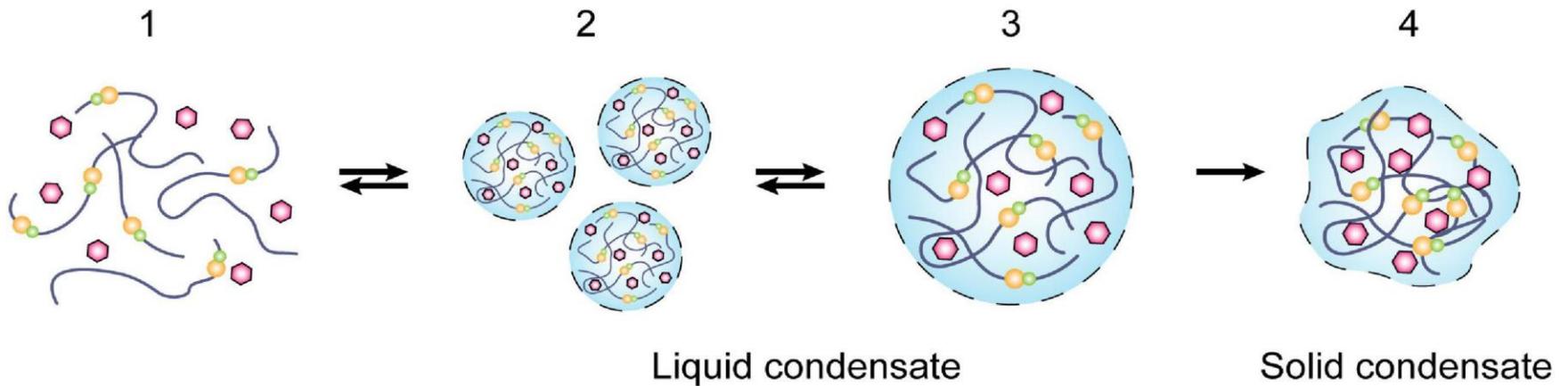
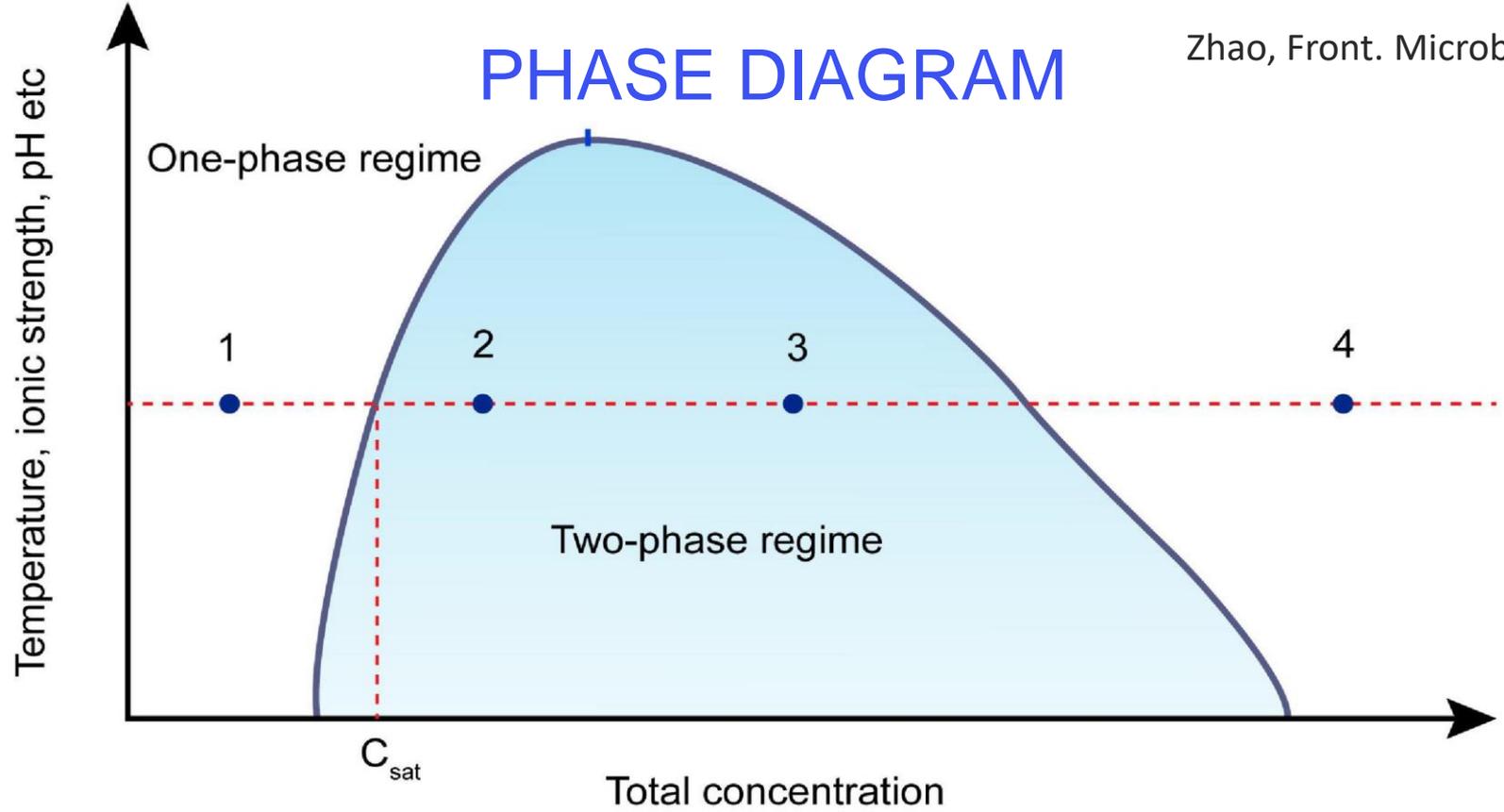
## Part 2: Aggregation of the Tau Protein



Coacervation = liquid liquid phase separation  
= formation of droplets  
= formation of biomolecular condensates



# PHASE DIAGRAM



# What drives phase separation?

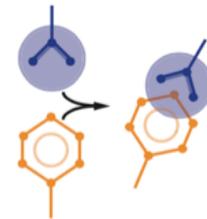
Charge pattern



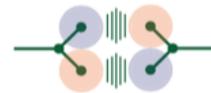
Chain charge density



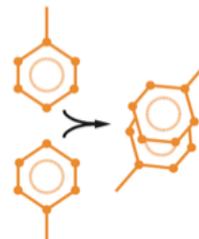
Short-ranged interactions



cation -  $\pi$



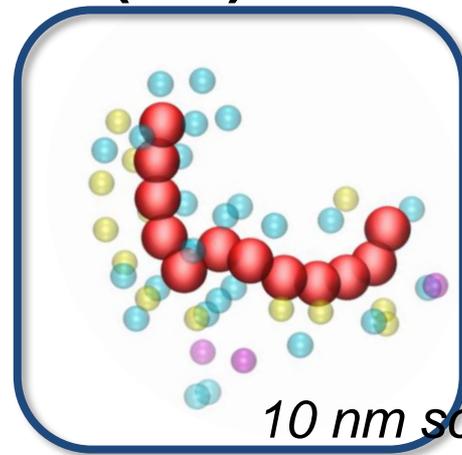
dipole - dipole



$\pi$  -  $\pi$  stacking

# Computational Approaches

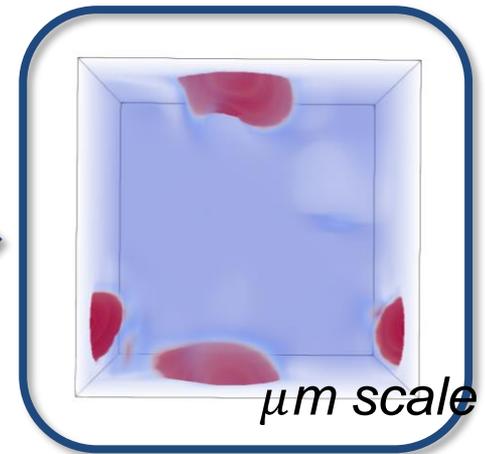
**Coarse-grained  
(CG) MD**



*10 nm scale*



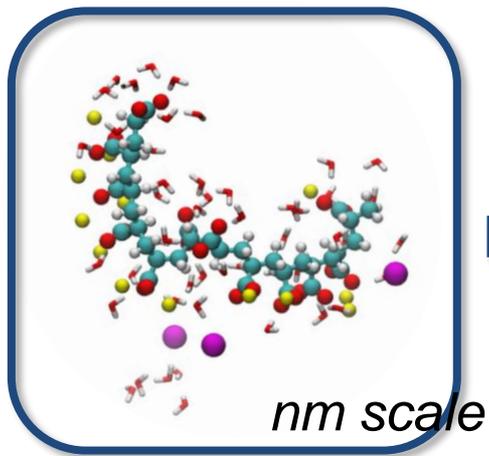
**Field theoretic  
simulations  
(FTS)**



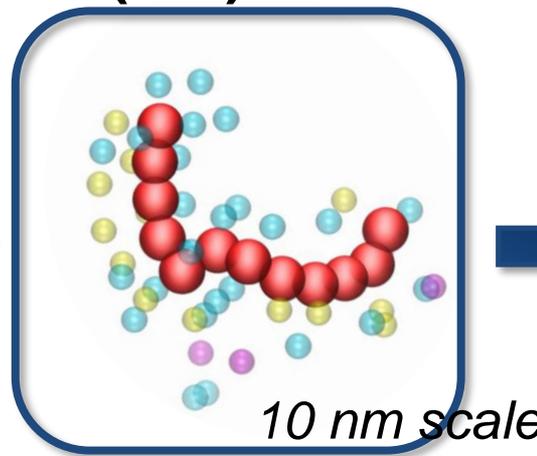
*$\mu\text{m}$  scale*

# Computational Approach

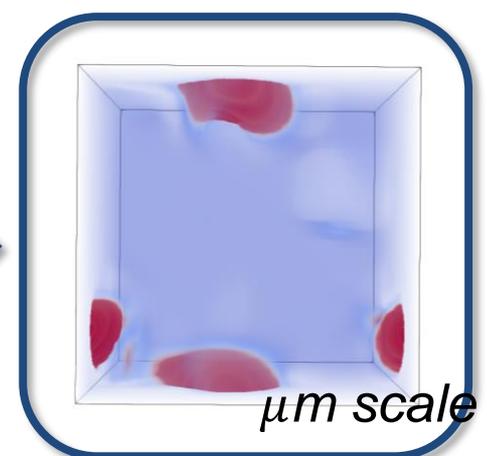
All-atom (AA) MD



Coarse-grained (CG) MD



Field theoretic simulations (FTS)



 **chemistry**

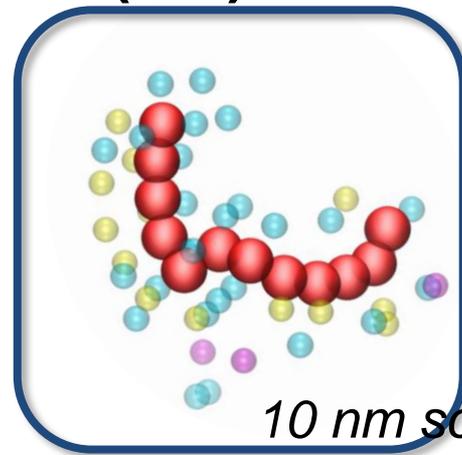
**macroscopic properties** 

- Clear connection to chemical details
- Short time/length scale
- Dilute conditions

- Can probe long time and length scales
- Efficient for high concentrations

# Computational Approaches

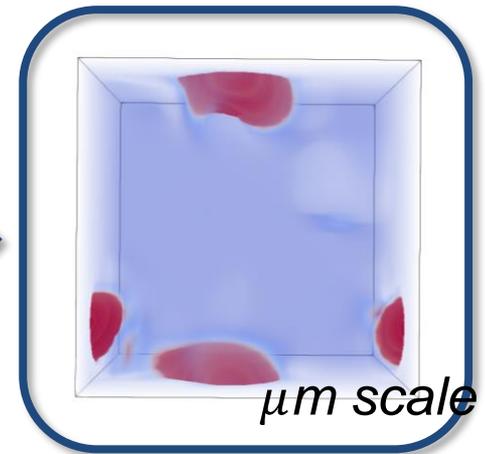
**Coarse-grained  
(CG) MD**



*10 nm scale*



**Field theoretic  
simulations  
(FTS)**



*$\mu\text{m}$  scale*

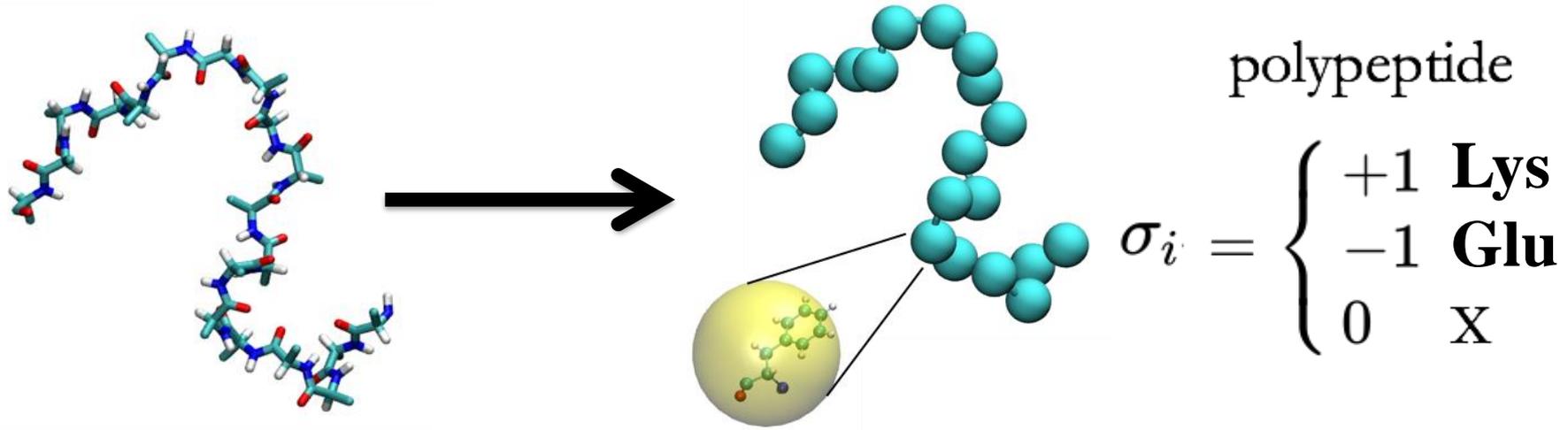




# Coarse-Grained Model

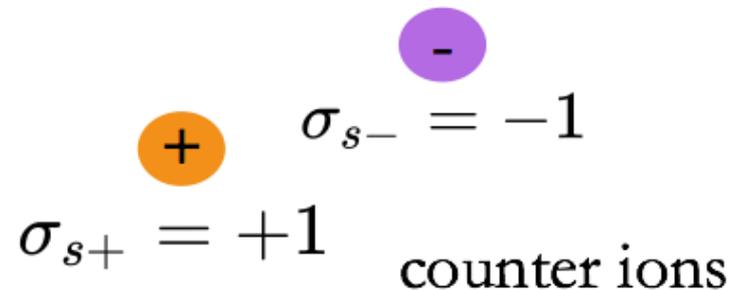
## Coarse-grained peptide chain

Each amino acid represented by a single site

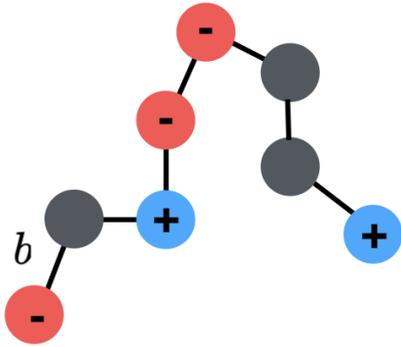


**Implicit solvent**

**Explicit salt ions**



# Discrete Gaussian chain polyelectrolyte model



## charge sequence

$$\{\sigma_i\} = \{\sigma_1, \sigma_2, \dots, \sigma_N\}$$

## charge density

$$\frac{1}{N} \sum_i |\sigma_i|$$

$N_l$  sites per molecule

$\sigma_i$  charge on bead  $i$

$b$  segment length

# Formally equivalent representations



Particle MD simulation



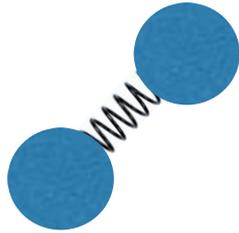
Field Theoretic simulation

low polymer density

high polymer density

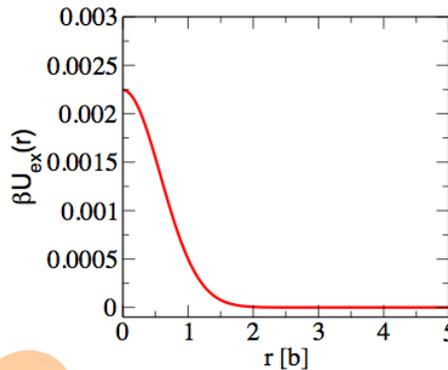
bond potential

$$\beta U_b = \frac{3}{2b^2} r^2$$



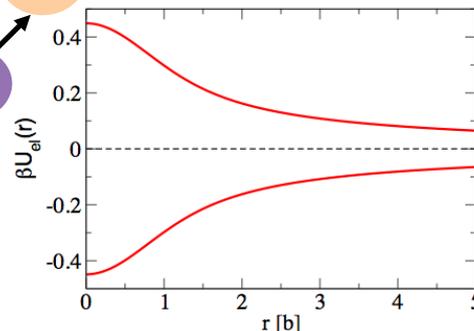
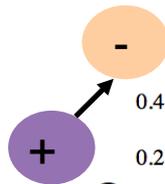
excluded volume parameter

$$\beta U_{ev}(r) = \frac{v}{8\pi^{3/2} a^3} e^{-r^2/4a^2}$$



electrostatic potential

$$\beta U_{es} = \frac{l_B z_i z_j}{r} \operatorname{erf}\left(\frac{r}{2a}\right)$$

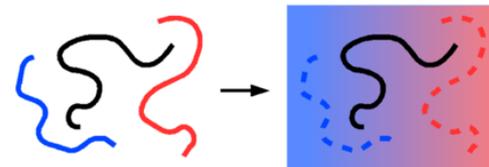


$$H[w, \psi] = \frac{1}{2B} \int d\mathbf{r} w(\mathbf{r})^2 + \frac{1}{2E} \int d\mathbf{r} |\nabla\psi|^2 - \sum_l \frac{CV\bar{\phi}_l}{N_l} \ln Q_l$$

Fluctuating chemical potential field

Fluctuating electrostatic potential field

Single chain partition function for polymers and salt ions



Polymers decoupled!

# Formally equivalent representations



Particle MD  
simulation



Field Theoretic  
simulation

low polymer density

high polymer density

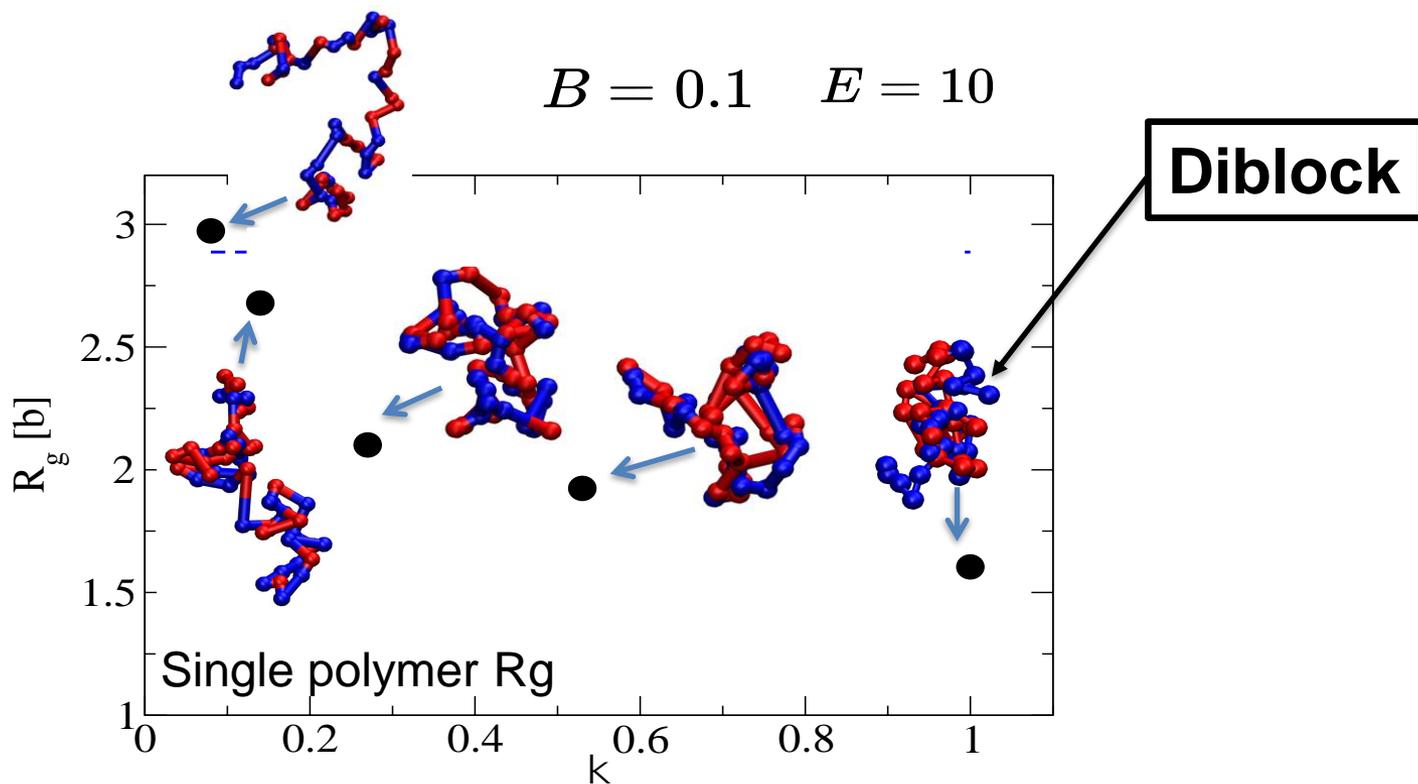
			SCD	$\kappa$
sv10			-2.10	0.08
sv15			-4.35	0.14
sv20			-7.37	0.27
sv25			-12.77	0.53
sv30			-27.84	1.00

# Particle Based Simulations

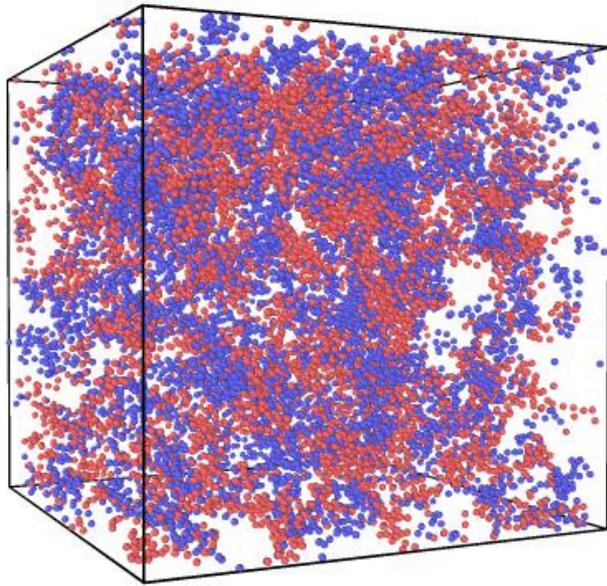
		SCD	$\kappa$
sv10		-2.10	0.08
sv15		-4.35	0.14
sv20		-7.37	0.27
sv25		-12.77	0.53
sv30		-27.84	1.00



James Mc Carthy

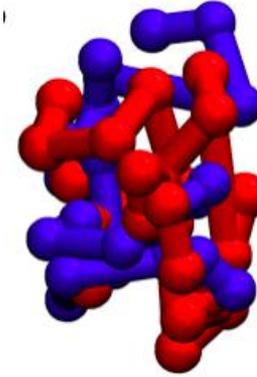


# Particle Based Simulations

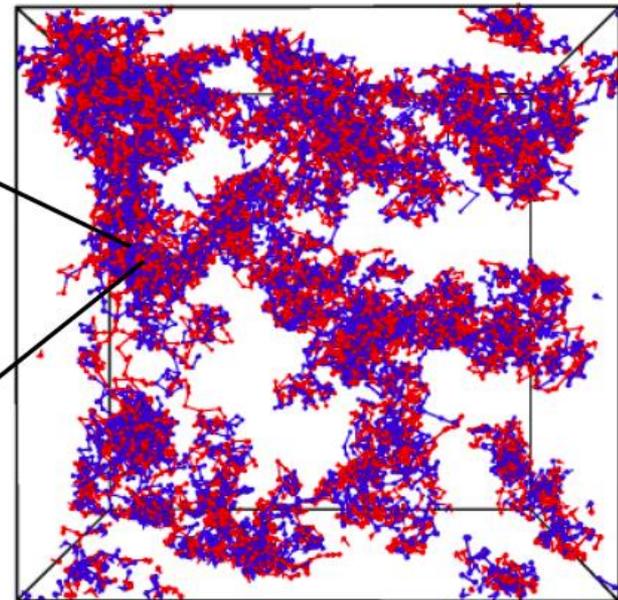
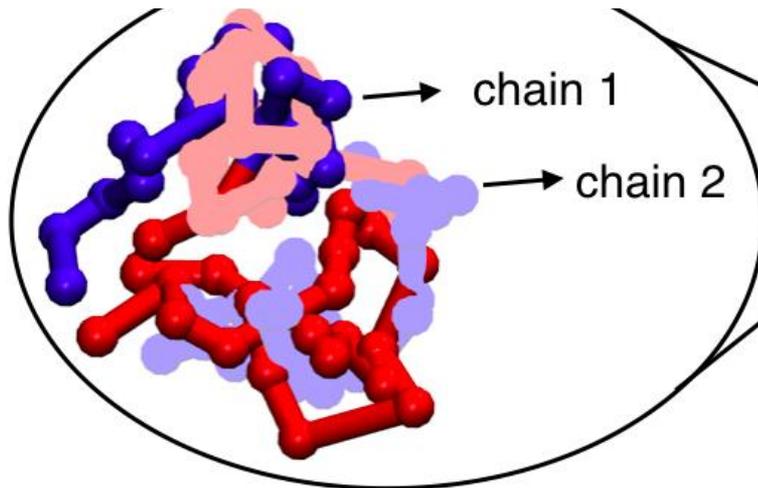


$$n = 300$$

$$N = 50$$



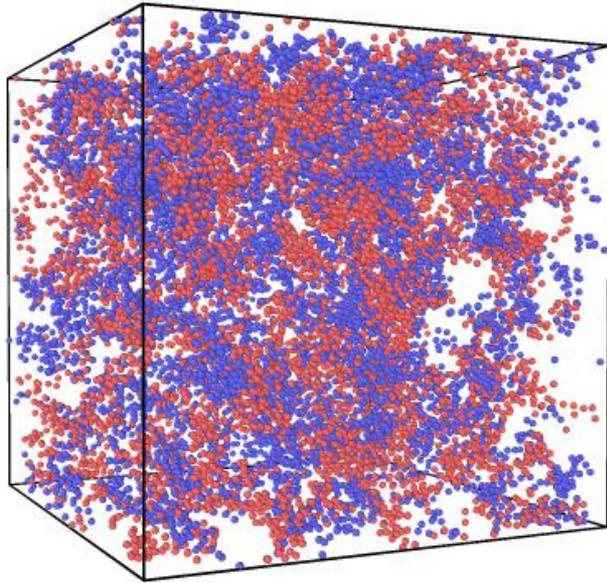
Single Chain



# Particle Based versus Field Theoretic Simulations

$n = 300$

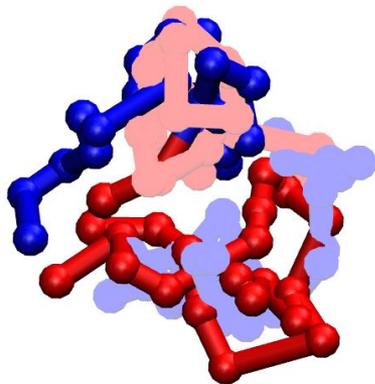
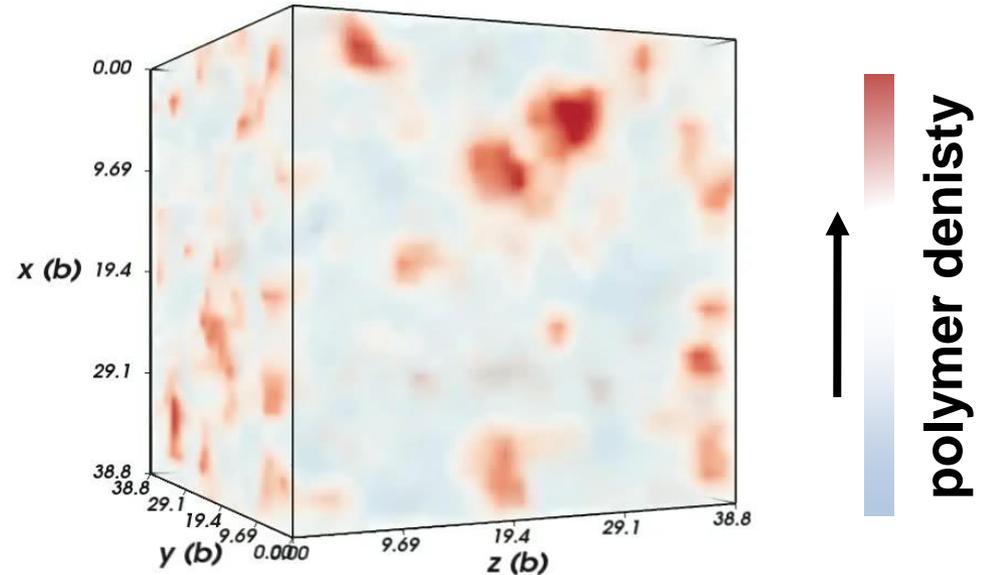
$N = 50$



$C = 0.015$

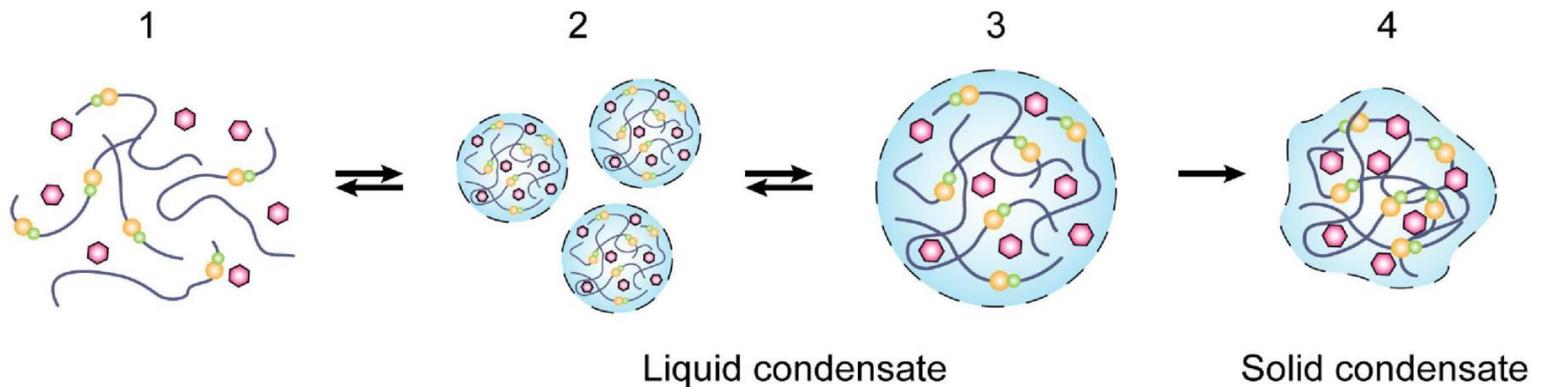
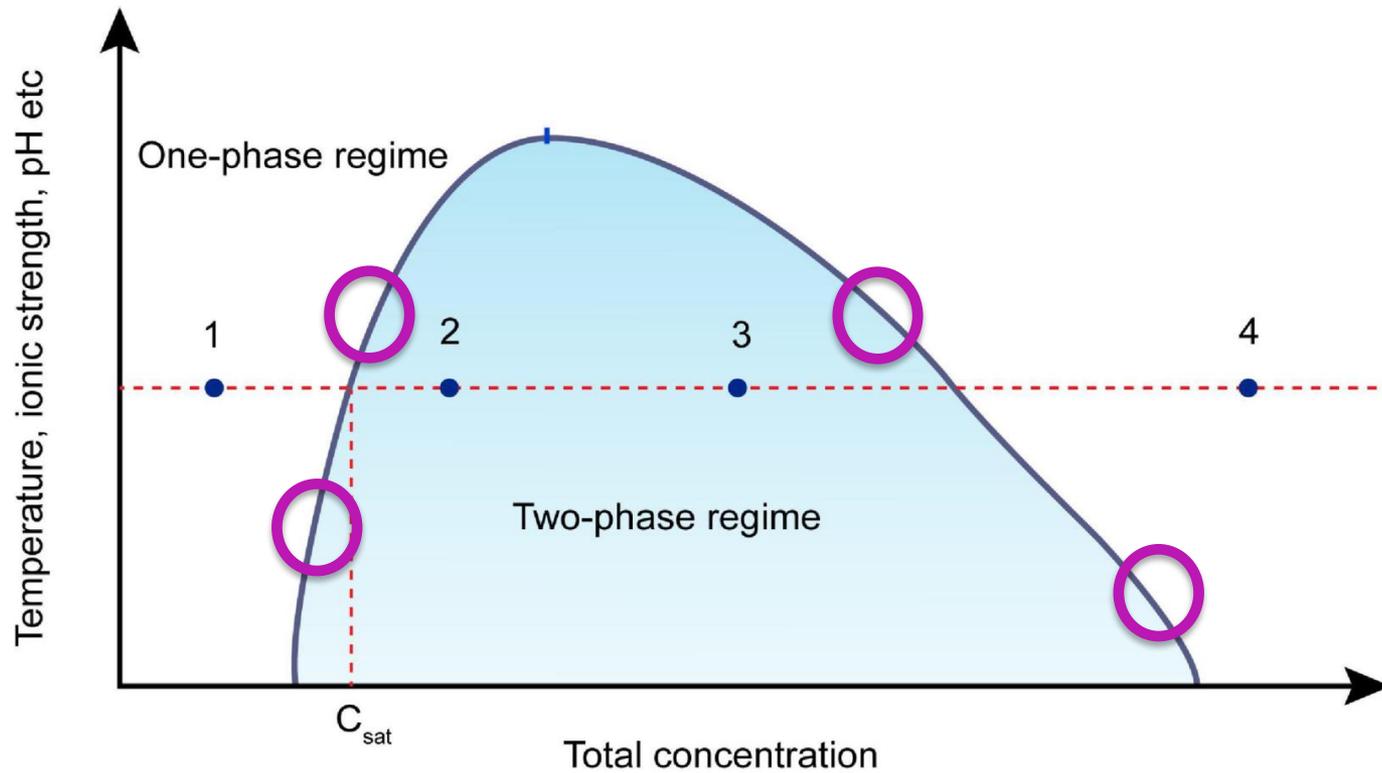


$B = 0.1$



Get a Phase Diagram

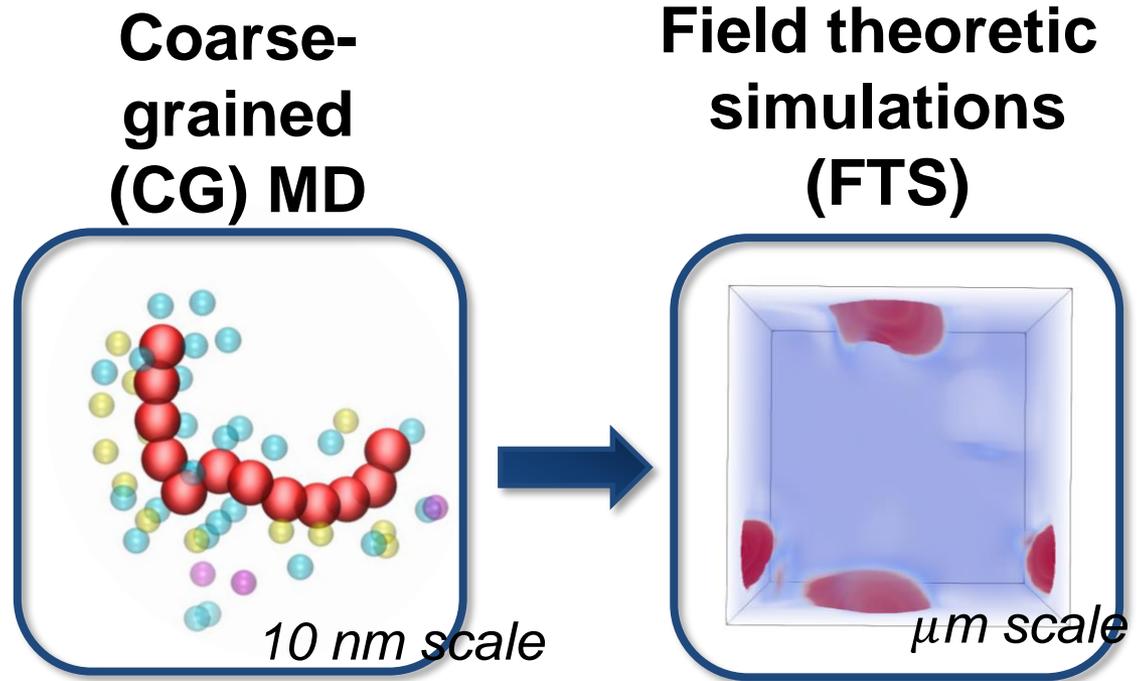
# We map the boundaries of the phase diagram







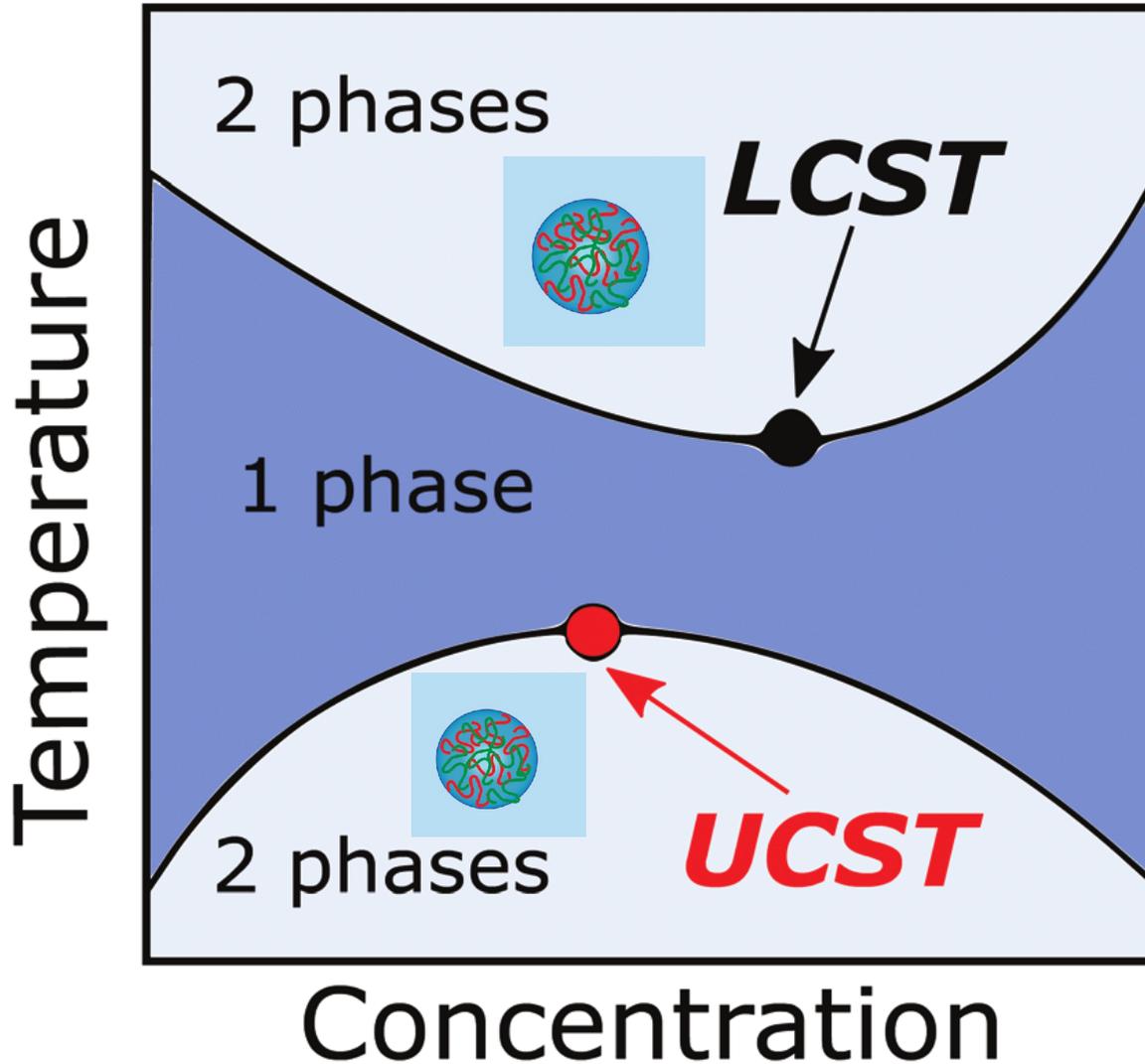
# Computational Approach



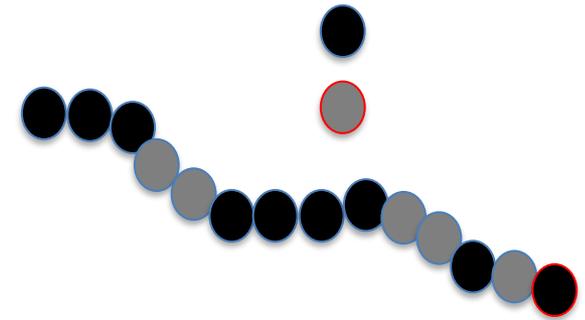
## Limitations

- No amino acid specificity
- Implicit solvent

# Upper (UCST) and Lower (LCST) critical solution temperature

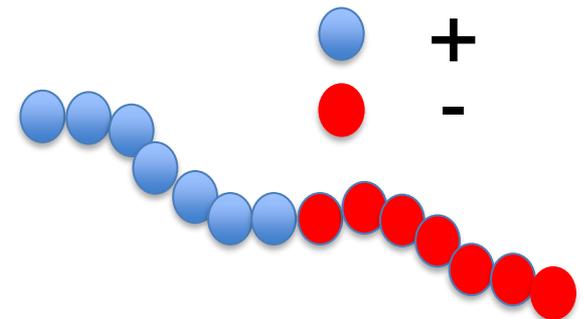


Hydrophobic amino acids

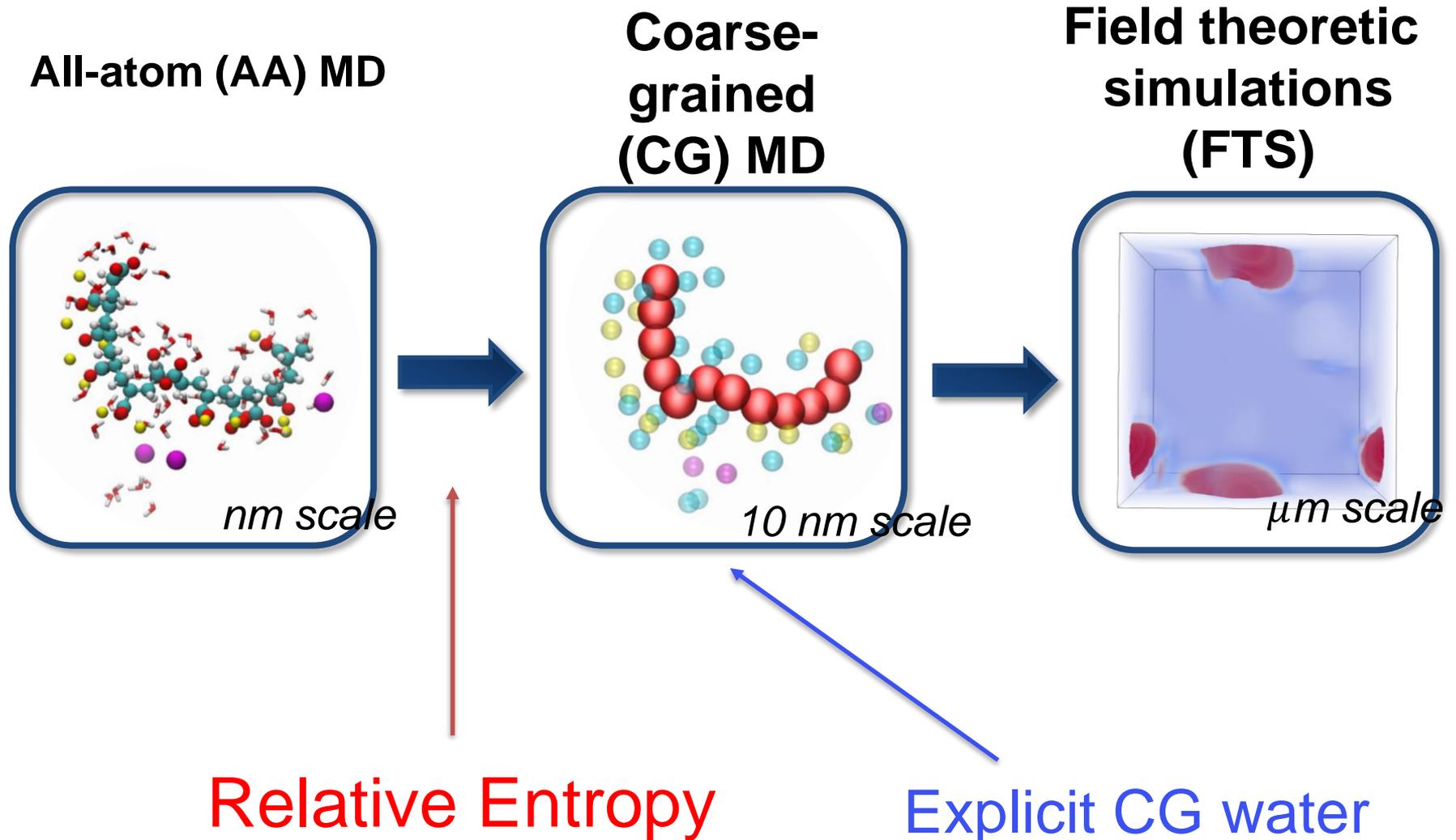


Release of water

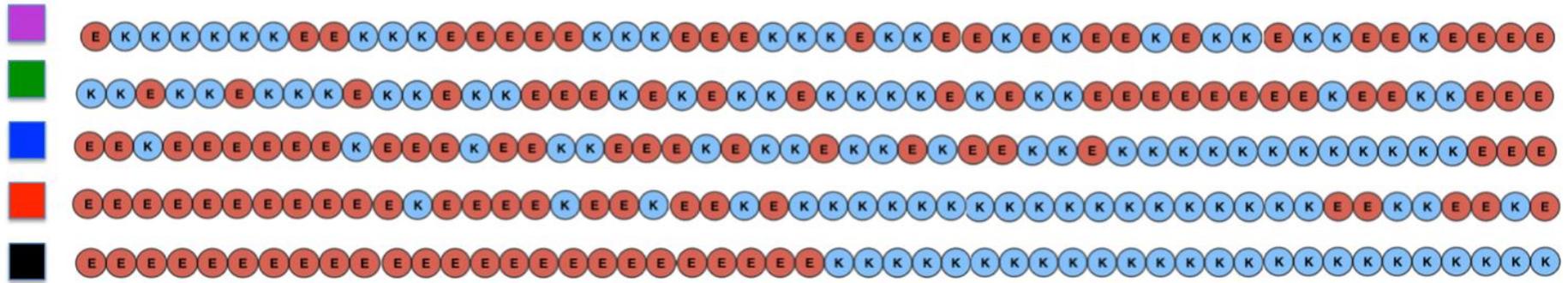
Charged amino acids



# Computational Approach



OLD: Model System: **KE** (Lys/Glu)sequence



**NEW**: Model System: **RE** (Arg/Glu)sequence

RE1: Poly-Arg/Glu 

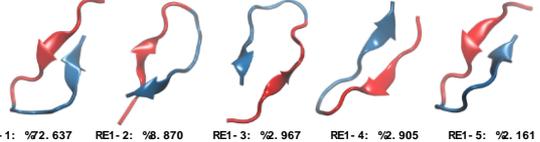
RE2: Poly-Arg/Glu 

RE3: Poly-Arg/Glu 

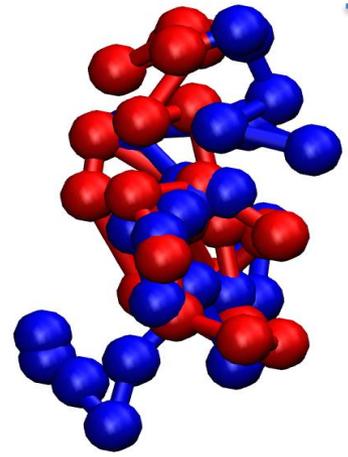
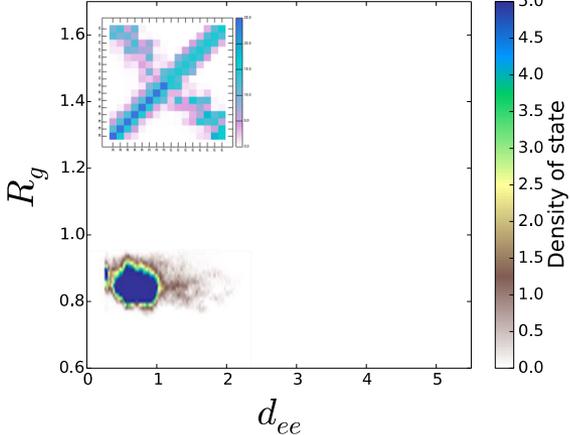
RE4: Poly-Arg/Glu 

# All atom replica exchange MD reference simulations

a) RE1: Poly-Arg/Glu ●●●●●●●● ●●●●●●●●

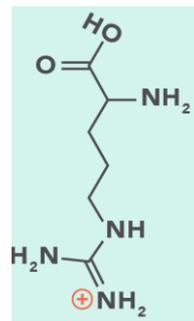


RE1-1: %2.637 RE1-2: %8.870 RE1-3: %2.967 RE1-4: %2.905 RE1-5: %2.161

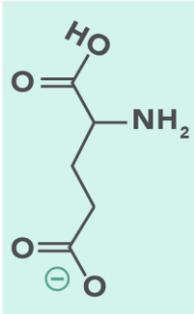


Original  
CG

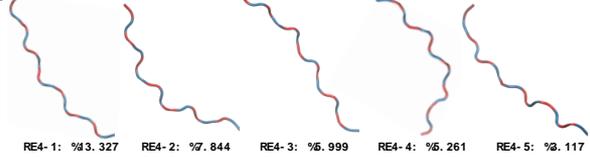
Arg



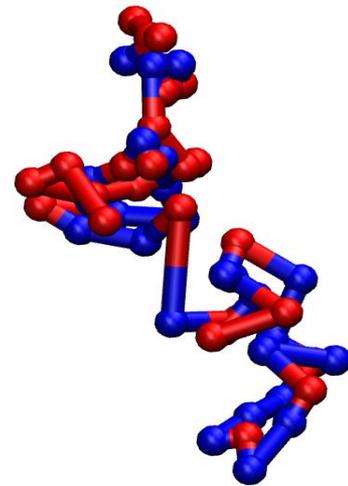
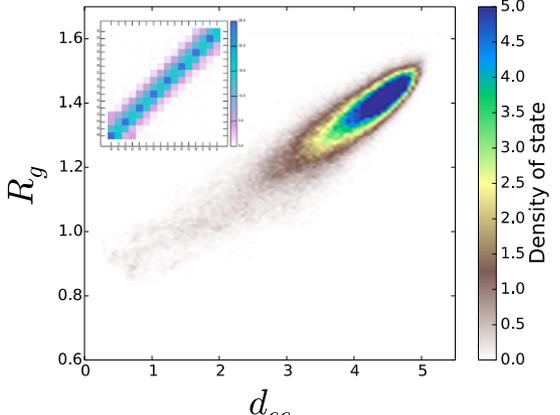
Glu



d) RE4: Poly-Arg/Glu ●●●●●●●● ●●●●●●●●



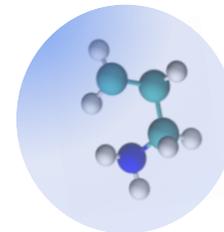
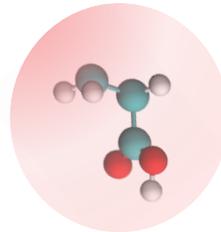
RE4-1: %3.327 RE4-2: %7.844 RE4-3: %6.999 RE4-4: %6.261 RE4-5: %8.117



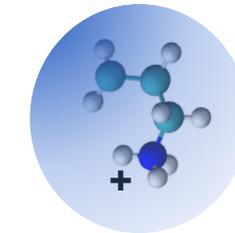
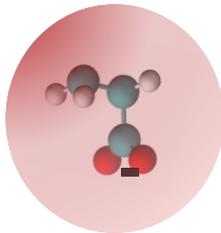
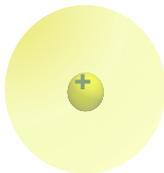
# Relative Entropy Coarse-graining

Minimizing the relative entropy:

$$S_{rel} = \int \int \rho_{AA}(\mathbf{r}) \ln \left( \frac{\rho_{AA}(\mathbf{r})}{\rho_{CG}(\mathbf{R})} \right) \delta(\mathbf{M}(\mathbf{r}) - \mathbf{R}) d\mathbf{r} d\mathbf{R}$$



Scott Shell

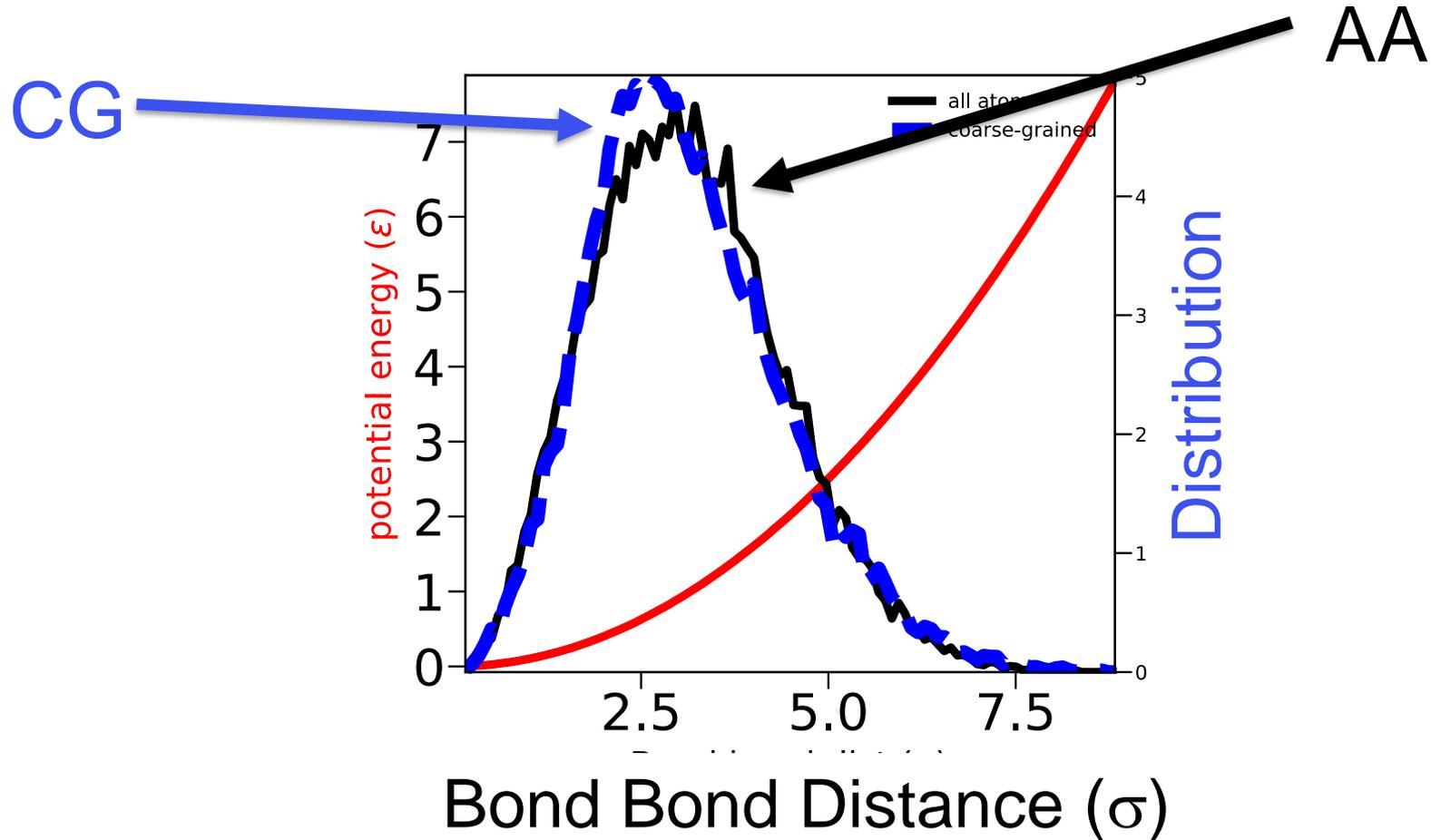


Water

ions

Amino acids

# Relative Entropy Parameterization



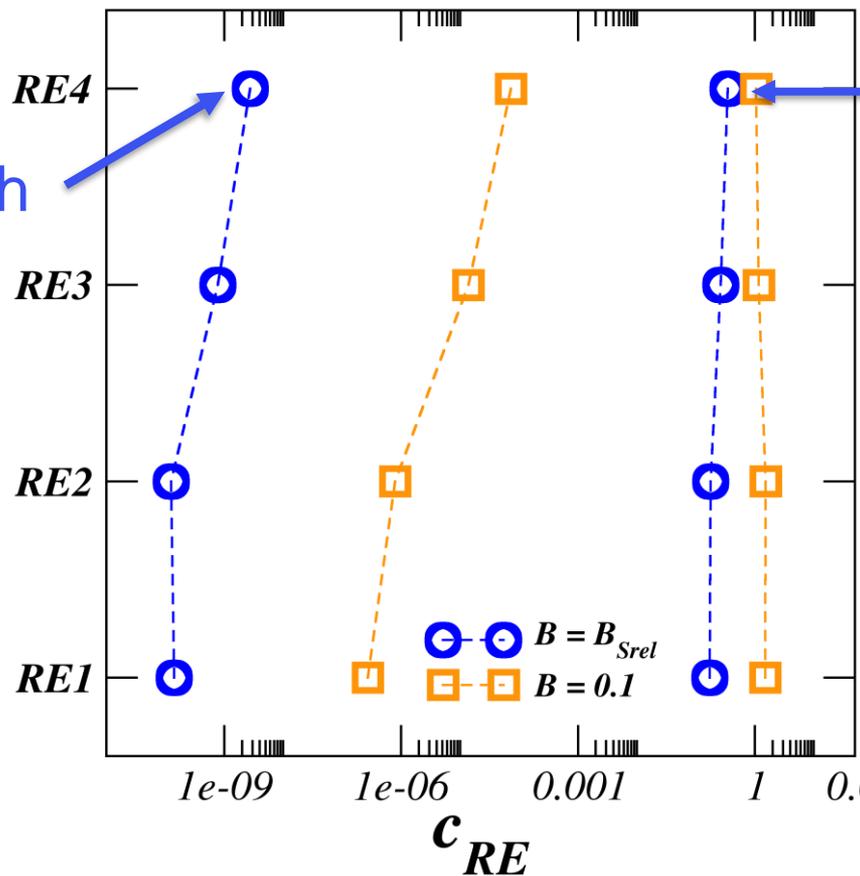
# RE peptides phase behavior

RE1: Poly-Arg/Glu ●●●●●●●● ●●●●●●●●

RE2: Poly-Arg/Glu ●●●●●●●● ●●●●●●●●

RE3: Poly-Arg/Glu ●●●●●●●● ●●●●●●●●

RE4: Poly-Arg/Glu ●●●●●●●● ●●●●●●●●



Dense Branch

Dilute Branch

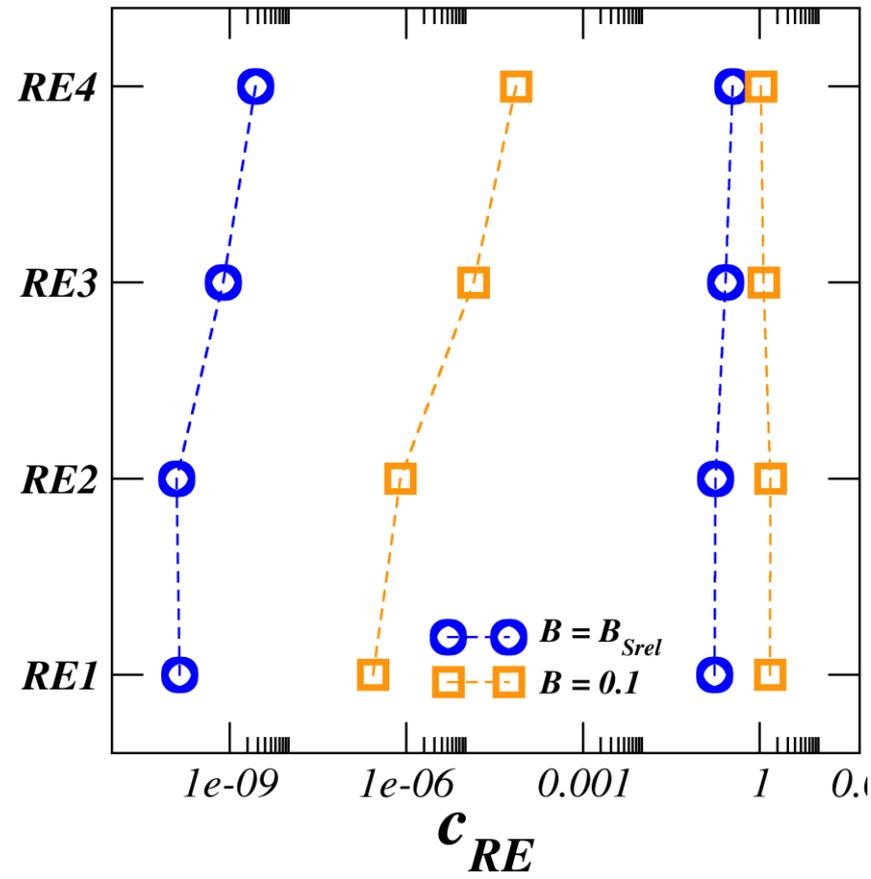
# RE peptides phase behavior

RE1: Poly-Arg/Glu ●●●●●●●● ●●●●●●●●

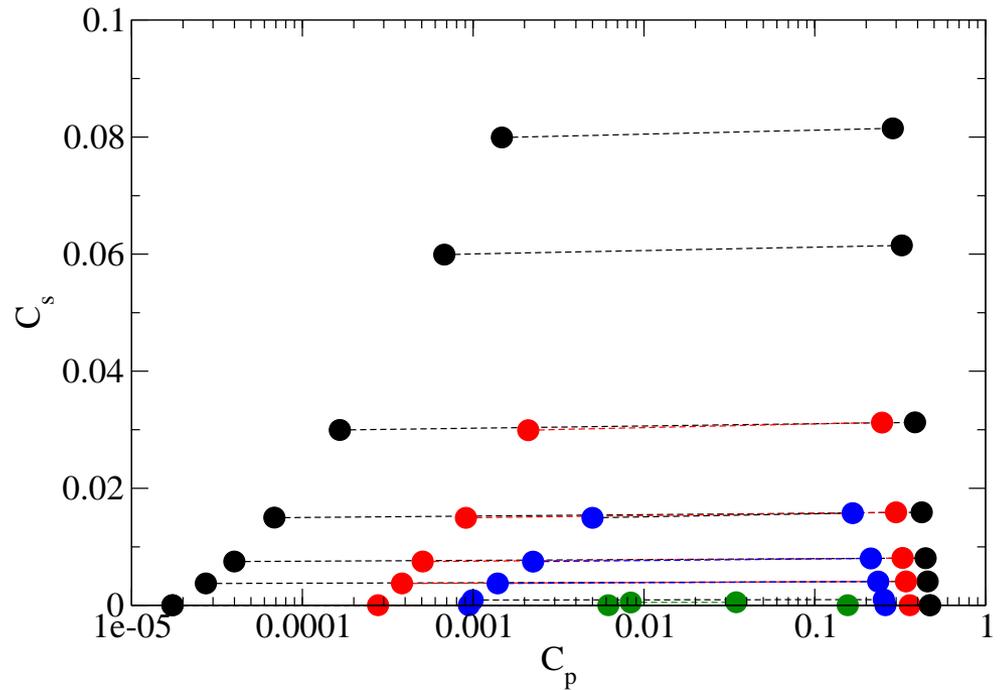
RE2: Poly-Arg/Glu ●●●●●●●● ●●●●●●●●

RE3: Poly-Arg/Glu ●●●●●●●● ●●●●●●●●

RE4: Poly-Arg/Glu ●●●●●●●● ●●●●●●●●



## “OLD” FTS

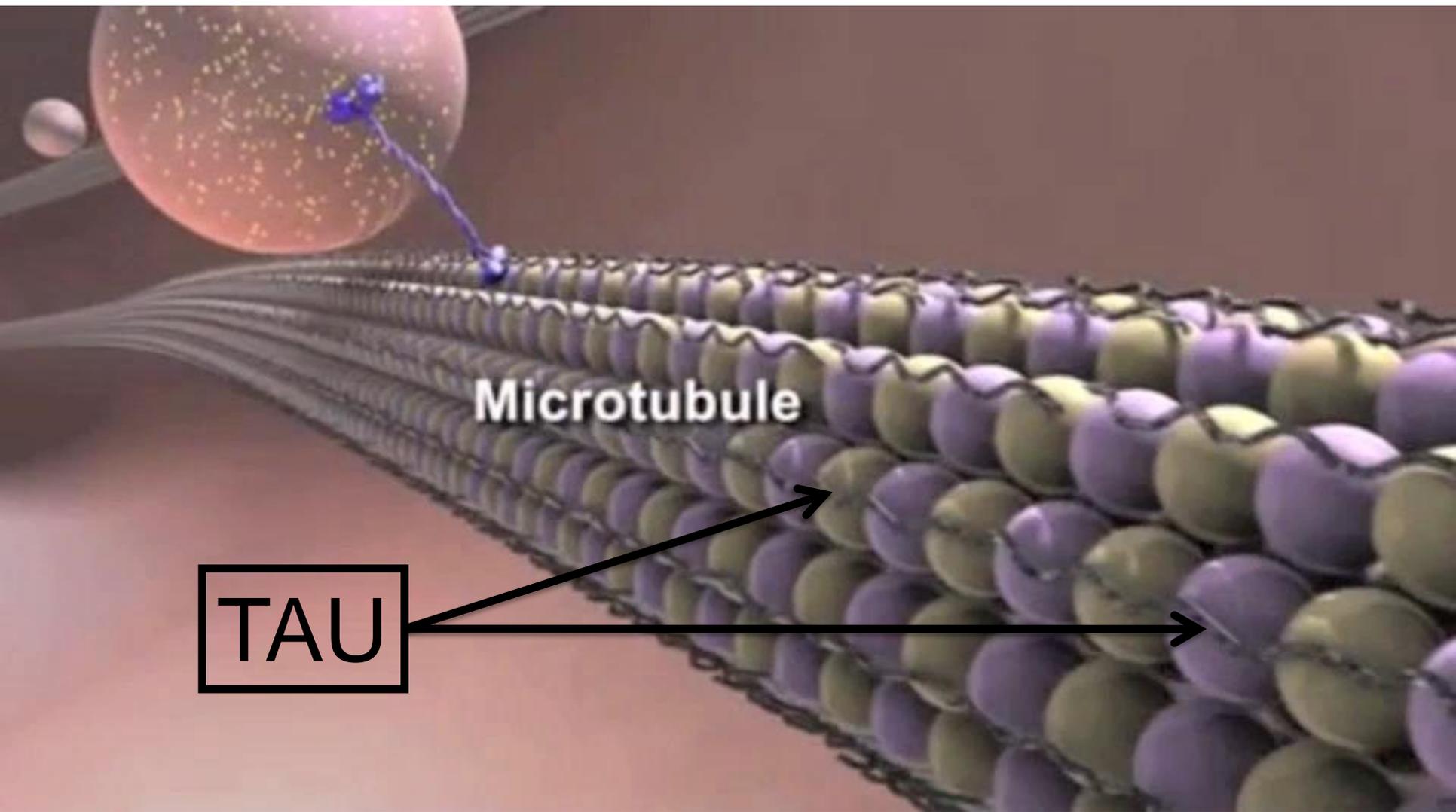


# Tau Protein

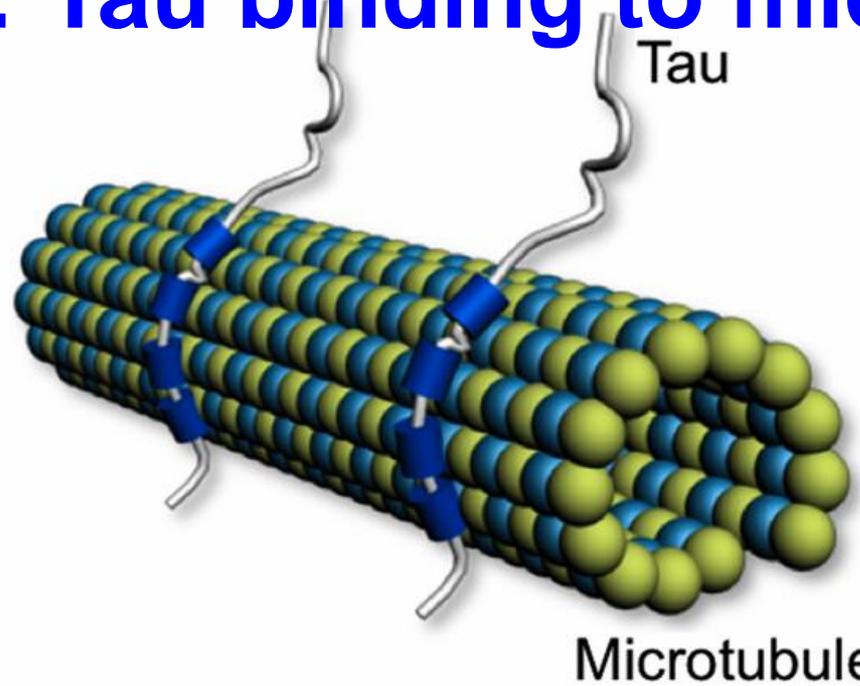
## Liquid Liquid Phase Separation and Fibrillization



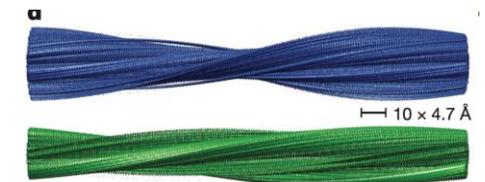
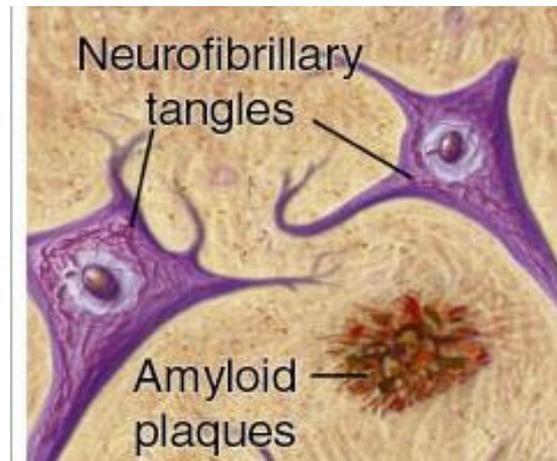
# MICROTUBULES ARE STABILIZED BY TAU PROTEINS



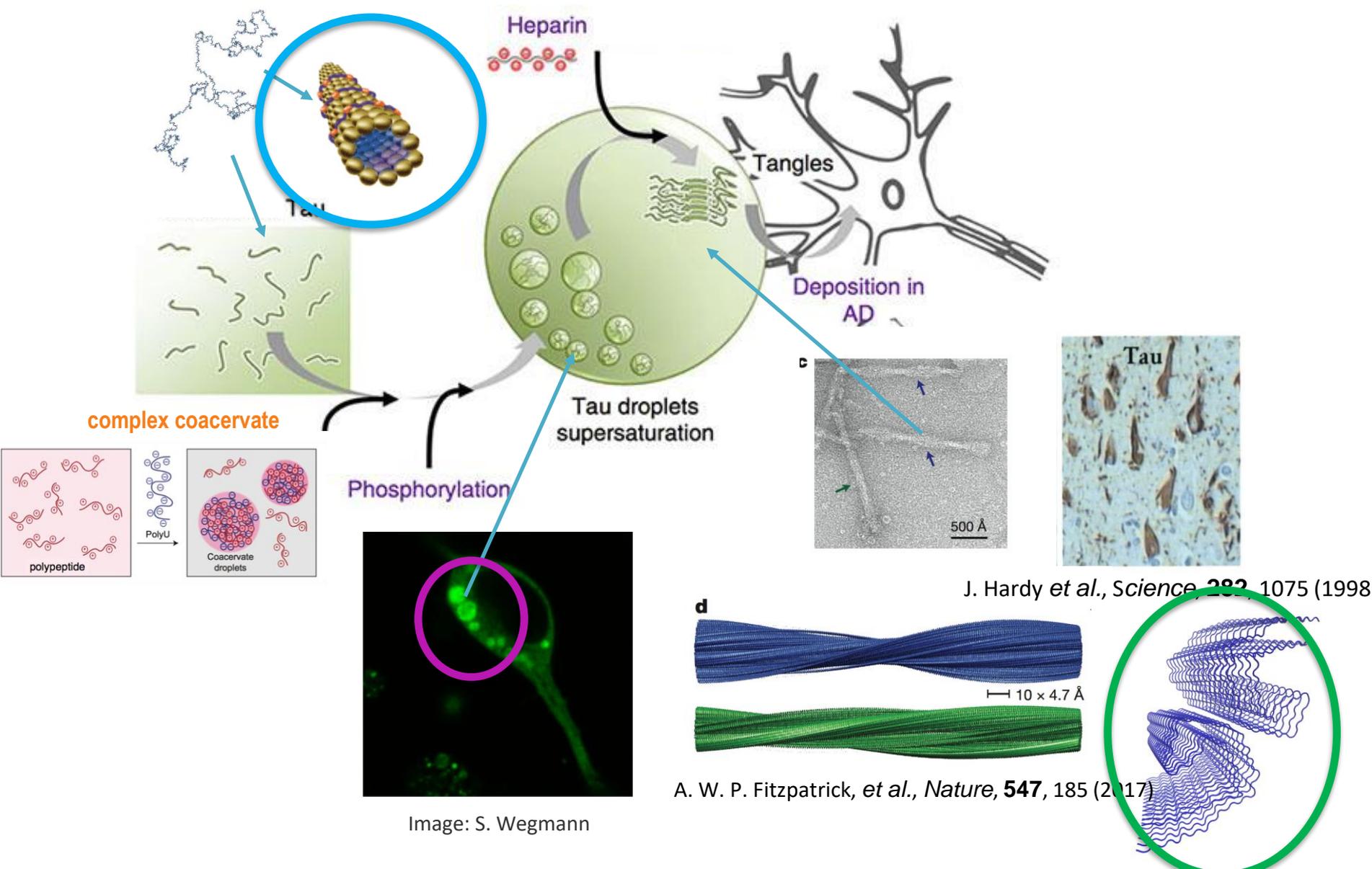
# Function: Tau binding to microtubule



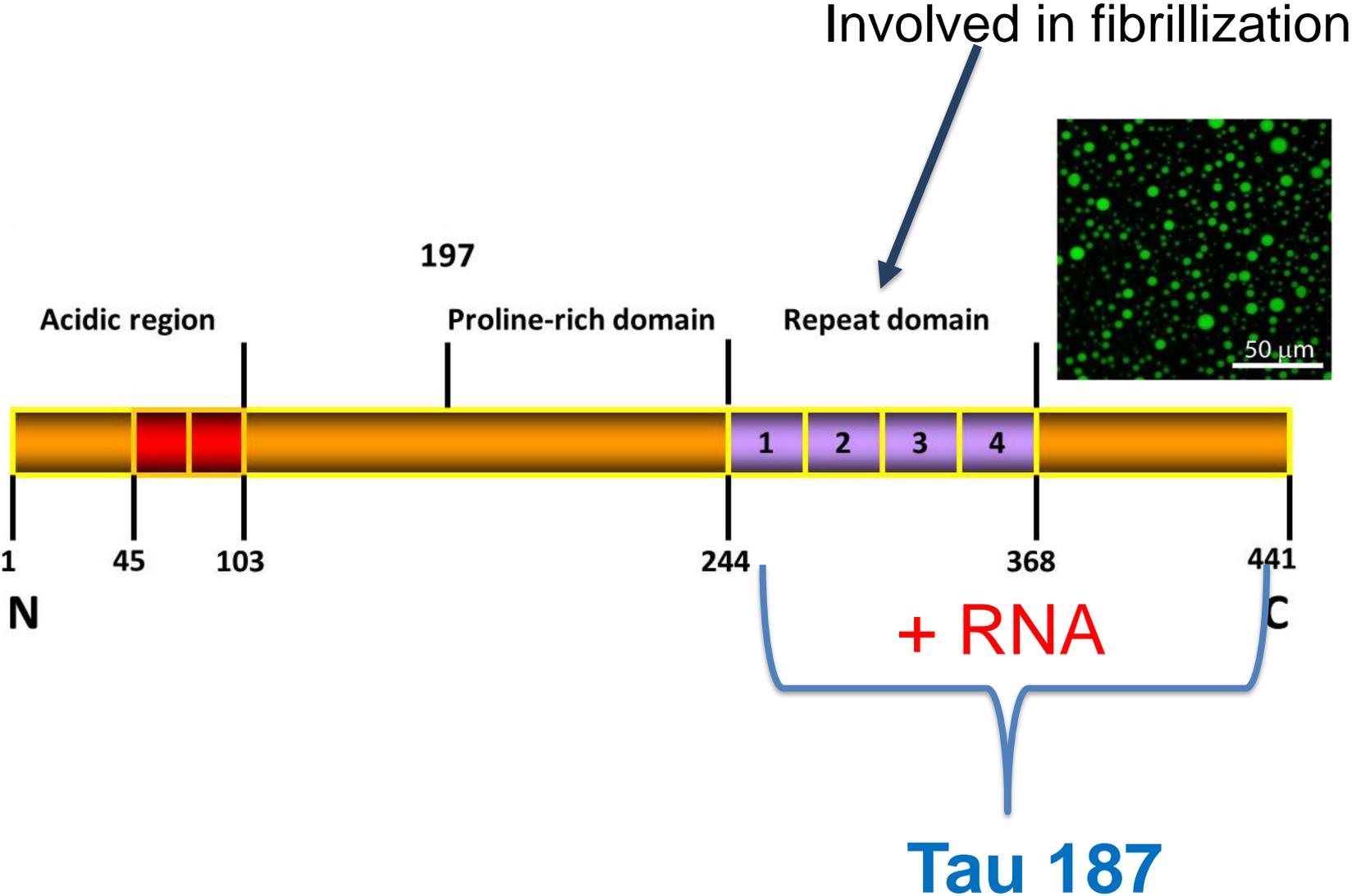
# Pathology: Tau and Aggregation



# Liquid-Liquid Phase Separation of Tau and Fibril Formation

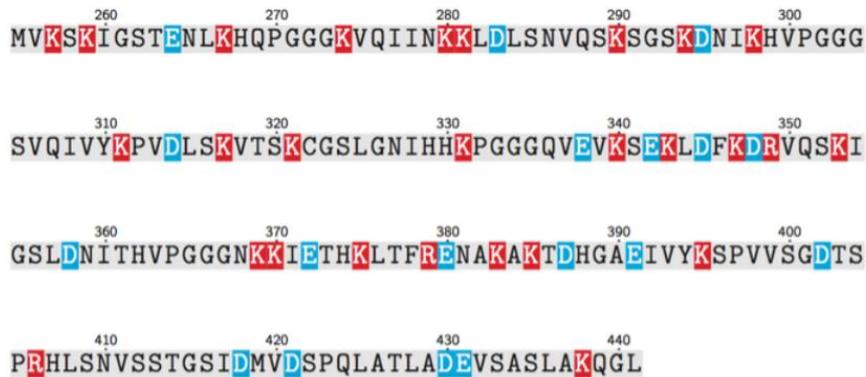


# Tau sequence

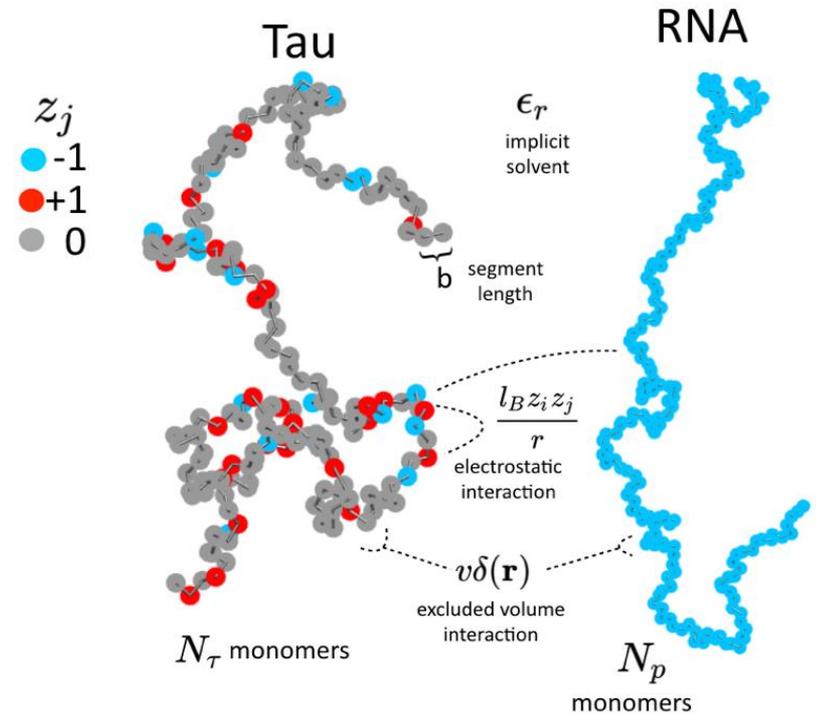


# Field Theory Modeling of Tau-RNA complex coacervation

Tau187 primary sequence

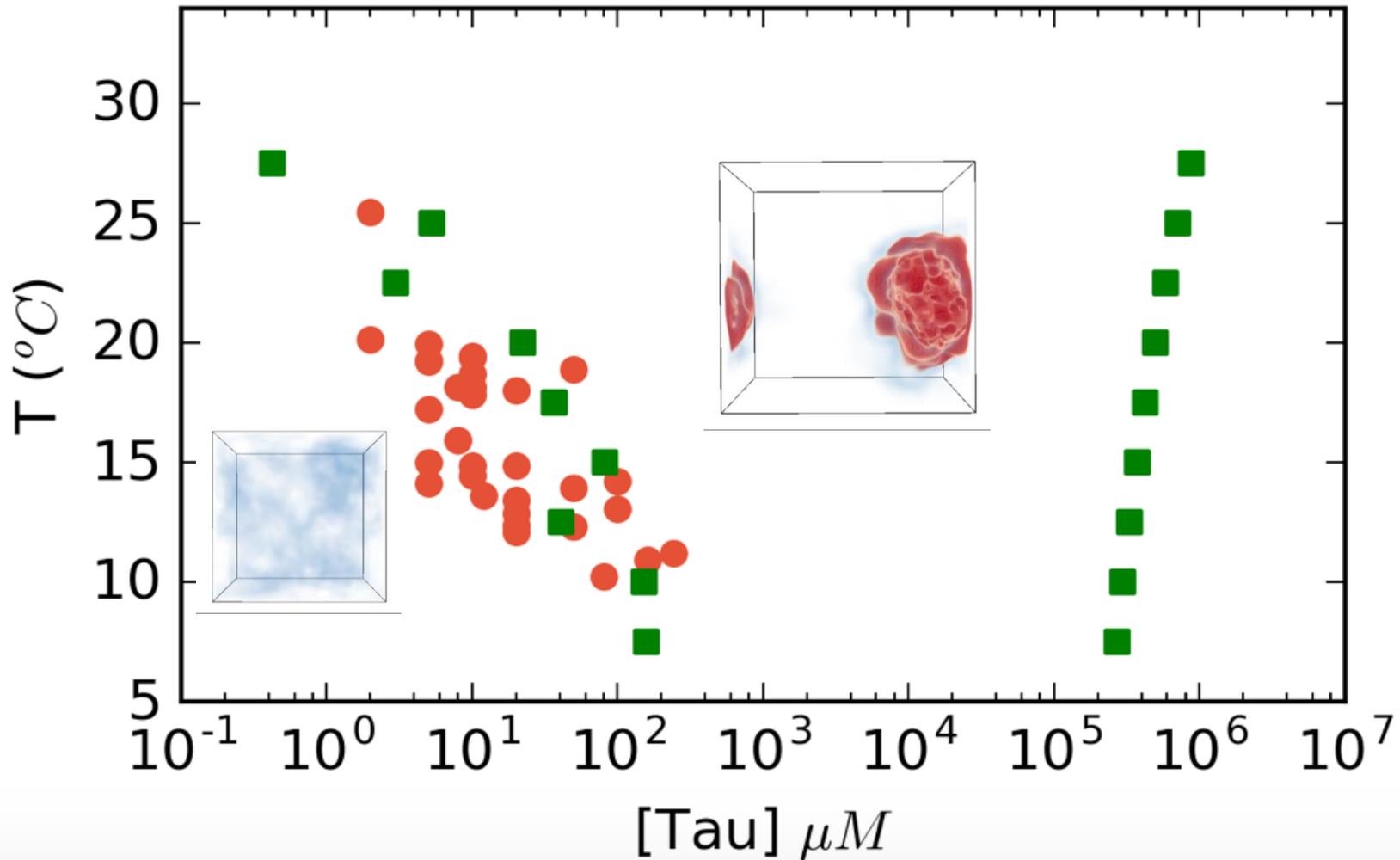


**X** basic (+)  
**X** acidic (-)  
**X** neutral (0)

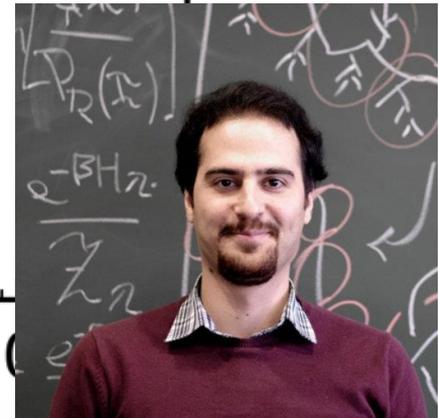
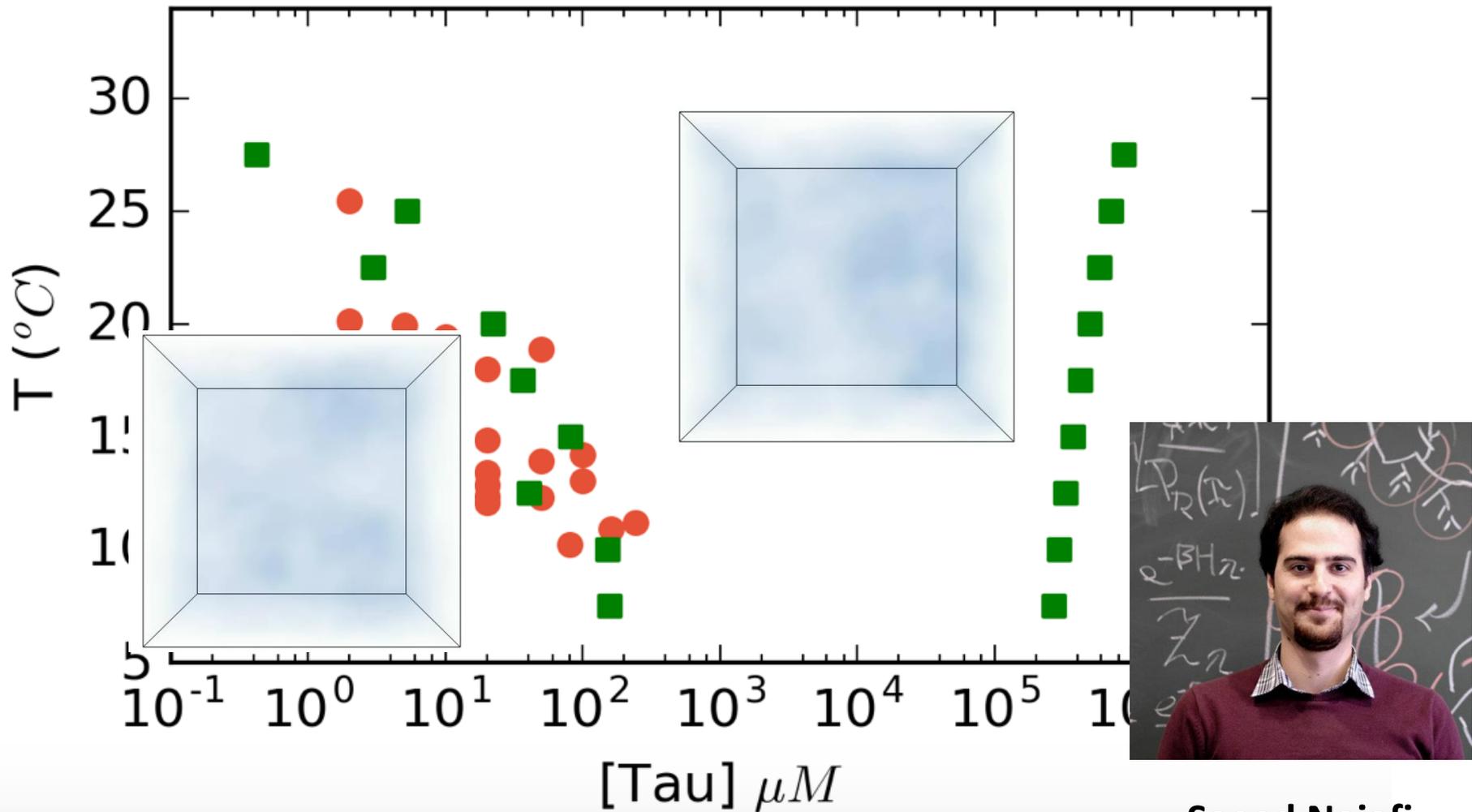


Fit excluded volume (B) and Bjerrum length (E) to experimental values for Tau

# Field Theory can map out the entire phase diagram of Tau-RNA Complex Coacervation

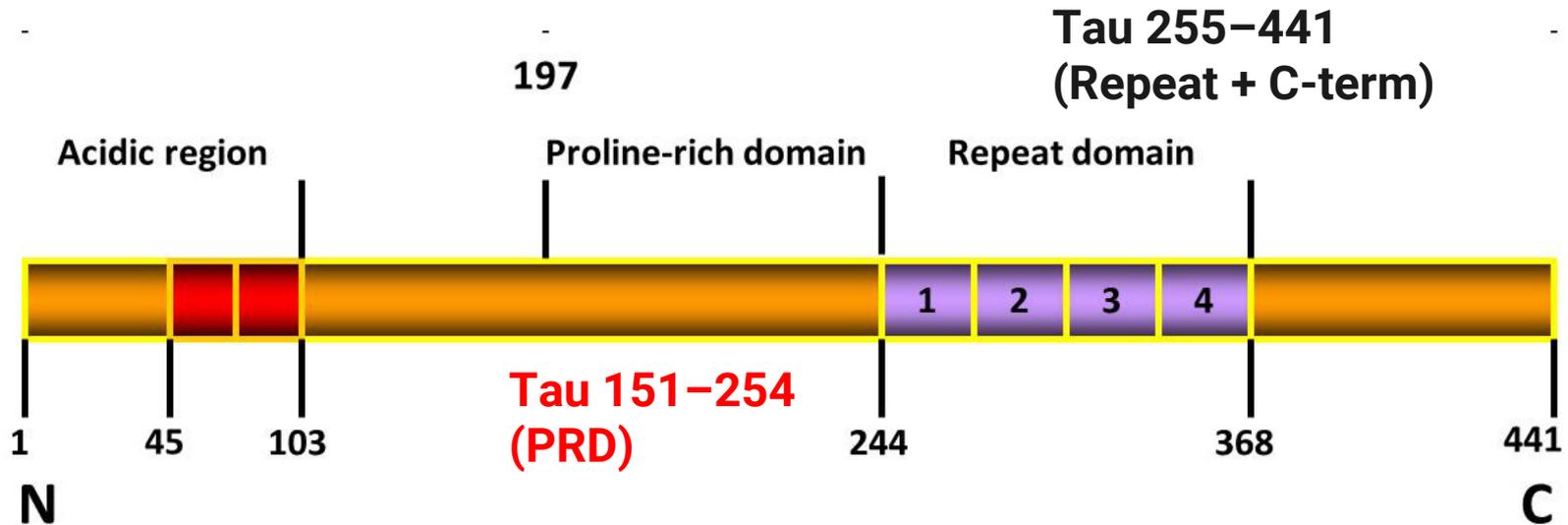


# Field Theory can map out the entire phase diagram of Tau-RNA Complex Coacervation



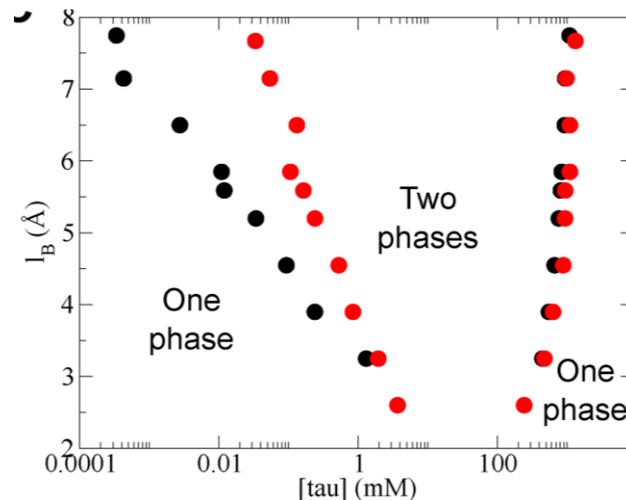
Saeed Najafi

# Tau sequence: The Proline-Rich Domain Liquid-Liquid Phase Separates in vitro

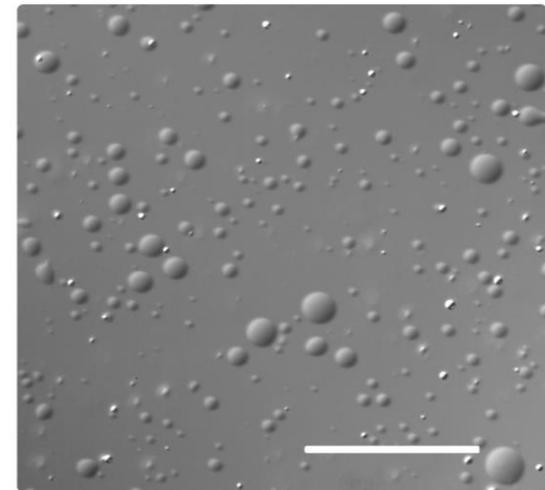


Tau 255–441  
(Repeat + C-term)

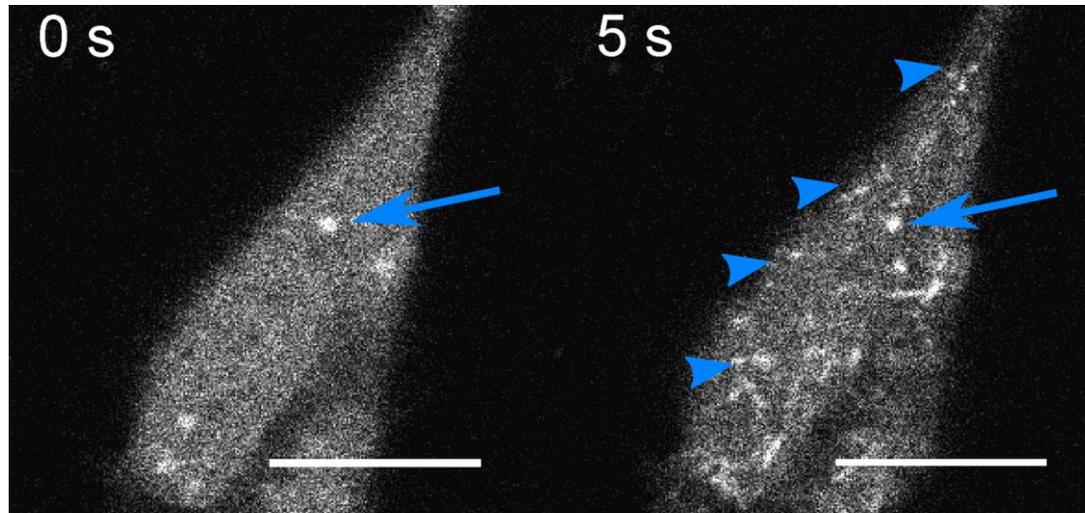
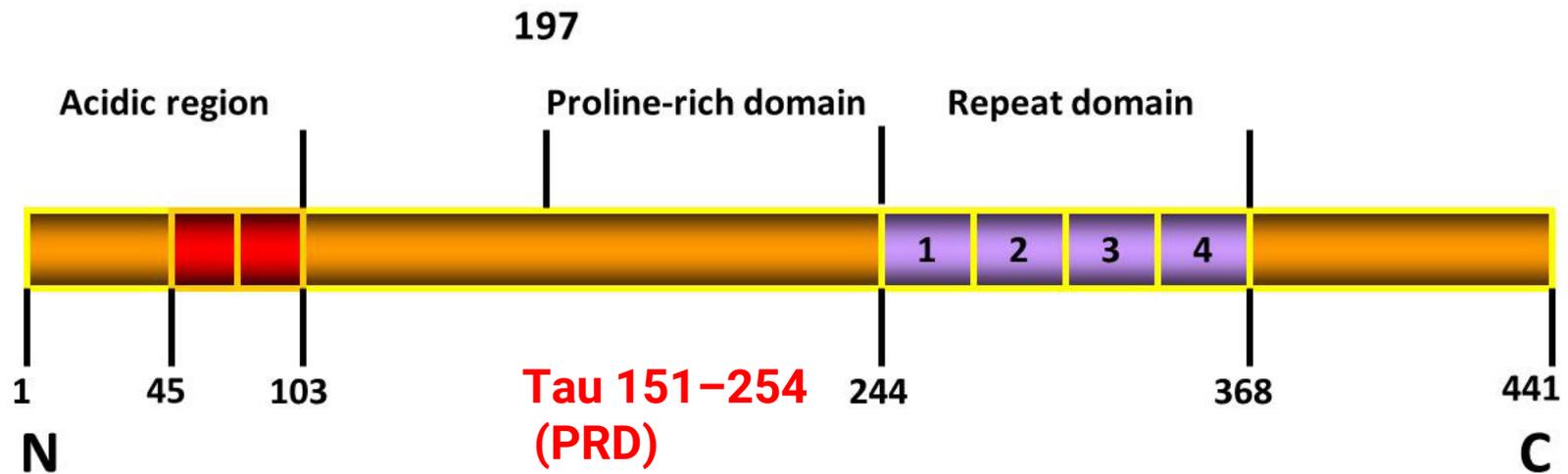
Tau 151–254  
(PRD)



In Vitro

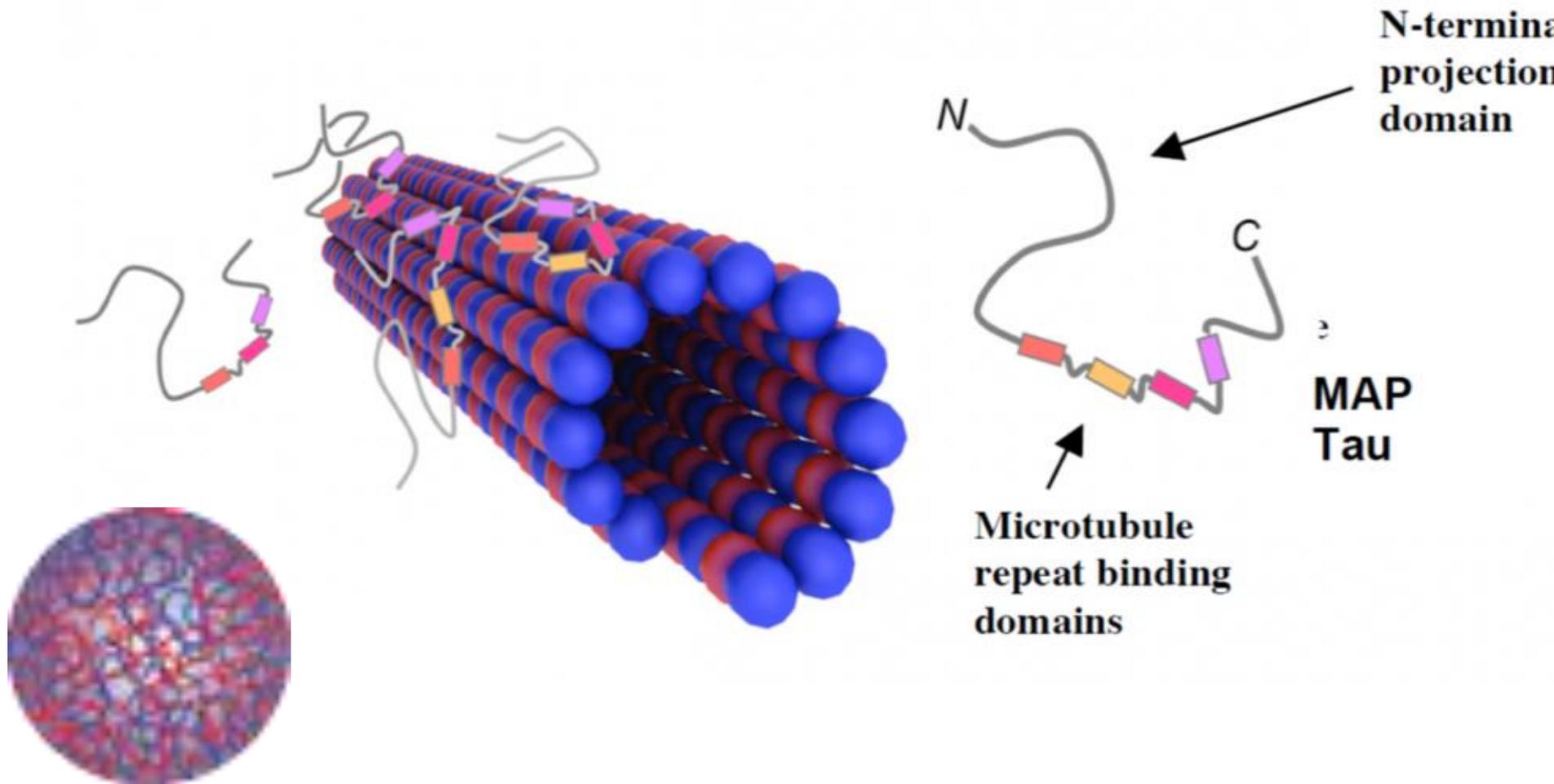


# The Proline-Rich Domain Condensates promote Tau association with MTs



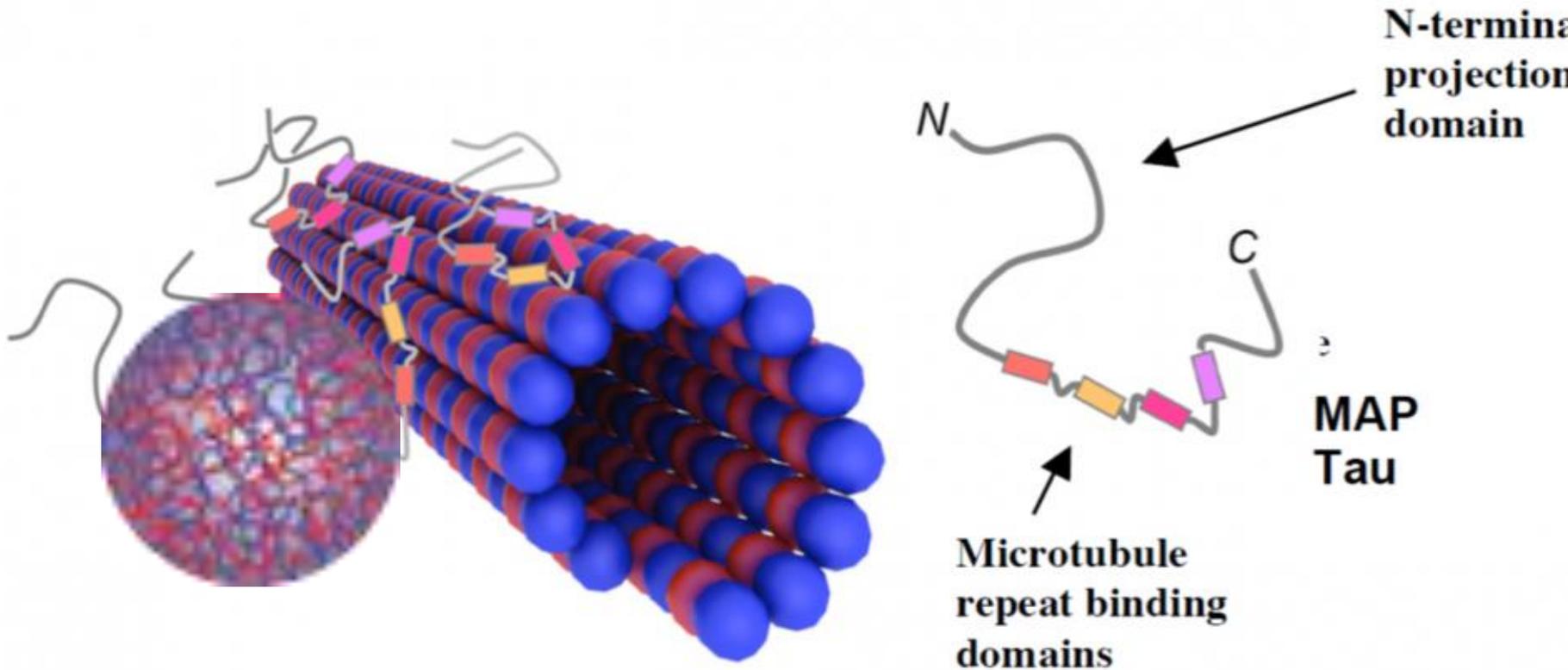
PRD  
condensates  
align along  
the MTs

# MICROTUBULES ARE STABILIZED BY TAU PROTEINS



**LLPS CONCENTRATES TAU AND FACILITATES BINDING**

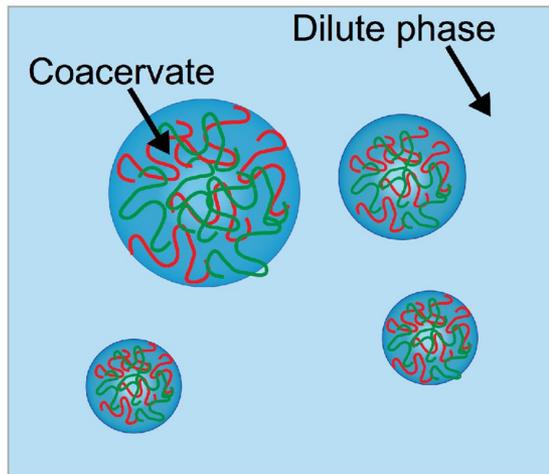
# MICROTUBULES ARE STABILIZED BY TAU PROTEINS



**LLPS CONCENTRATES TAU AND FACILITATES BINDING**

# Proteins can assemble in different ways

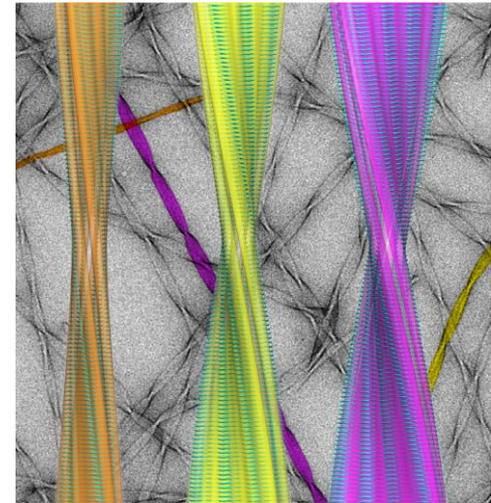
**“Liquid”**



Droplets  
Biomolecular condensates  
Coacervates

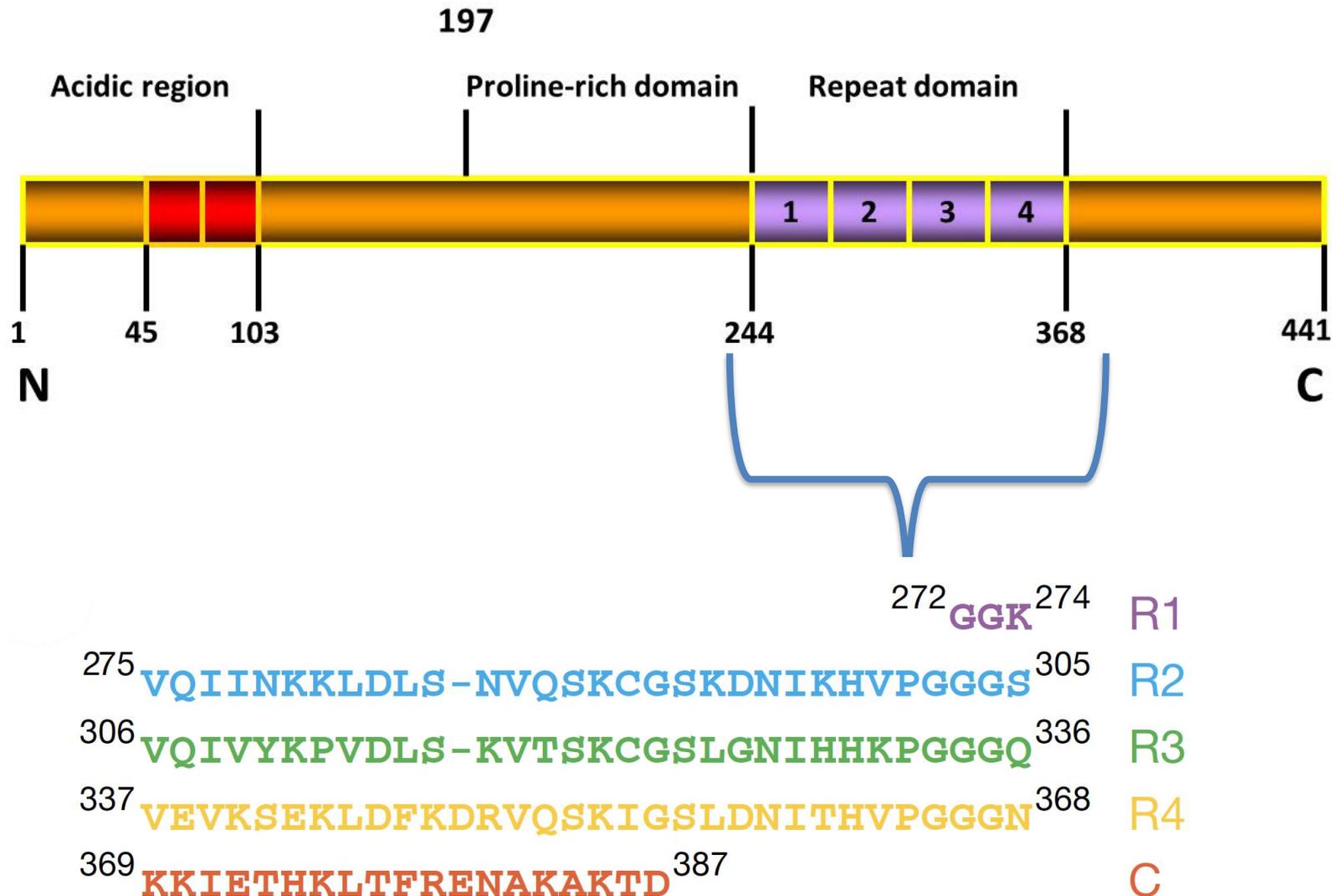
**“Solid”**

Aging  
→

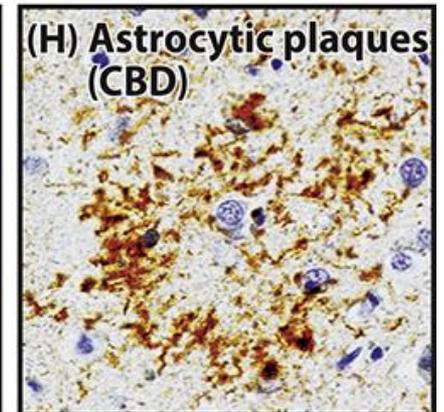
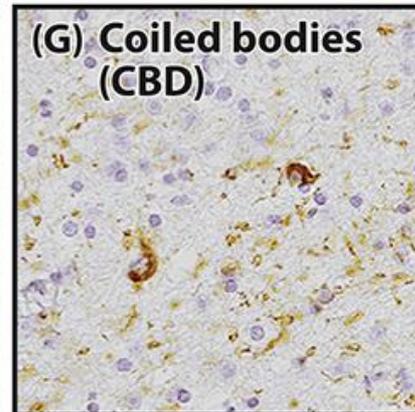
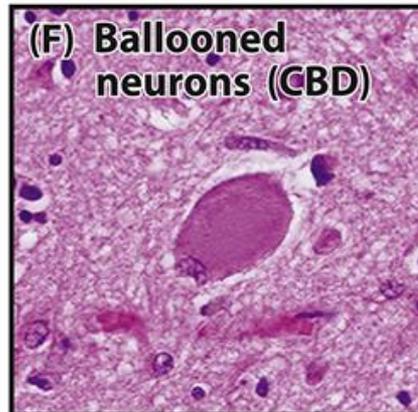
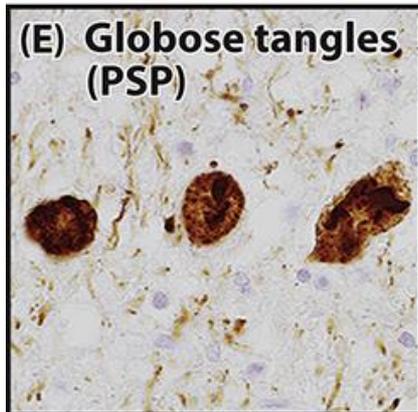
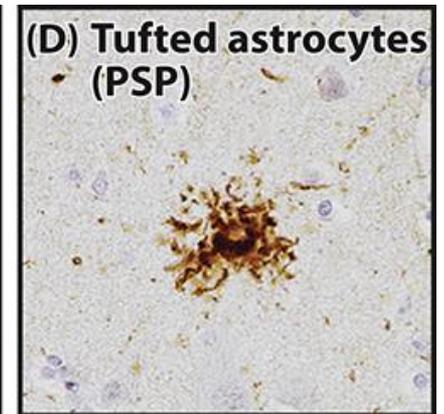
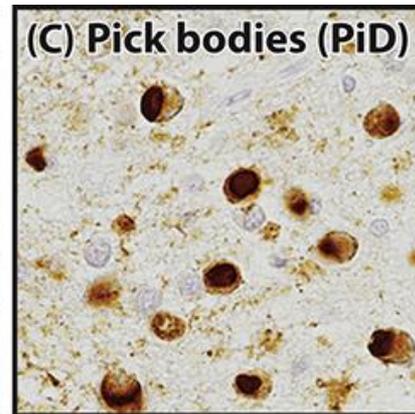
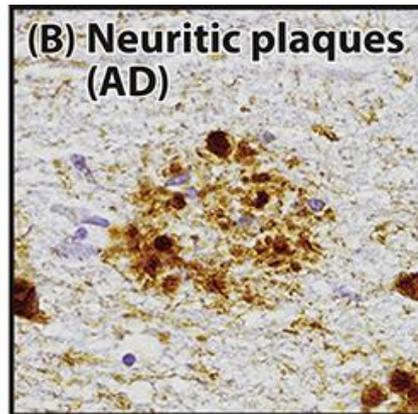
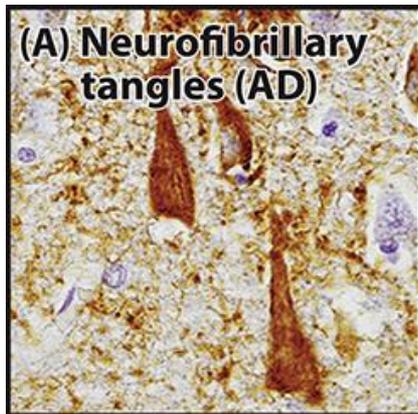


Amyloid Fibrils

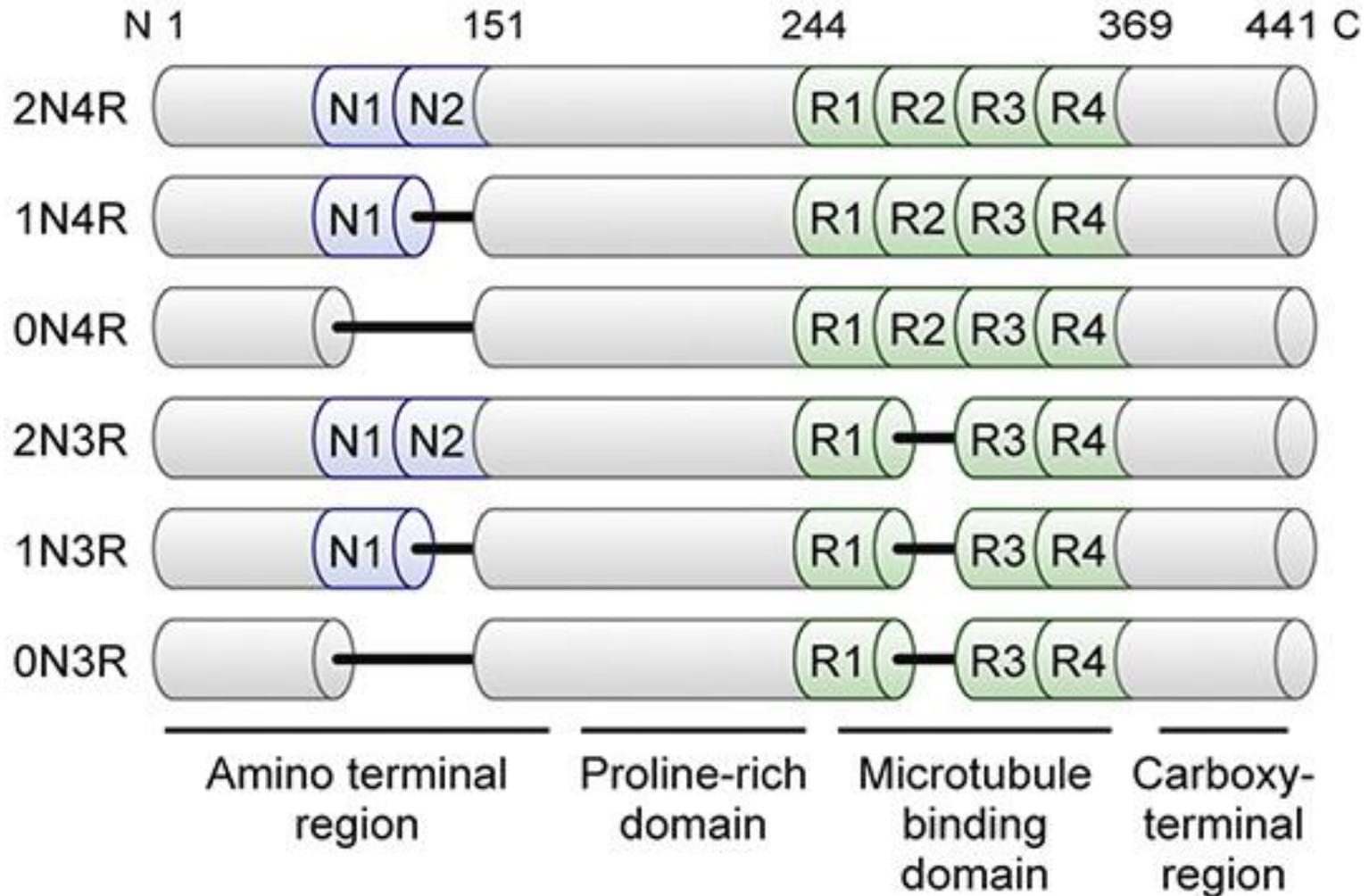
# Tau Aggregation: Repeat Domain makes up the core of Tau Fibrils



# There are many neurodegenerative diseases associated with Tau

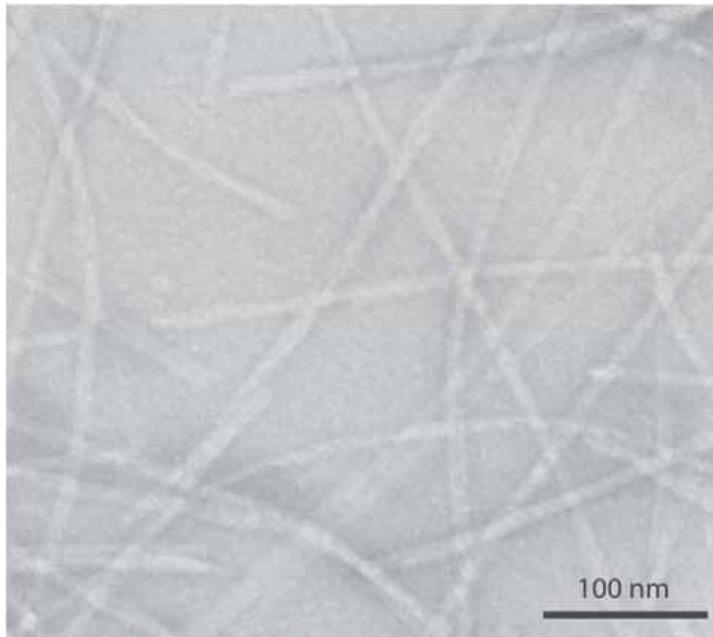


# There are many forms of Tau



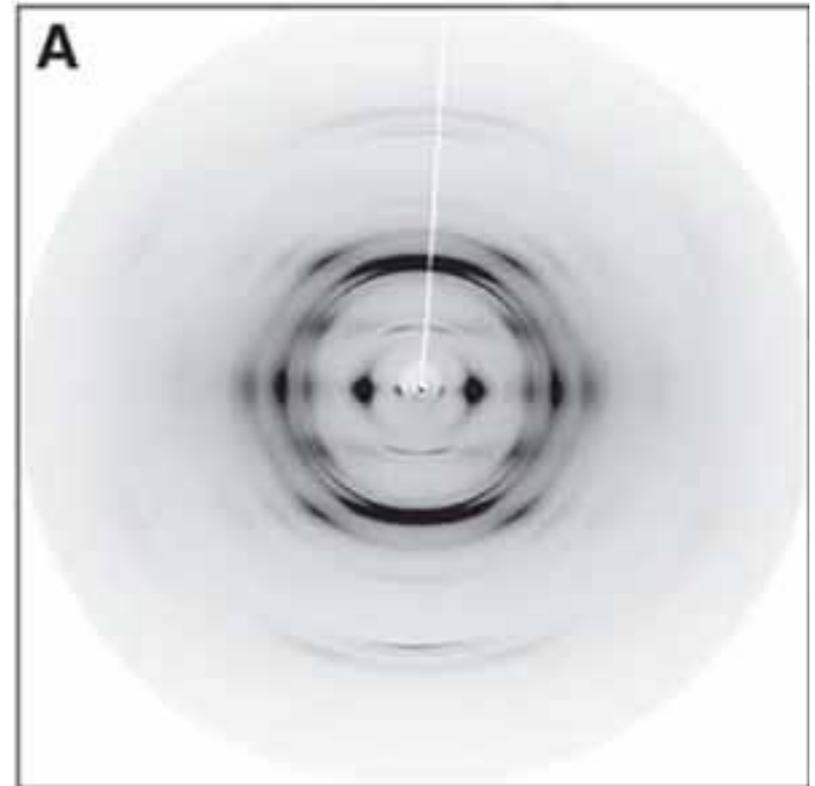
At first glance they all look the same

Transmission Emission  
Microscopy (TEM)



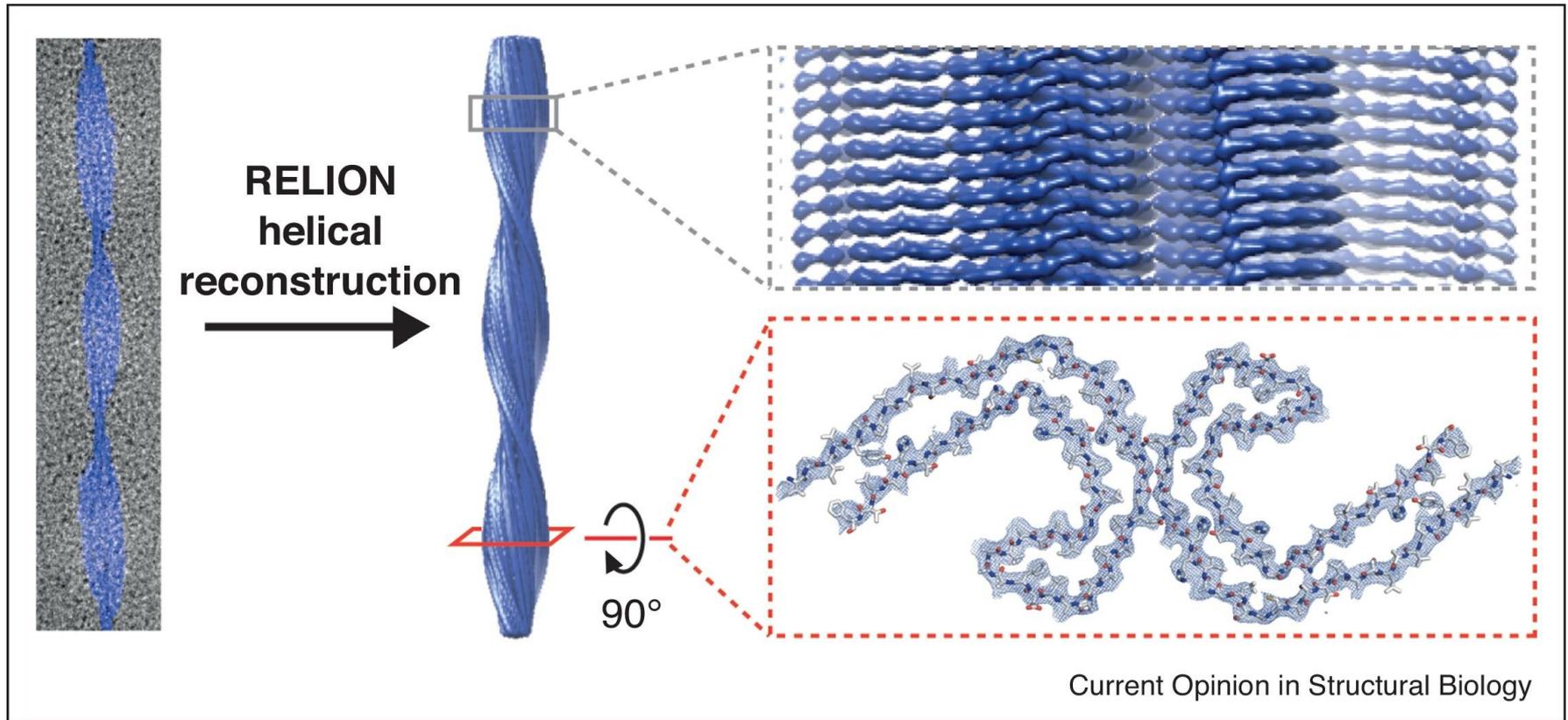
Fibrils

X-Ray diffraction



Cross-beta structure

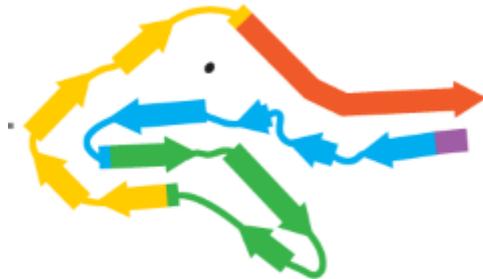
# But Cryo-EM shows (subtle) differences



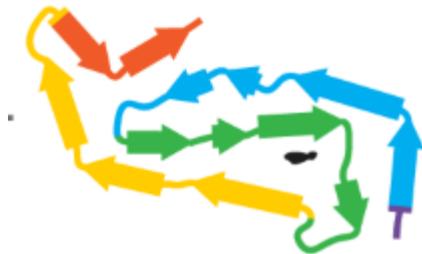
# The specific fibril shape is a signature of a specific disease



CBD



AGD  
ARTAG  
+3 or +16



PSP



GGT-I  
GGT-II



GPT

# COMMON STRAND-LOOP-STRAND MOTIF



CBD

AGD



# Created: 19 Amino-acid Peptide



WILD TYPE: jR2R3

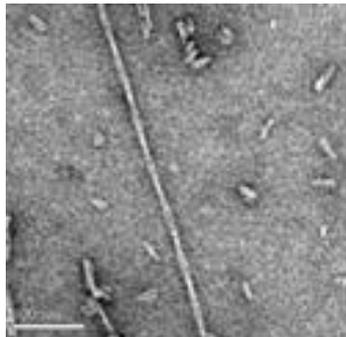


MUTANT: jR2R3-P301L

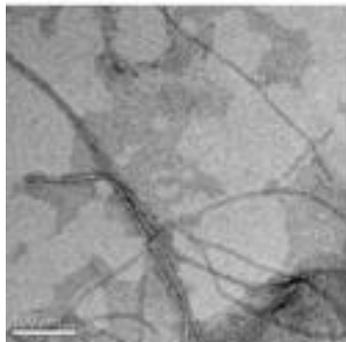


**jR2R3-P301L mutant** aggregates faster than jR2R3 and shows more fibril morphologies

jR2R3

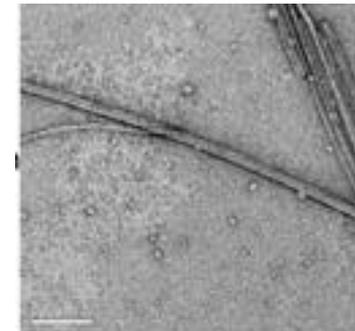
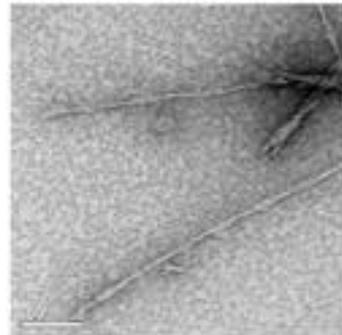


PHF

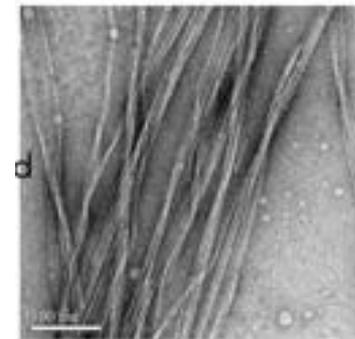
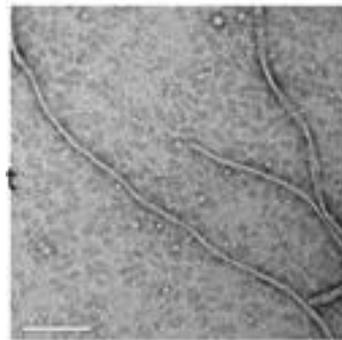


Straight

**jR2R3-P301L**



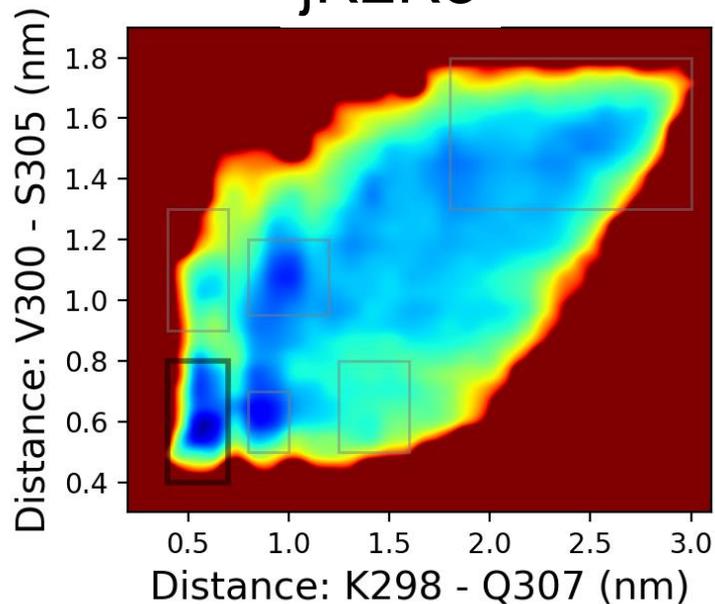
Ribbon



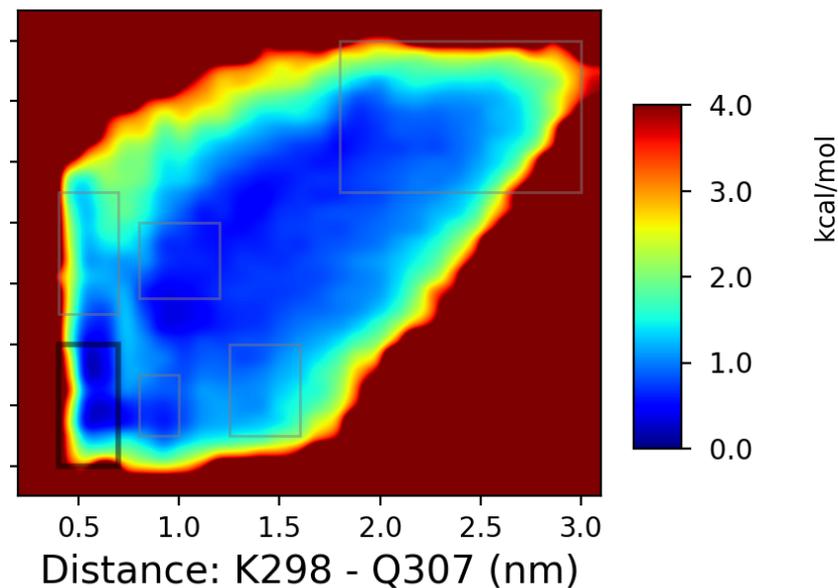
Bundle

# jR2R3-P301L can explore more conformational space

## jR2R3

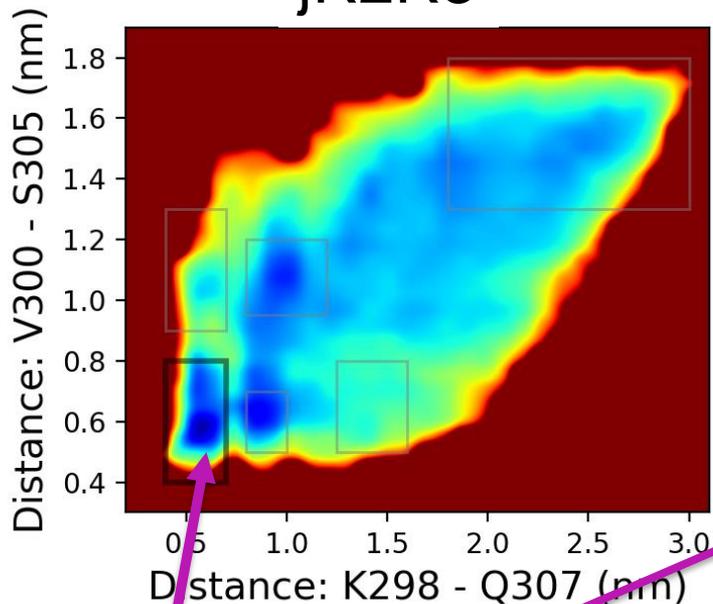


## jR2R3-P301L (mutant)

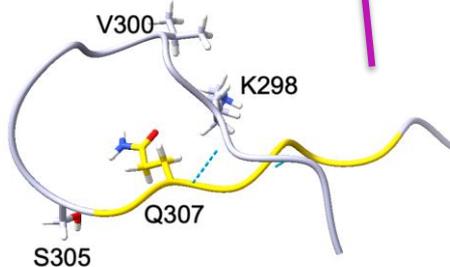
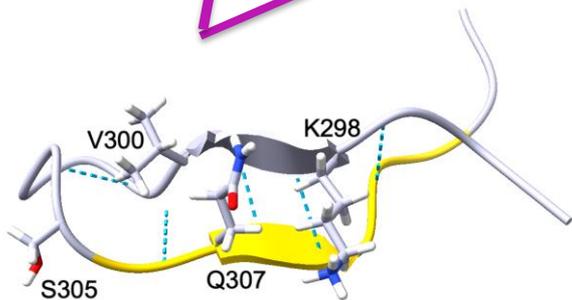
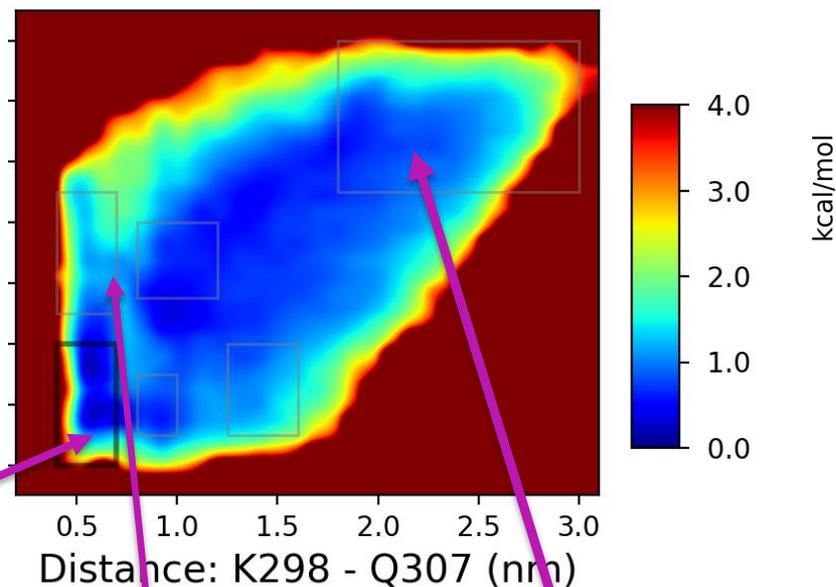


# jR2R3-P301L can get out of a "fibril protective" hairpin

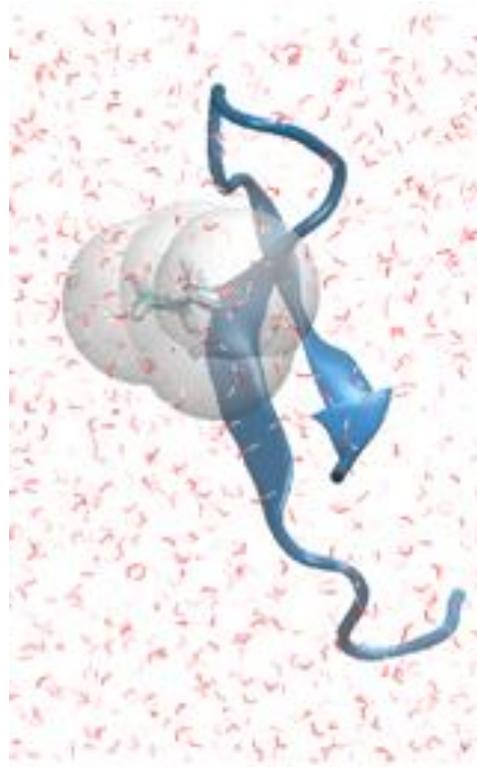
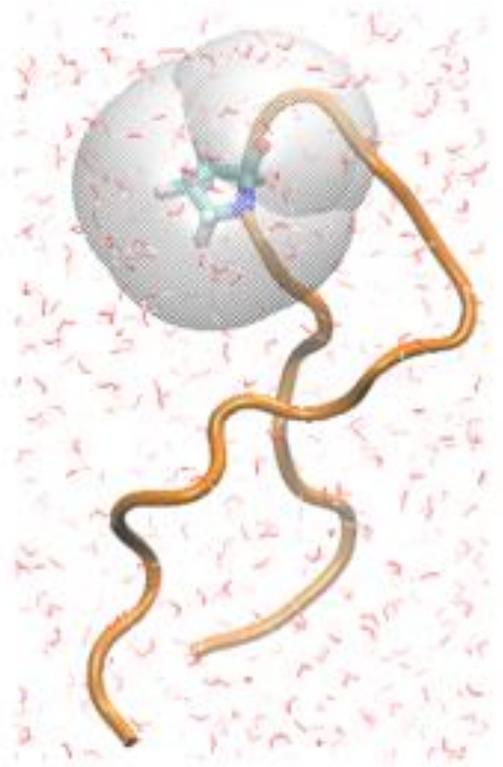
## jR2R3



## jR2R3-P301L (mutant)



Overhauser Dynamic Nuclear Polarization (ODNP) experiments show a reduction in hydration water dynamics around 301 site for the P→L mutant (jR2R3-P301L)



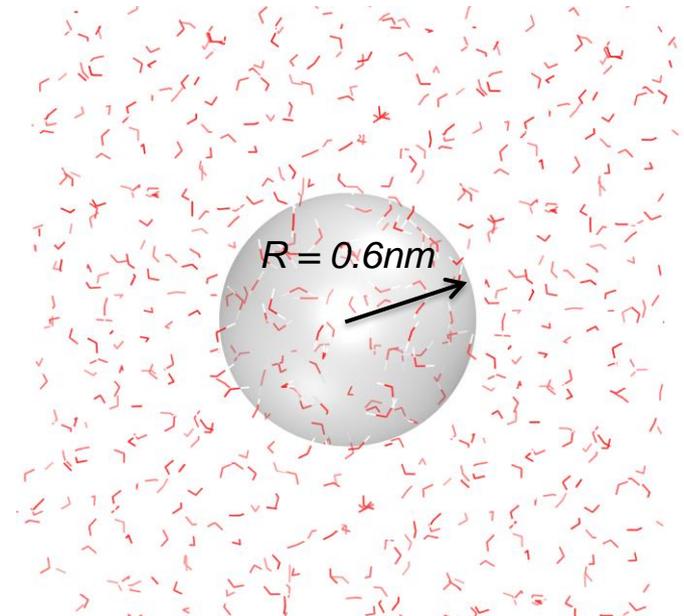
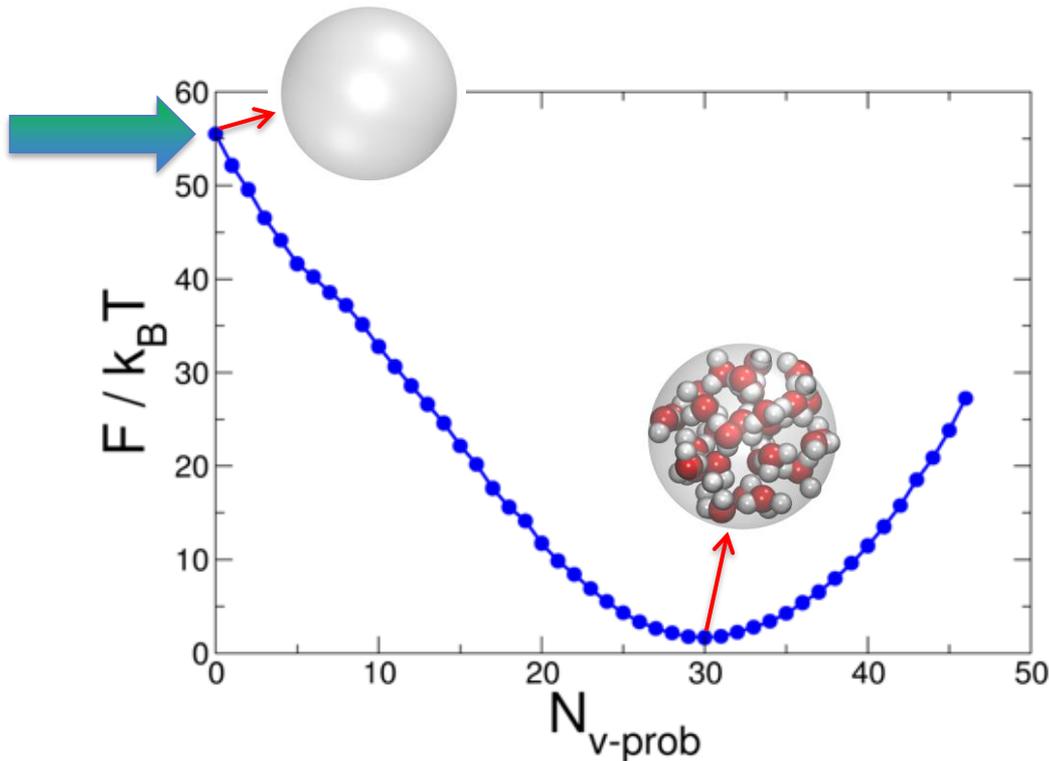
Increased ordering of water around mutation site

→ locally more hydrophobic

# Probing Hydrophobicity Computationally through Umbrella Sampling (INDUS)

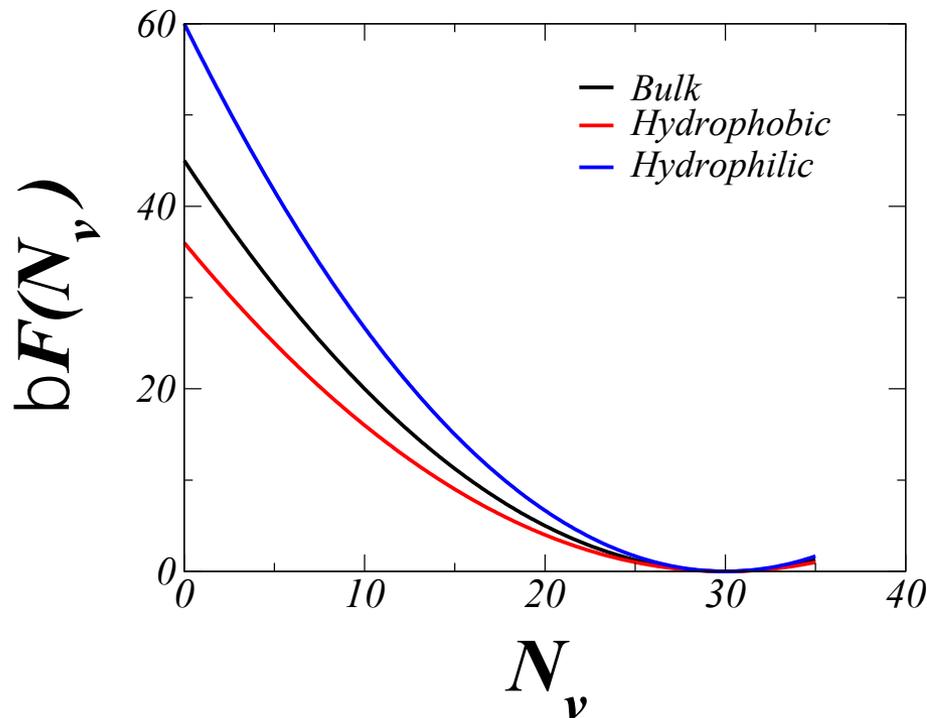
Free energy of dewetting a spherical volume in bulk

$$\mu_{\text{ex}} = \mathbf{F}(\mathbf{0}) = -\ln \mathbf{P}_{\mathbf{v}}(\mathbf{0})$$

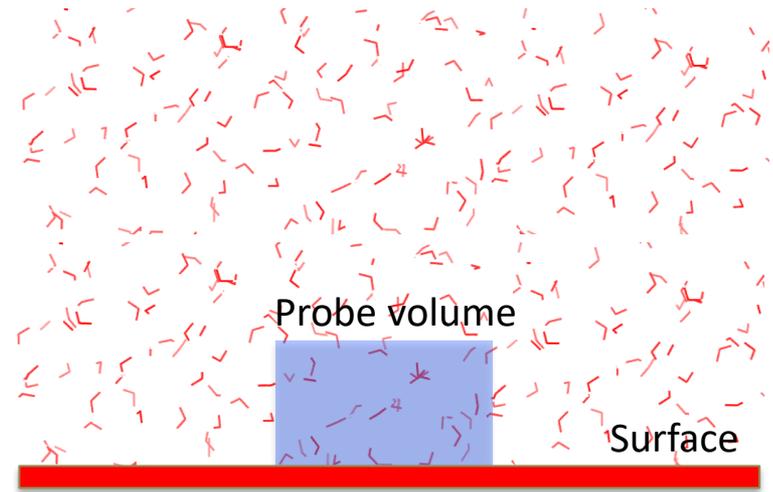


# Free energy of dewetting the probe volume in vicinity of a surface

Excess chemical potential is an indication of hydrophilicity or hydrophobicity of the surface

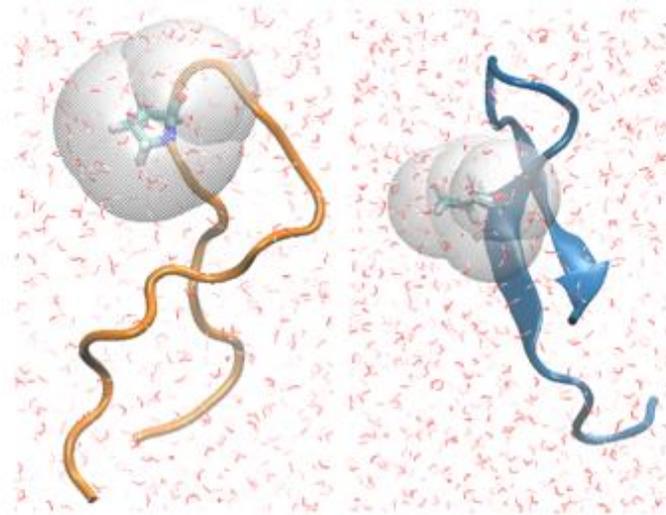


$$\mu_{\text{ex}}^{\text{phil}} > \mu_{\text{ex}}^{\text{bulk}} > \mu_{\text{ex}}^{\text{phob}}$$



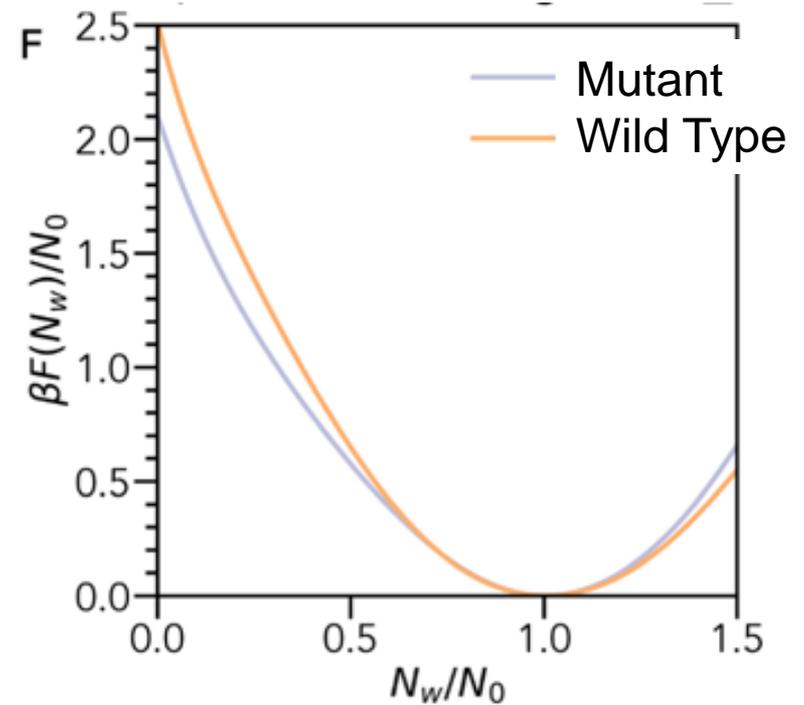
$$\mu_{\text{ex}} = \mathbf{F}(0) = -\ln P_v(0)$$

# Free energy of dewetting lower for jR2R3-P301L: an additional factor favoring association of jR2R3-P301L

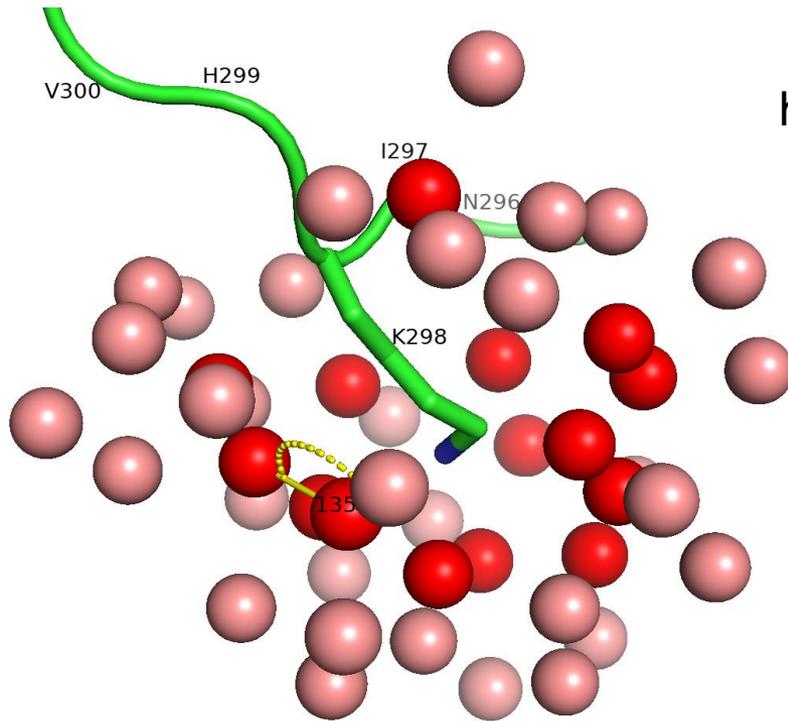


Wild Type

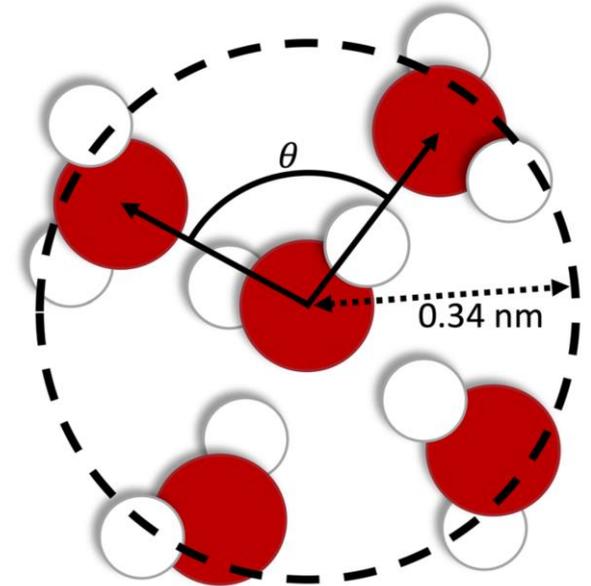
Mutant



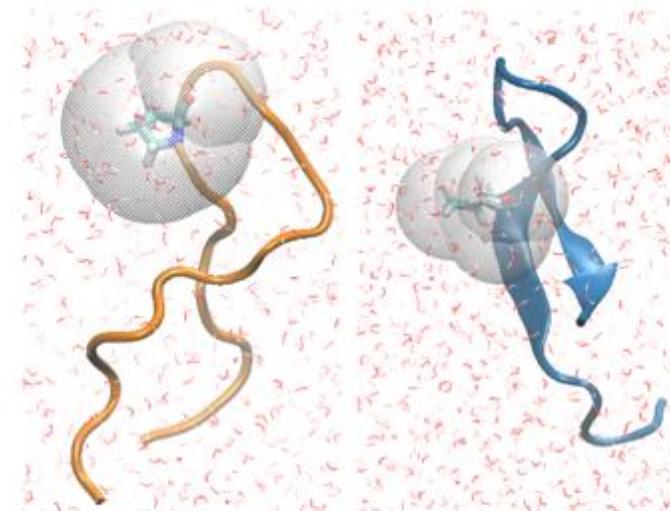
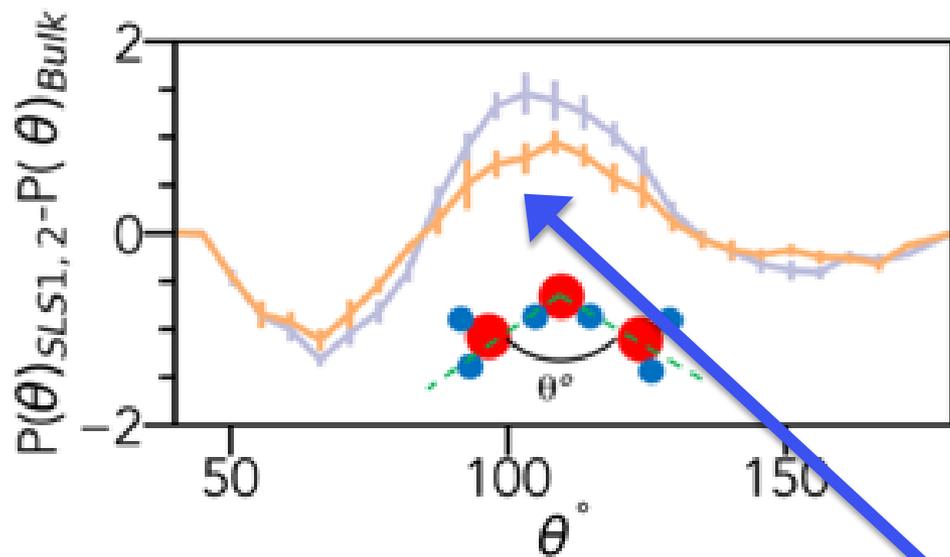
# Quantifying Water Structure: *water triplet distribution*



histogram  
the  
angles



Increased tetrahedral ordering of hydration waters near the L301 (mutant) compared to P301 (wild type)

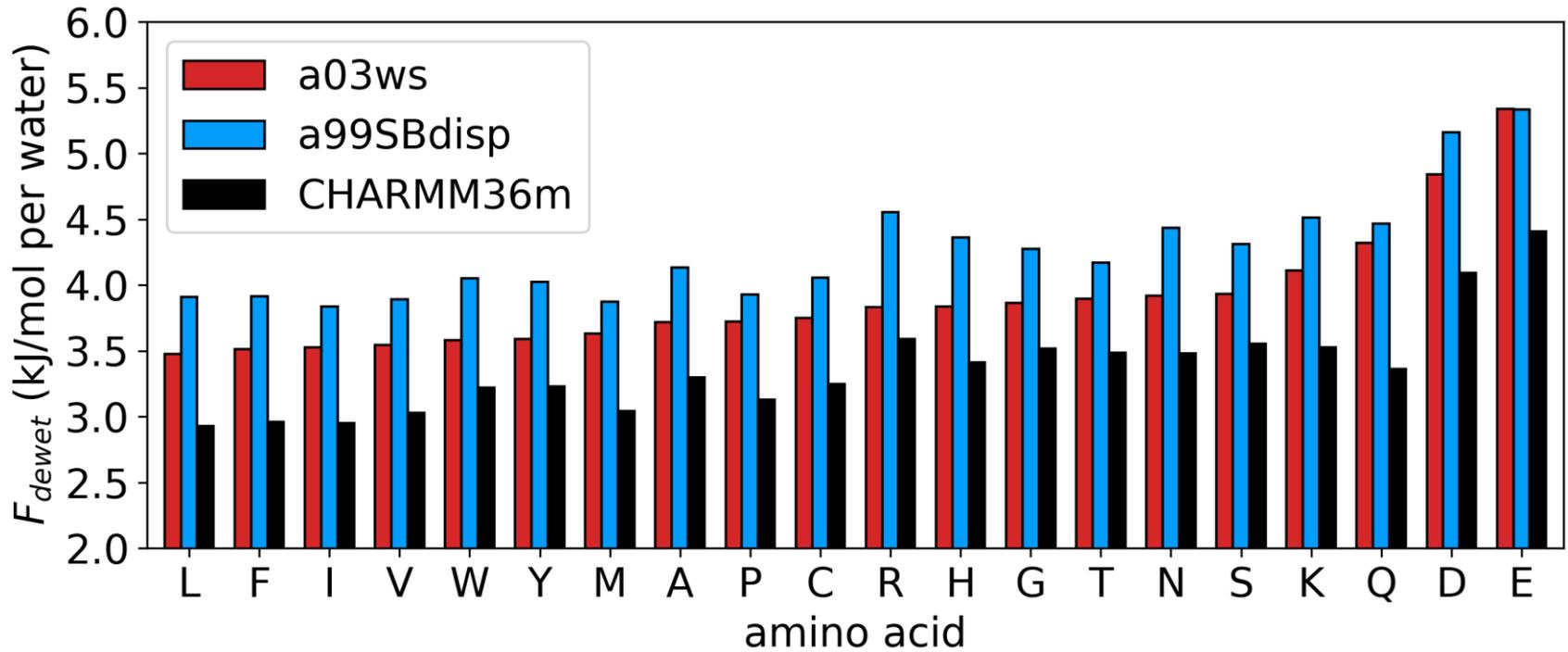


Wild Type

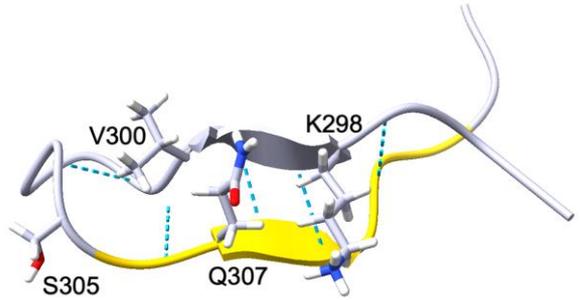
Mutant

TETRAHEDRAL  
“hydrophobic”

# Different Force Fields Show Different Hydrophobicity



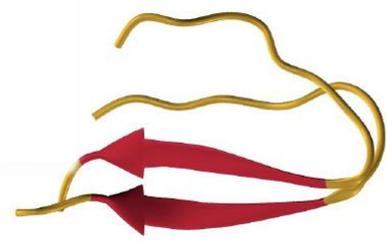
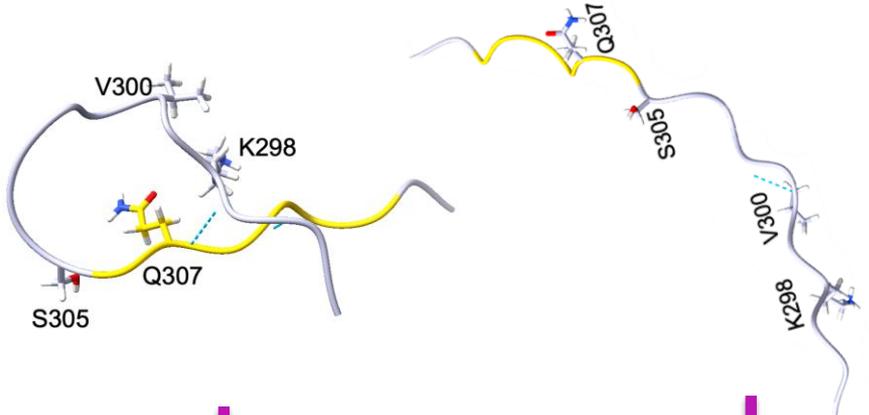
”fibril protective structure”



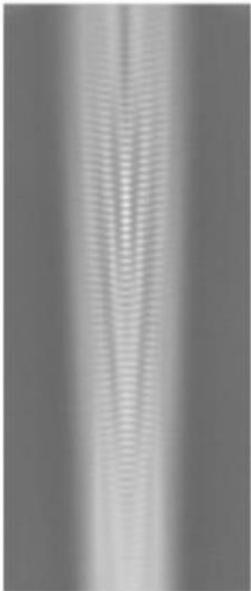
Dead End



”fibril inducing structure”



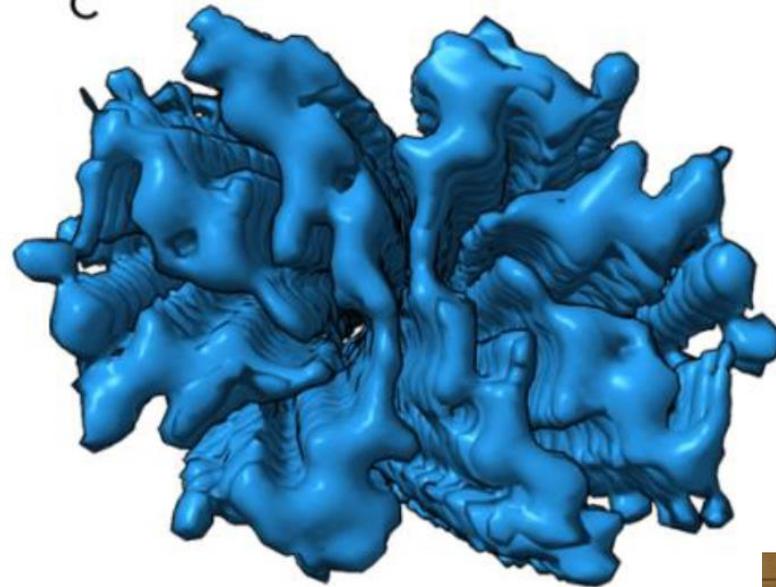
# Recent CryoEM structure of jR2R3-P301L



B



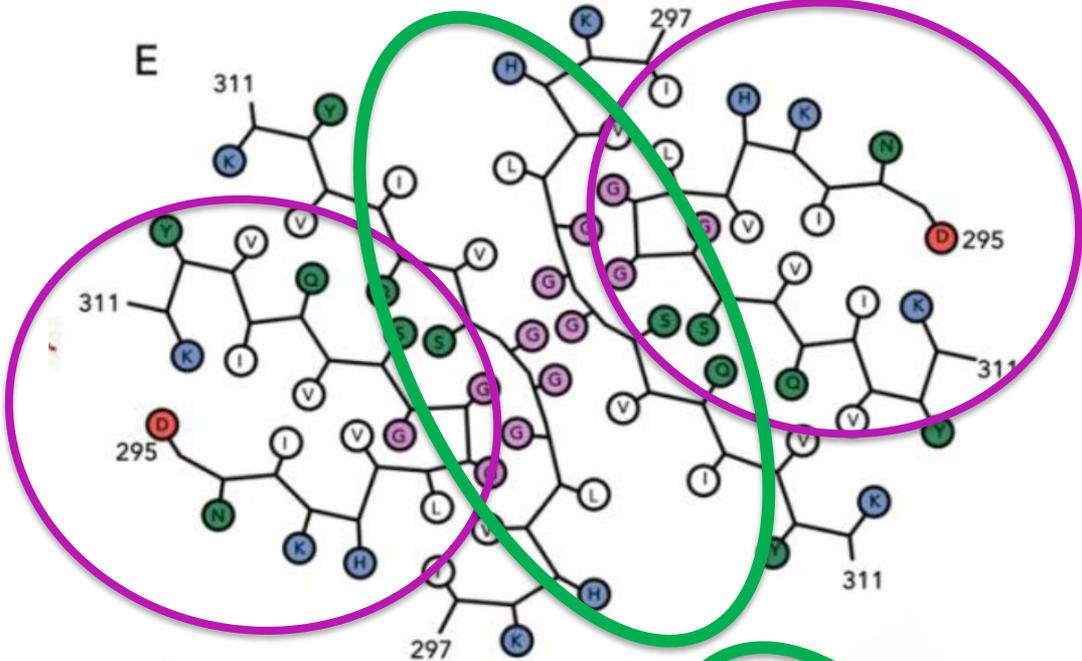
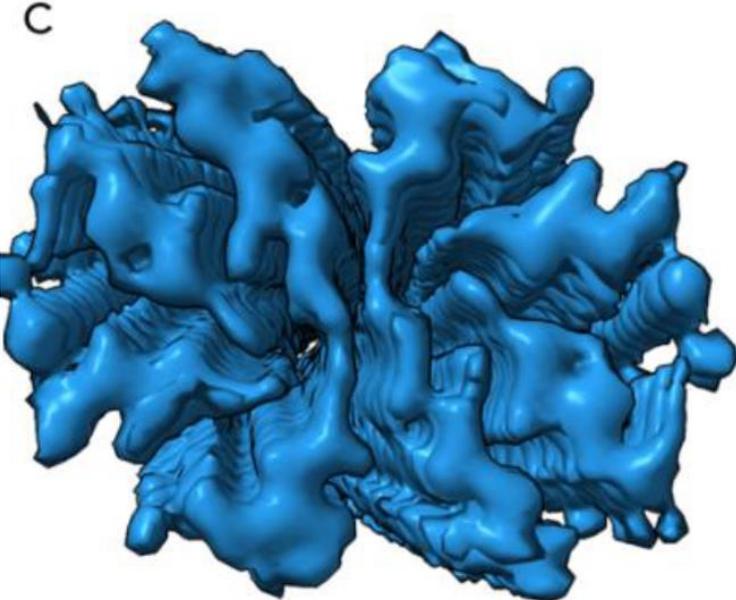
C



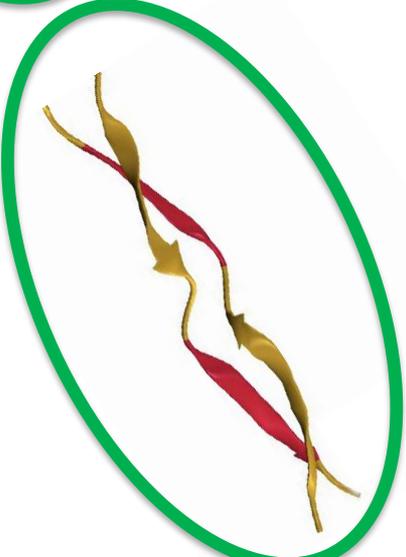
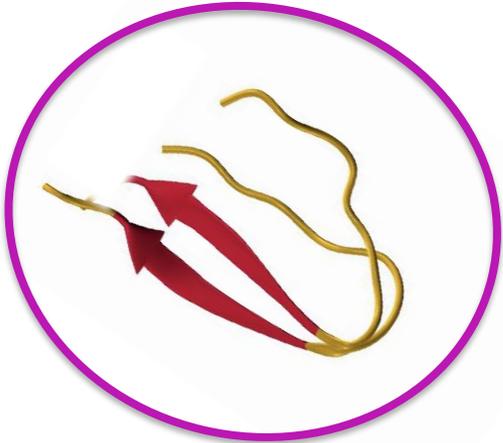
Professor Songi Han  
UCSB/  
Northwestern University



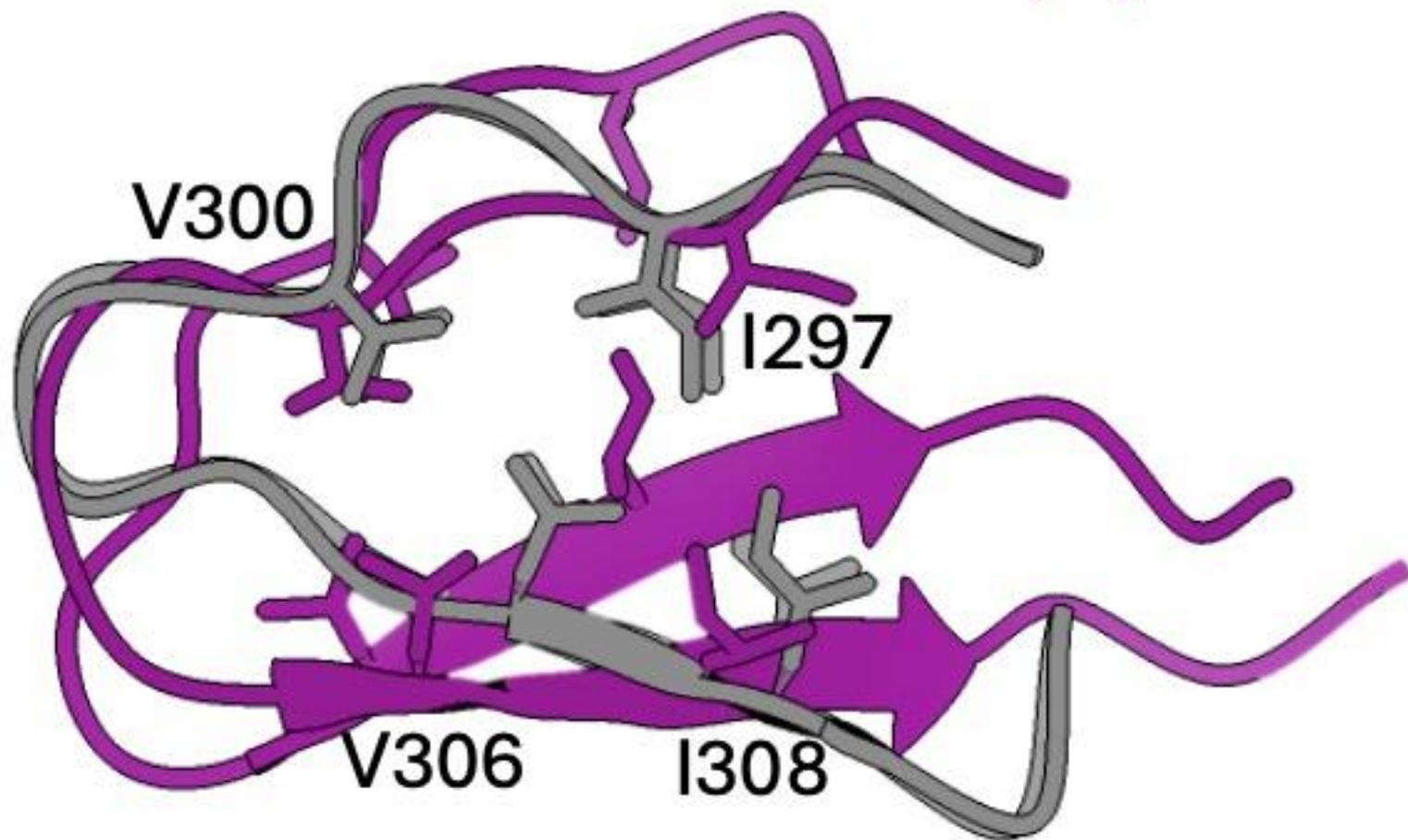
# CryoEM structure of jR2R3-P301L



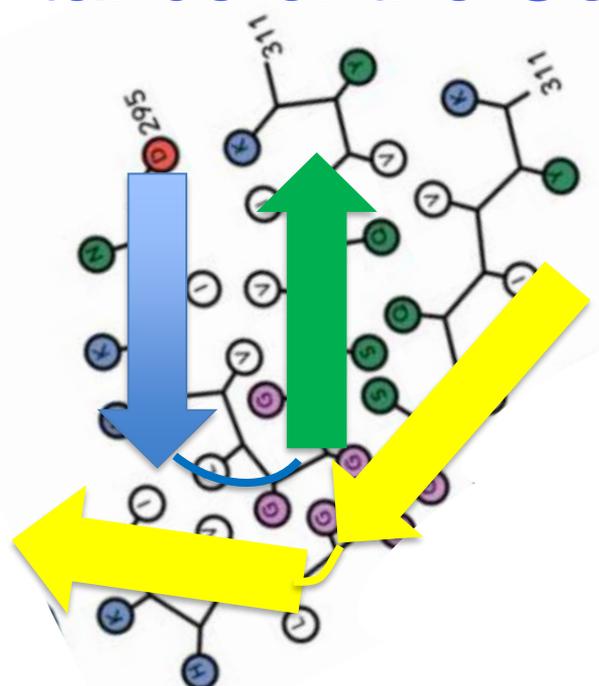
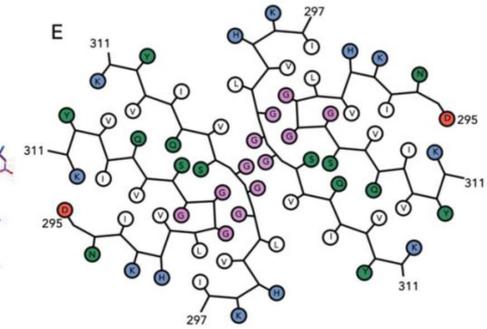
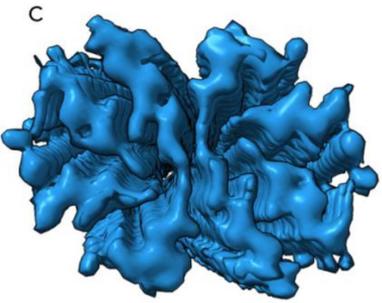
Simulation



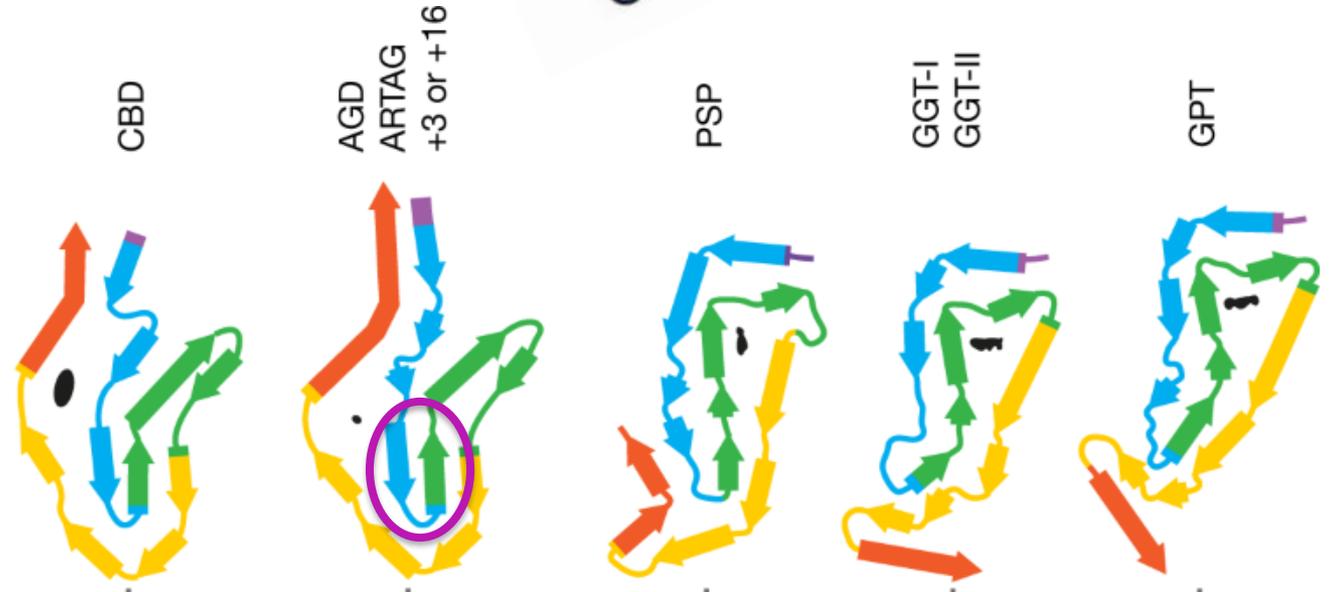
cryo-EM structure  
from CHARMM36m (iv)



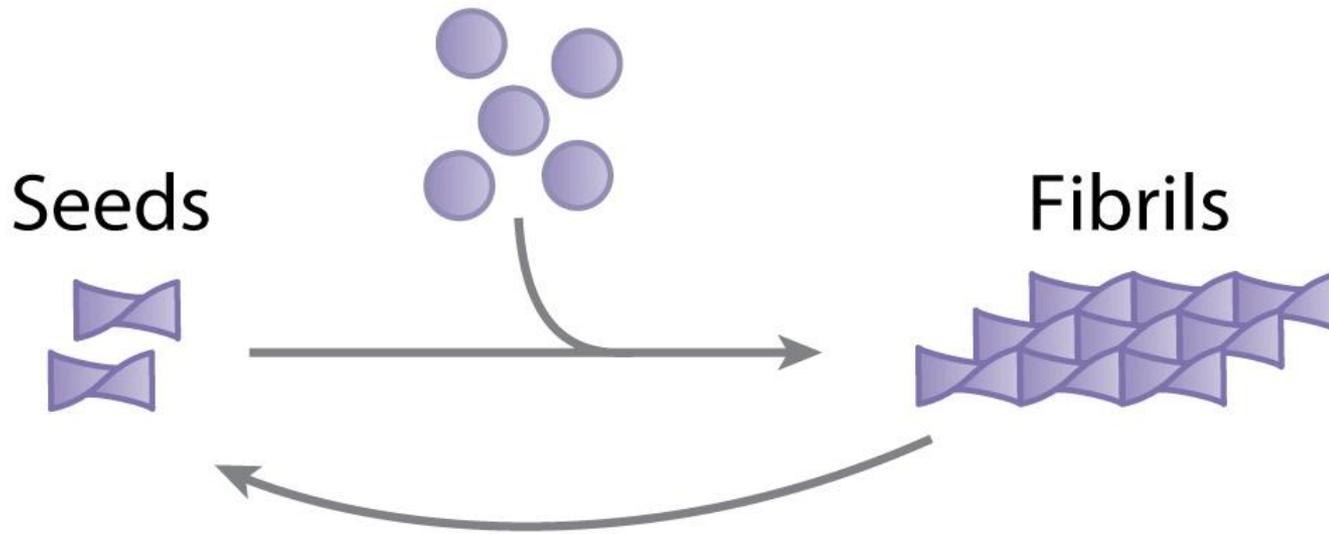
# Importance of the Counter-Strand



jR2R3-P301L

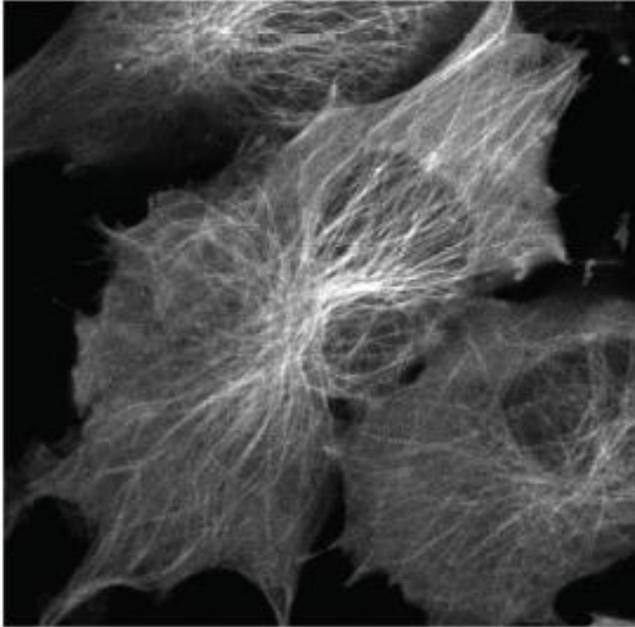


# jR2R3-P301L can seed the fibrillization of full length Tau in Vivo

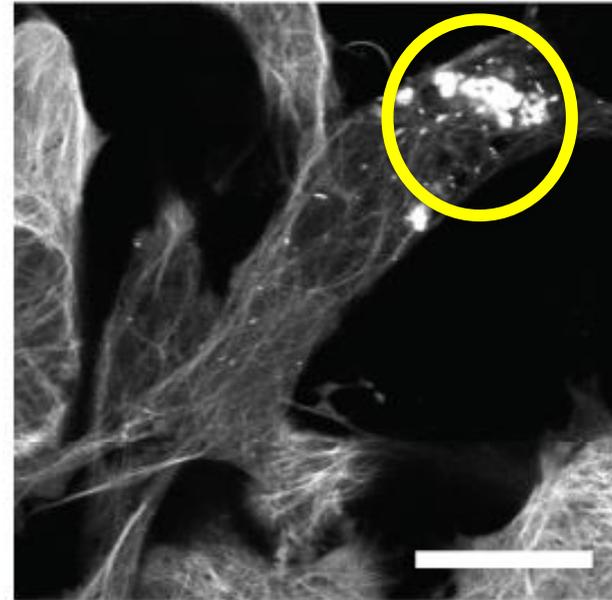


# jR2R3-P301L can seed the fibrillization of full length Tau in Vivo

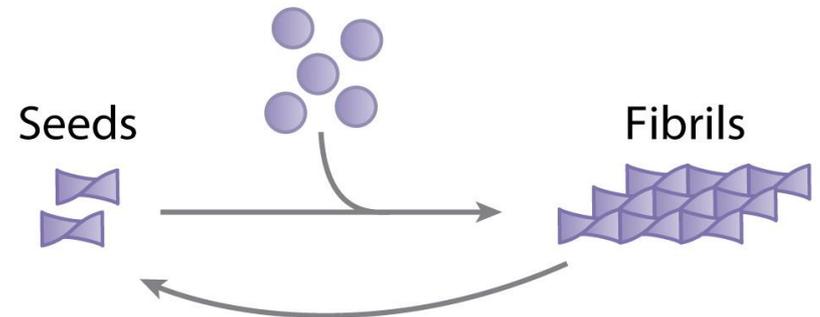
Before jR2R3 P301L



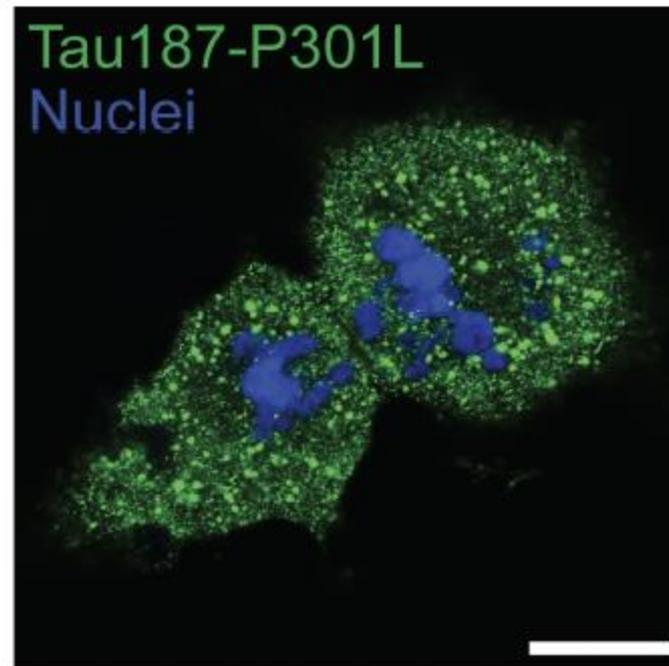
After jR2R3 P301L



Prof. Ken  
Kosik, UCSB



# jR2R3-P301L acts as a prion: propagates the strain

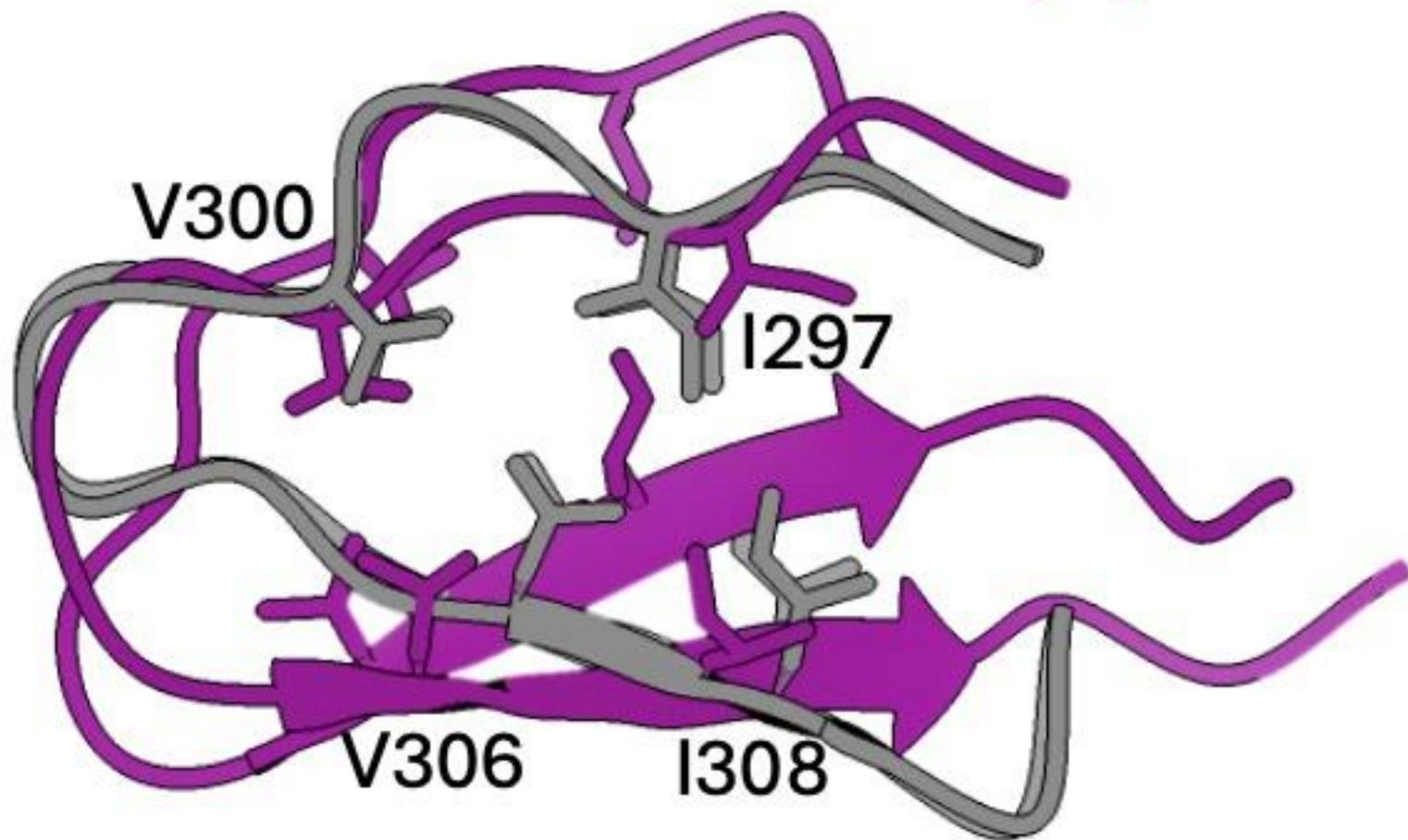


PNAS (2024)  
121 (15) e2320456121

Cells seeded with jR2R3-P301L fibrils undergo division and propagate aggregates to daughter cells



cryo-EM structure  
from CHARMM36m (iv)

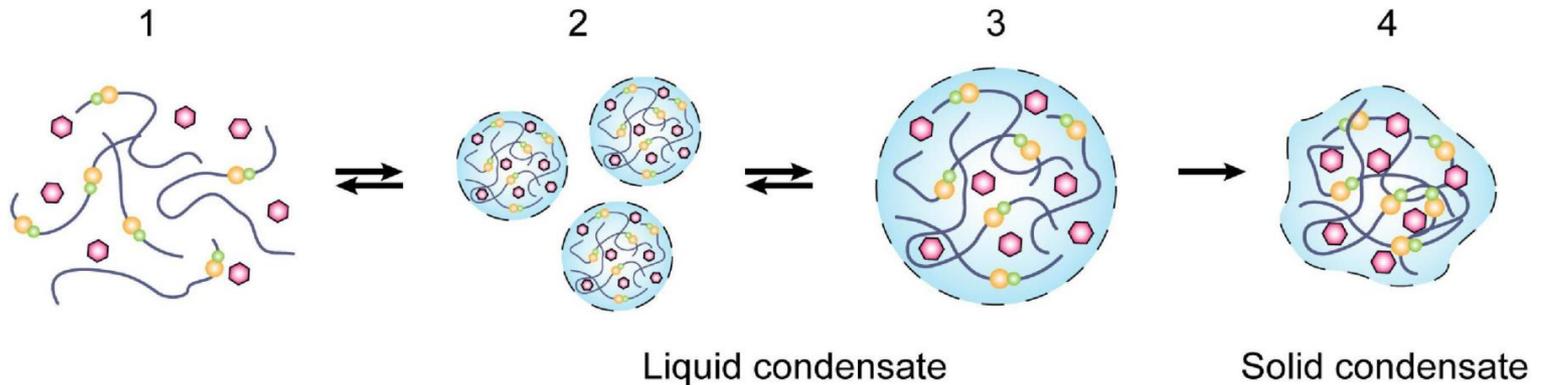
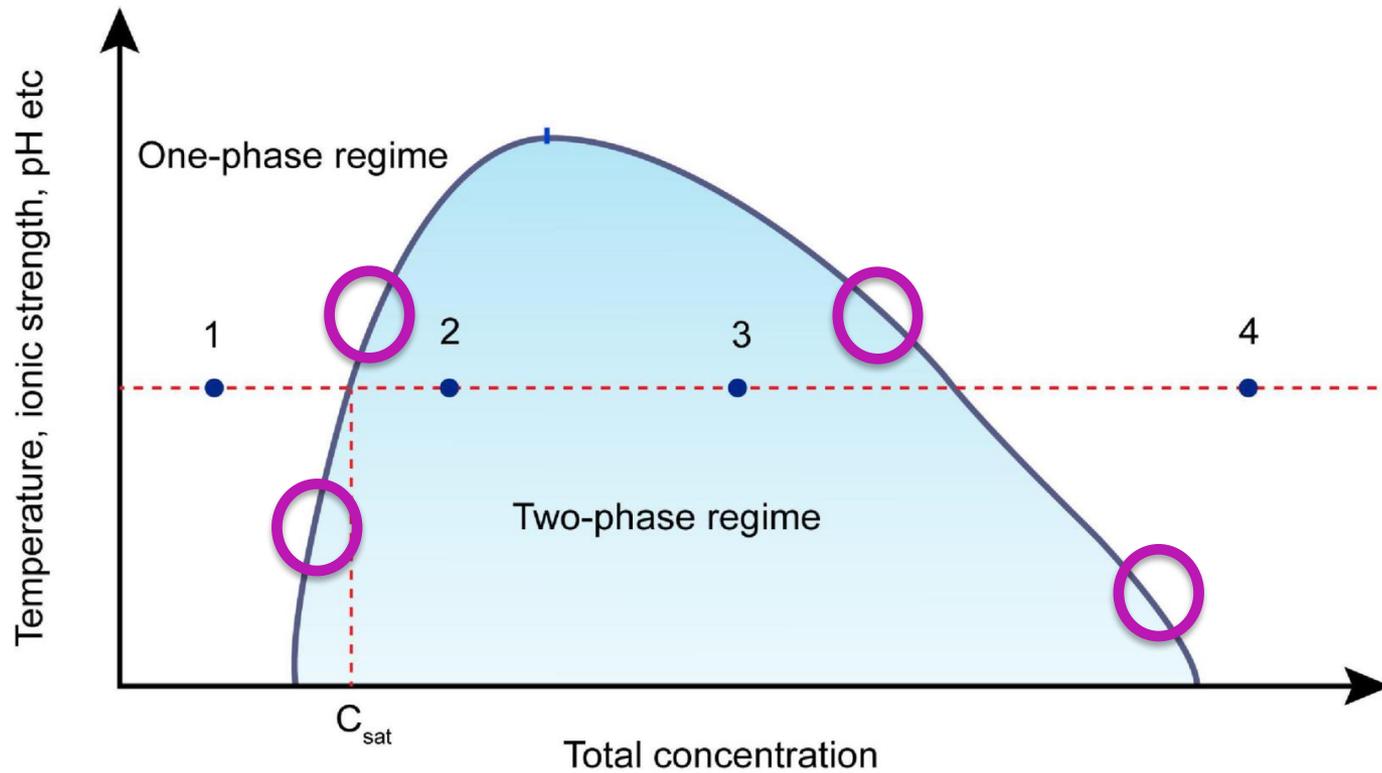


Back-up Slides





# We map the boundaries of the phase diagram



# Field Theoretic Model

Partition function for a system of charged Gaussian chains and salt in implicit solvent

$$Z = Z_0 \int Dw \int D\psi e^{-H[w,\psi]}$$

Excluded volume interactions

Electrostatic interactions

$$H[w, \psi] = \frac{1}{2B} \int d\mathbf{r} w(\mathbf{r})^2 + \frac{1}{2E} \int d\mathbf{r} |\nabla\psi|^2 - \sum_l \frac{CV\bar{\phi}_l}{N_l} \ln Q_l$$

Fluctuating chemical potential field

Fluctuating electrostatic potential field

Single chain partition function for polymers and salt ions (complex valued)

**B:** dimensionless excluded volume parameter

**E:** dimensionless Bjerrum length

**C:** dimensionless monomer density

# Field Theoretic Simulations

Complex Langevin equations of motion

$$\frac{\partial w(r, t)}{\partial t} = -\lambda_w \frac{\delta H[w, \psi]}{\delta w(r, t)} + \eta_w(r, t)$$
$$\frac{\partial \psi(r, t)}{\partial t} = -\lambda_\psi \frac{\delta H[w, \psi]}{\delta \psi(r, t)} + \eta_\psi(r, t)$$

← real Gaussian white noise

Ensemble averages over the field configurations

$$\langle O \rangle = \frac{\int Dw \int D\psi O e^{-H[w, \psi]}}{\int Dw \int D\psi e^{-H[w, \psi]}}$$



# Discrete Gaussian chain polyelectrolyte model

dimensionless monomer density

$$\boxed{C} \sim \rho b^3$$

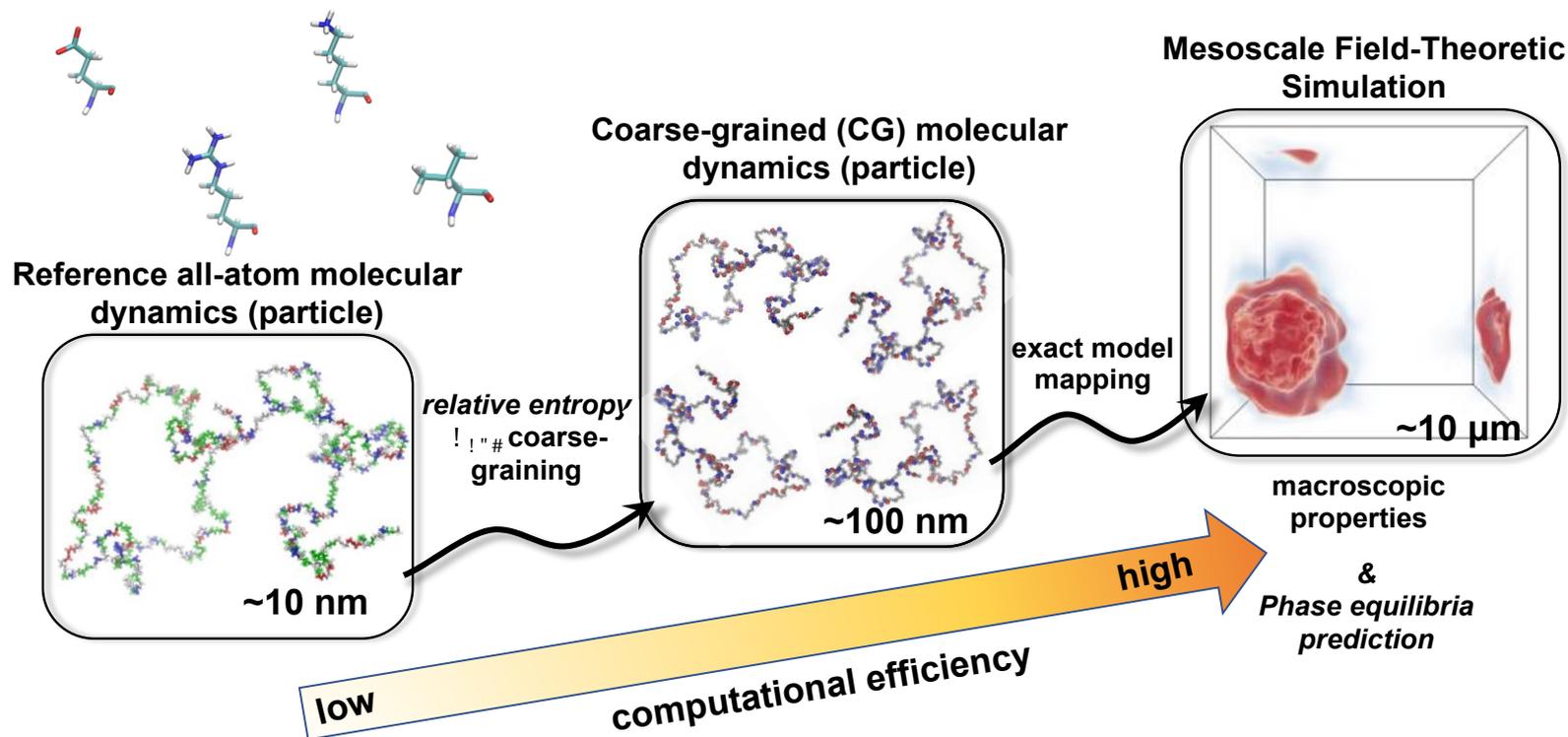
dimensionless excluded volume parameter

$$\beta U = \frac{v}{2} \int d\mathbf{r} \bar{\rho}^2(\mathbf{r}) \quad \boxed{B} \sim \frac{v}{b^3}$$

dimensionless Bjerrum length

$$\beta U = \frac{l_B}{2} \int d\mathbf{r} \int d\mathbf{r}' \frac{\bar{\rho}_e(\mathbf{r}) \bar{\rho}_e(\mathbf{r}')}{|\mathbf{r} - \mathbf{r}'|} \quad \boxed{E} \sim \frac{l_B}{b}$$

# Coarse graining through minimizing $S_{rel}$ (Information Loss)



## Chemical Detail & transferability

Minimizing the relative entropy (information-loss):

$$S_{rel} = \int \int \rho_{AA}(\mathbf{r}) \ln \left( \frac{\rho_{AA}(\mathbf{r})}{\rho_{CG}(\mathbf{R})} \right) \delta(\mathbf{M}(\mathbf{r}) - \mathbf{R}) d\mathbf{r} d\mathbf{R}$$

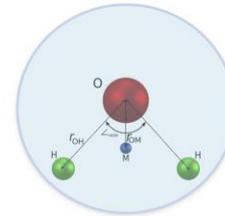
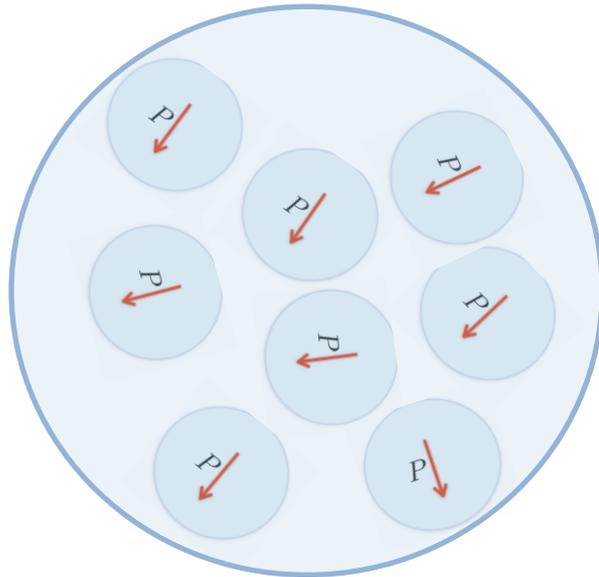
Shell Adv.  
Chem. Phys.  
2016



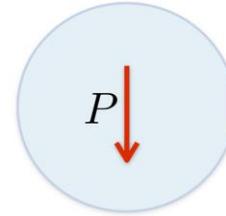
# Field theoretic simulation water model

## FTS simulation in NVT ensemble

8 AA waters → 1 CG water



Atomistic model

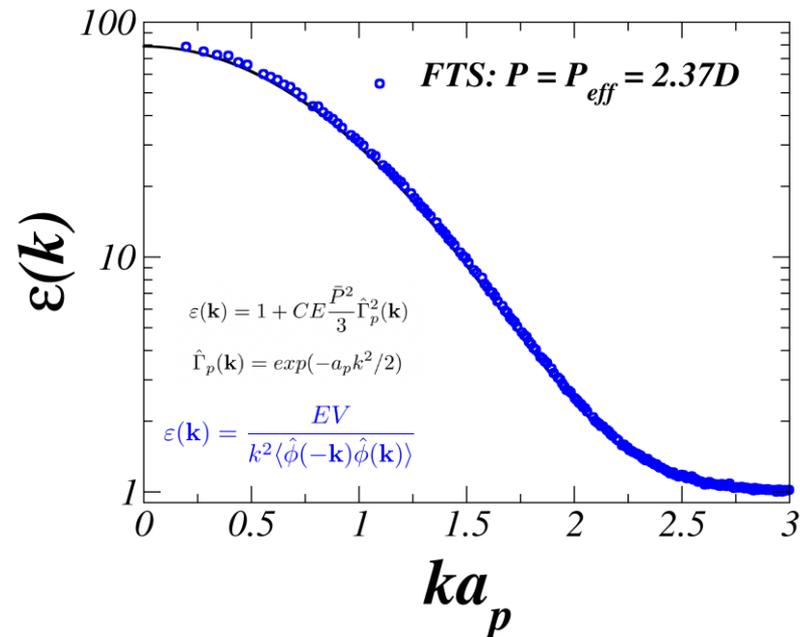


Dimer model

$$\beta\bar{U}[\mathbf{r}] = \frac{\nu}{2} \int d\mathbf{r} \bar{\rho}^2(\mathbf{r}) + \frac{l_B}{2} \iint d\mathbf{r} d\mathbf{r}' \frac{\bar{\rho}_e(\mathbf{r})\bar{\rho}_e(\mathbf{r}')}{|\mathbf{r} - \mathbf{r}'|}$$

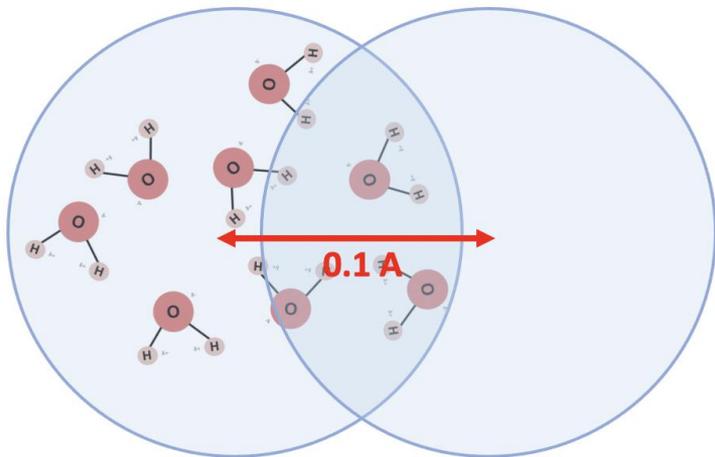
$$\bar{\rho}(\mathbf{r}) = \int d\mathbf{r}' \Gamma(|\mathbf{r} - \mathbf{r}'|) \hat{\rho}(\mathbf{r}')$$

$$\Gamma(r) = \left( \frac{1}{2\pi a^2} \right)^{3/2} \exp(-r^2/2a^2)$$



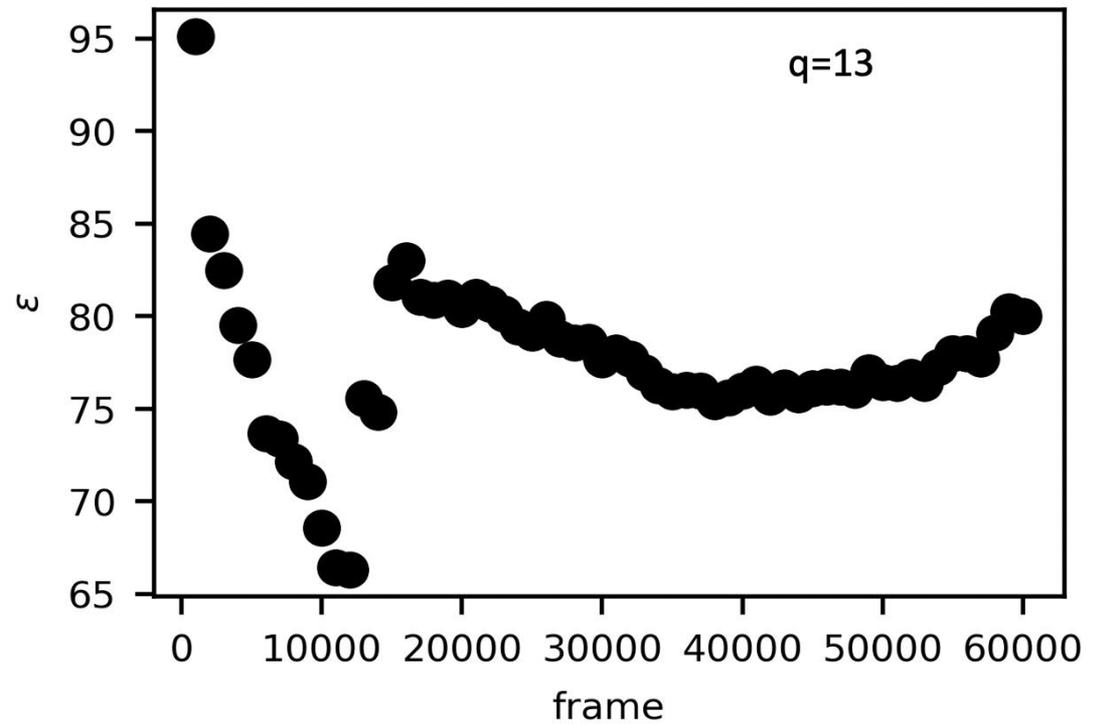
# MD coarse-grained water model

8 AA waters  $\rightarrow$  1 CG water



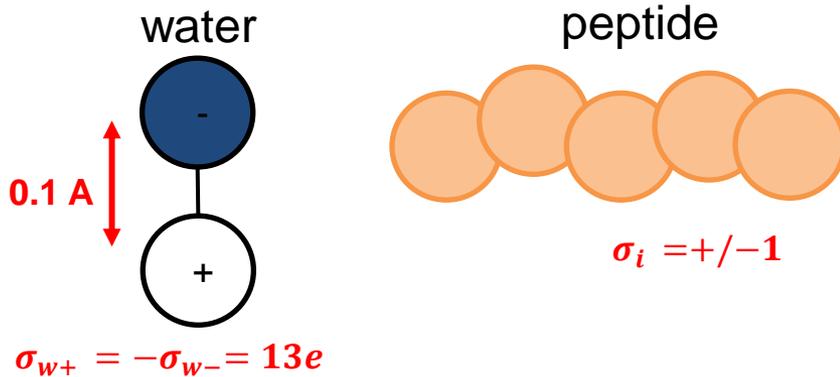
$$\sigma_{w+} = -\sigma_{w-} = 13 e$$

Match  $\epsilon = 80$  at 300 K



# Relative entropy parametrization of peptide and water interaction

RE1: Poly-Arg/Glu 



$$\sigma = a_i = R_w = 0.158 \text{ nm}$$

- $\beta U_{bond,water}$ : stiff harmonic with  $b = 0.01 \text{ nm}$
- $\beta U_{bond,residue}$ : harmonic with  $b = \sqrt{6} \sigma$
- Electrostatics:  $\beta U_{el,W-,W+}, \beta U_{el,W+,W+}, \beta U_{el,W-,W-}$

- Excluded volumes:

- $u_{W-,W-} = u_{Lys,Lys} = u_{Cl,Cl} = u_{W-,Cl} = u_{Lys,Cl} = 0.1 \text{ kT } \sigma^3$
- $u_{W-,residue} = [B^{Srel} (4\pi\sigma^2)^{\frac{3}{2}} \sigma^3] N_{ref}^2 \sigma^{-3} = B_{W-,residue}^{Srel} (4\pi)^{3/2} \sigma^3$

## Coarse-grain interactions

$$U_{CG} = \sum_{bonds} U_{bond} + \sum_i^{N_T-1} \sum_{j=i+1}^{N_T} U_{ev}(r_{ij}) + U_{el}(r_{ij})$$

$$\beta U_{bond}(r_{ij}) = \frac{3}{2b^2} (r_{ij} - r_0)^2$$

$$\beta U_{ev}(r_{ij}) = \frac{u_{ij}}{(2\pi(a_i^2 + a_j^2))^{3/2}} e^{-r_{ij}^2/2(a_i^2 + a_j^2)}$$

$$\beta U_{el}(r_{ij}) = \frac{l_B \sigma_i \sigma_j}{r_{ij}} \text{erf} \left( \frac{r_{ij}}{2\sqrt{a_i^2/2 + a_j^2/2}} \right)$$

$a_i = 0.316 \text{ nm}$ : bead radius

$b$ : statistical segment length

$r_0 = 0$ : equilibrium bond distance

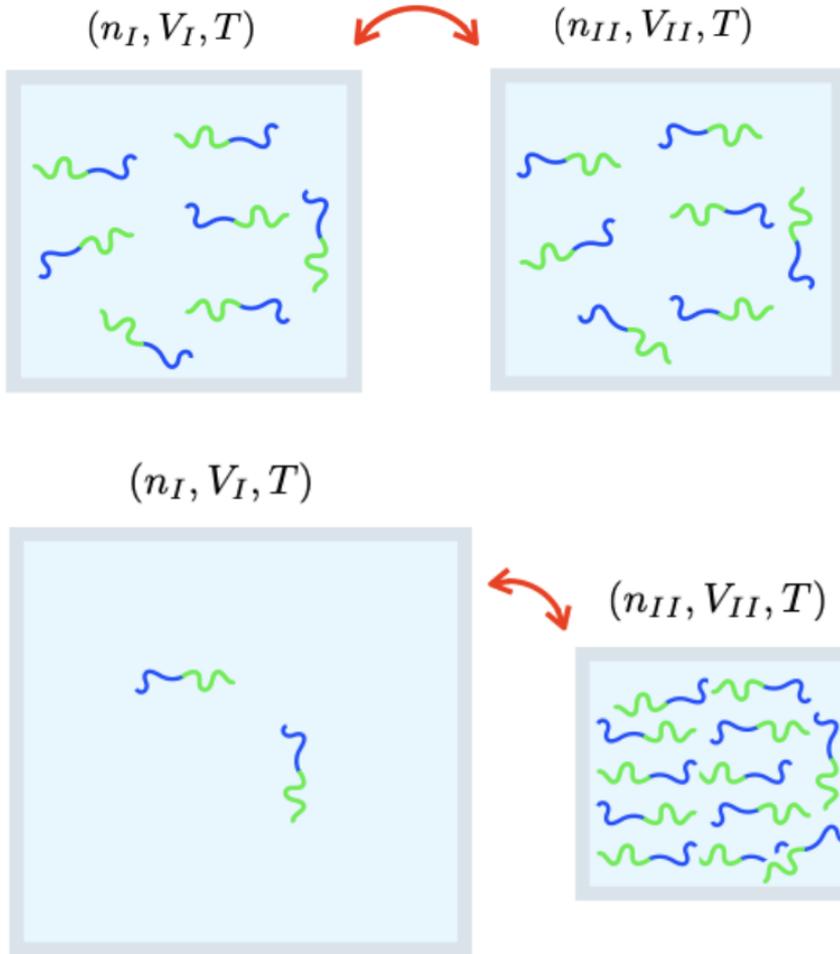
$u_{ij}$ : excluded volume

$\sigma_i$ : charge

$l_B = 561.6 \text{ nm}$ : Bjerrum length



# Gibbs Ensemble Simulations



$$V_T = V_I + V_{II} \quad n_T = n_I + n_{II}$$

$$F(n, V_T, T) = F_I(n_I, V_I, T) + F_{II}(n_{II}, V_{II}, T)$$

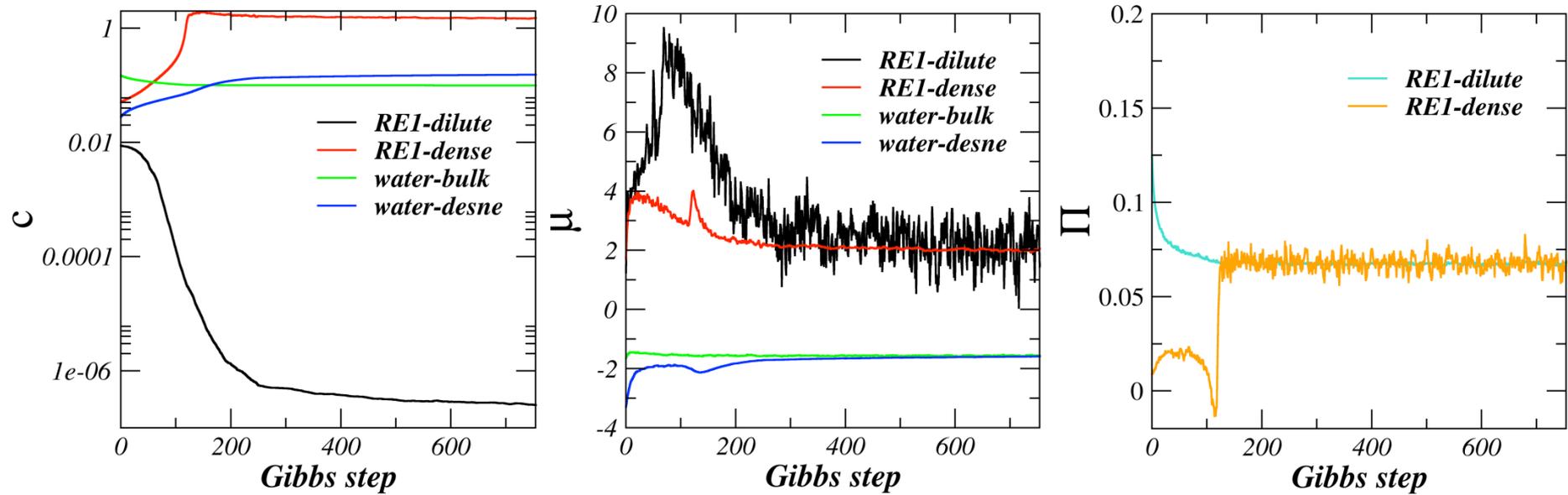
$$\frac{\partial F}{\partial V_I} = -(\Pi_I - \Pi_{II})$$

$$\frac{\partial F}{\partial n_I} = (\mu_I - \mu_{II})$$

$$\frac{\partial F}{\partial V_I} = -(\Pi_I - \Pi_{II}) = 0$$

$$\frac{\partial F}{\partial n_I} = (\mu_I - \mu_{II}) = 0$$

# Gibbs Ensemble Field Theory Simulation (FTS) convergence



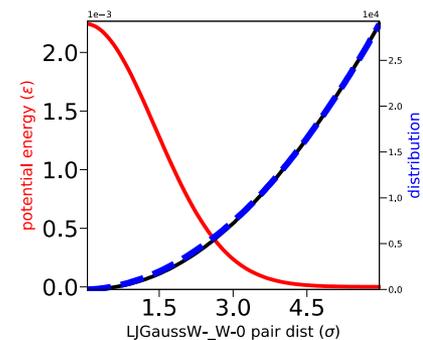
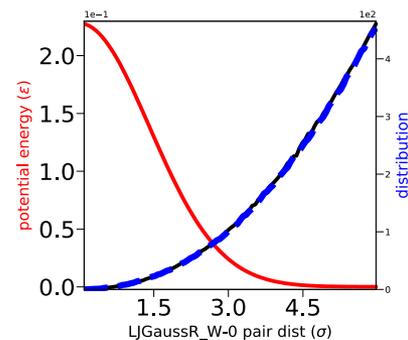
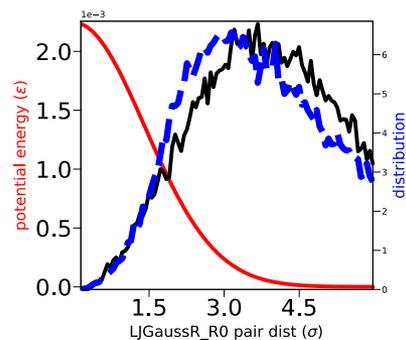
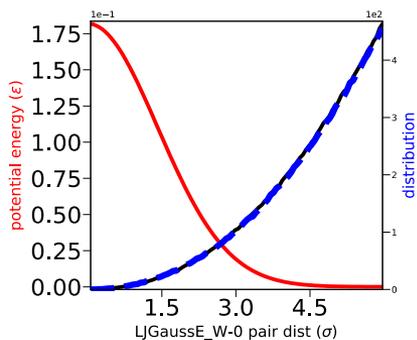
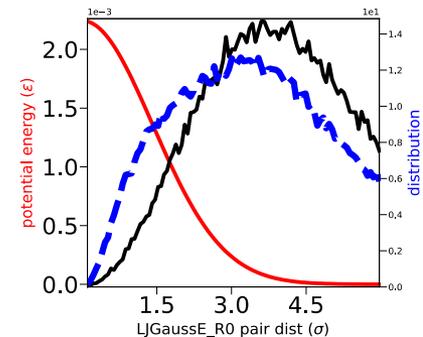
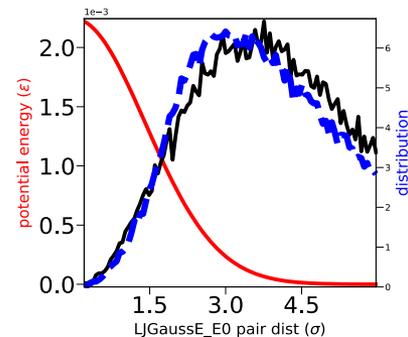
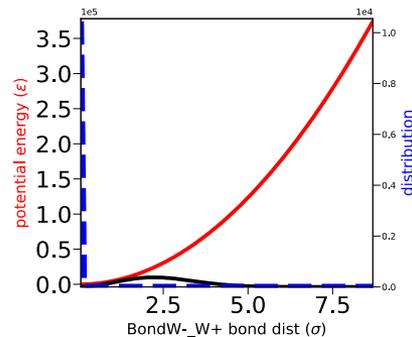
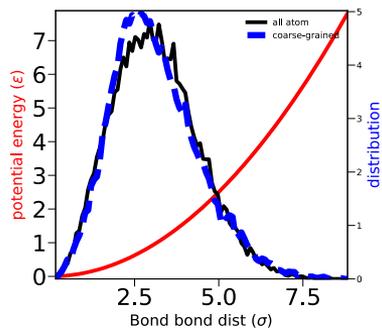
# CG parametrization of peptide-water interaction using relative-entropy

Optimized parameters:

Blurring sigma 0.35

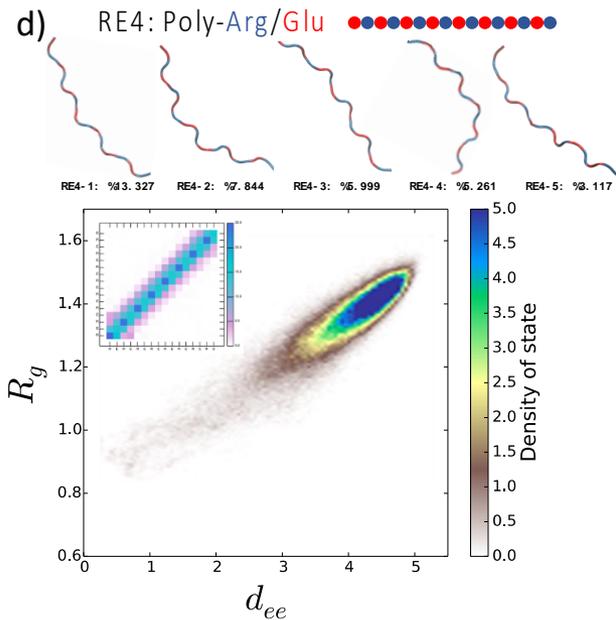
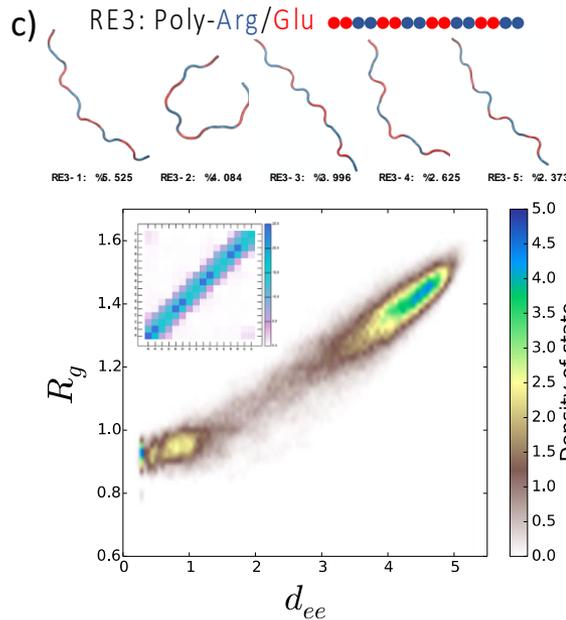
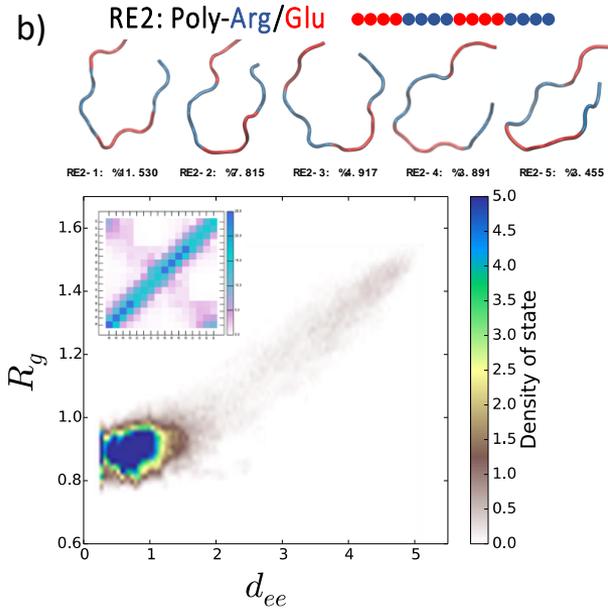
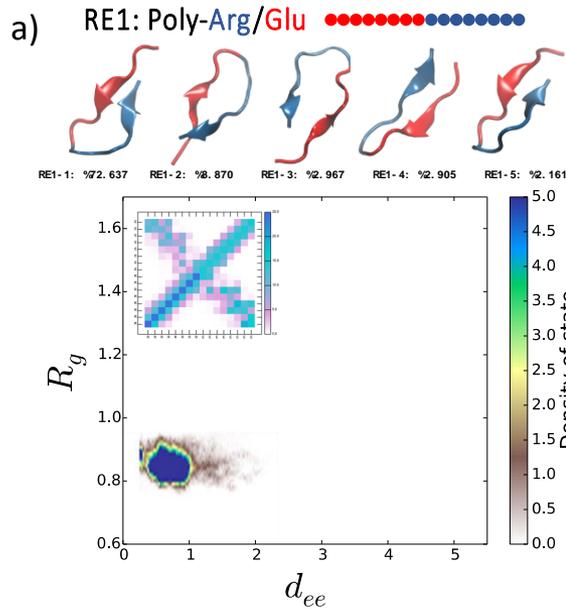
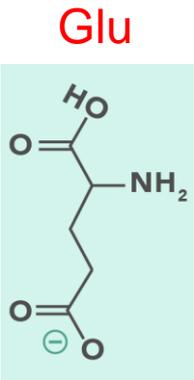
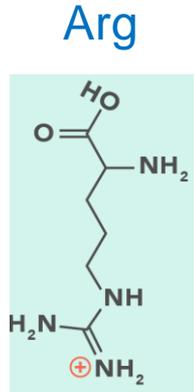
E-W B=0. 182

R-W B=0.228



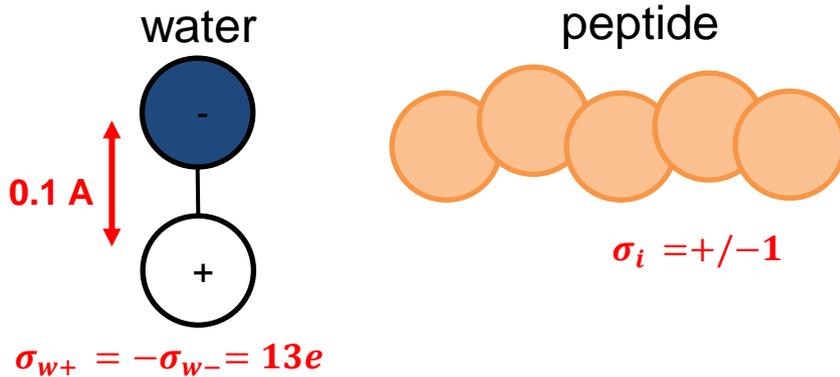
- The implicit solvent model cannot capture the heterogeneous dielectricity that may occur within the peptide-dense phase. In contrast, the explicit solvent model accounts for environment-dependent dielectricity, unlike the uniform dielectricity assumed in the implicit solvent model.
- This explicit solvent model can be parameterized to incorporate the effect of the peptide sequence on the effective interaction between residues and water into the coarse-graining (CG) process. Previous CG methods totally ignore this.
- As a result, the phase diagram derived from accurately transferring chemical details to the CG system is expected to capture any mesoscopic phase transitions and morphological complexities.
-

# All atom reference simulations for coarse graining



# Relative entropy parametrization of peptide and water interaction

RE1: Poly-Arg/Glu 



$$\sigma = a_i = R_w = 0.158 \text{ nm}$$

- $\beta U_{bond,water}$ : stiff harmonic with  $b = 0.01 \text{ nm}$
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- Excluded volumes:

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$a_i = 0.316 \text{ nm}$ : bead radius

$b$ : statistical segment length

$r_0 = 0$ : equilibrium bond distance

$u_{ij}$ : excluded volume

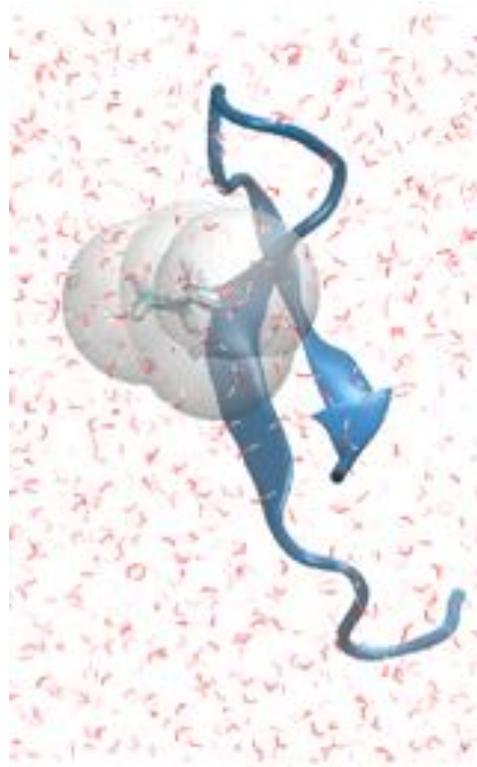
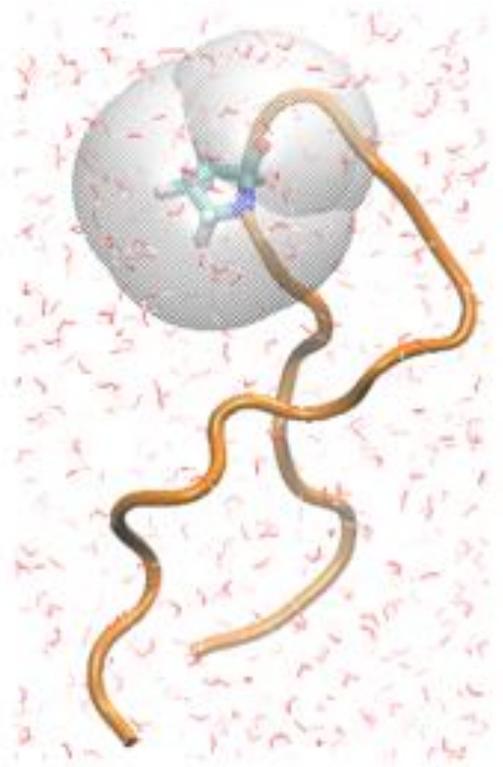
$\sigma_i$ : charge

$l_B = 561.6 \text{ nm}$ : Bjerrum length

# Tau and Traumatic Brain Injury



Overhauser Dynamic Nuclear Polarization (ODNP) experiments show a reduction in hydration water dynamics around 301 site for the P→L mutant (**jR2R3-P301L**)



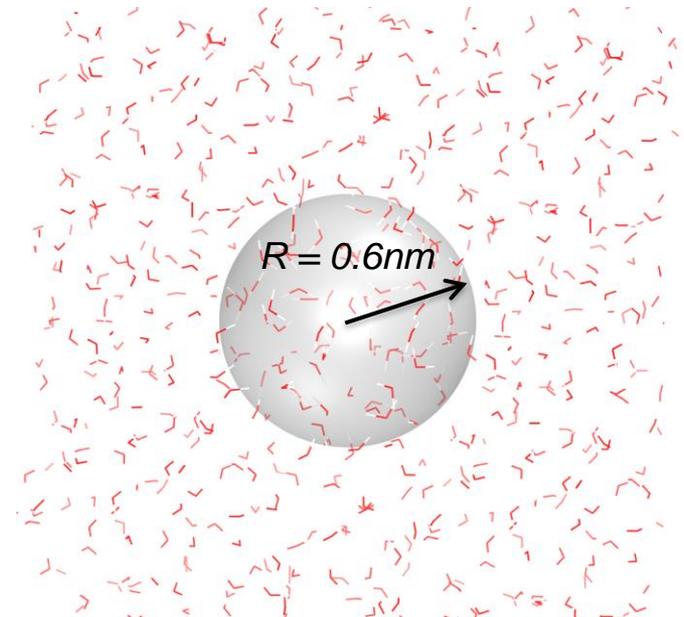
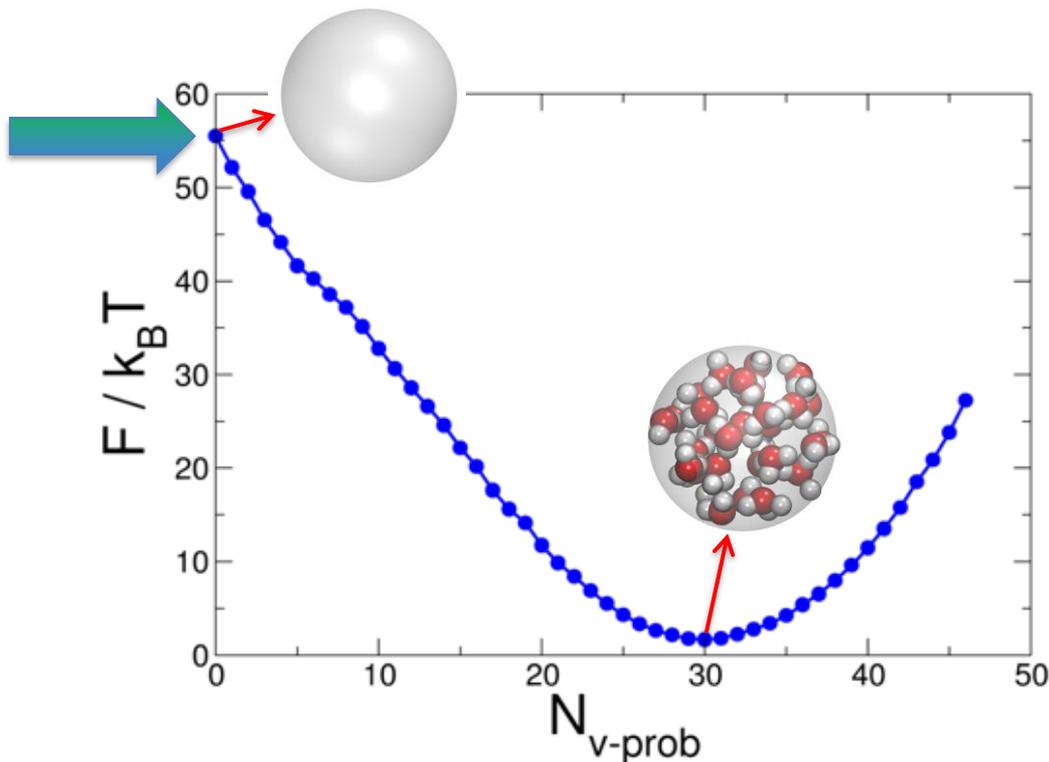
Increased ordering of water around mutation site

→ locally more hydrophobic

# Probing Hydrophobicity Computationally through Umbrella Sampling (INDUS)

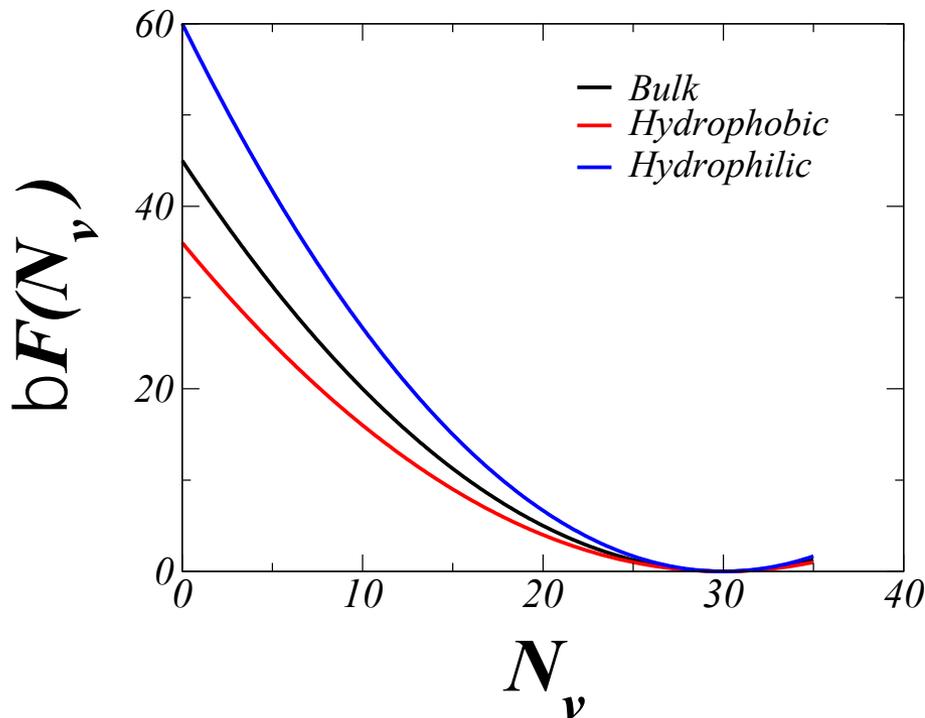
Free energy of dewetting a spherical volume in bulk

$$\mu_{\text{ex}} = \mathbf{F}(\mathbf{0}) = -\ln \mathbf{P}_{\mathbf{v}}(\mathbf{0})$$

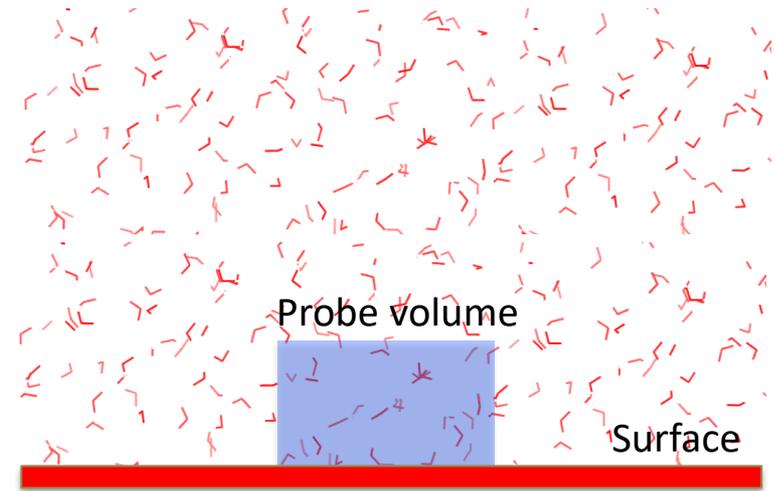


# Free energy of dewetting the probe volume in vicinity of a surface

Excess chemical potential is an indication of hydrophilicity or hydrophobicity of the surface

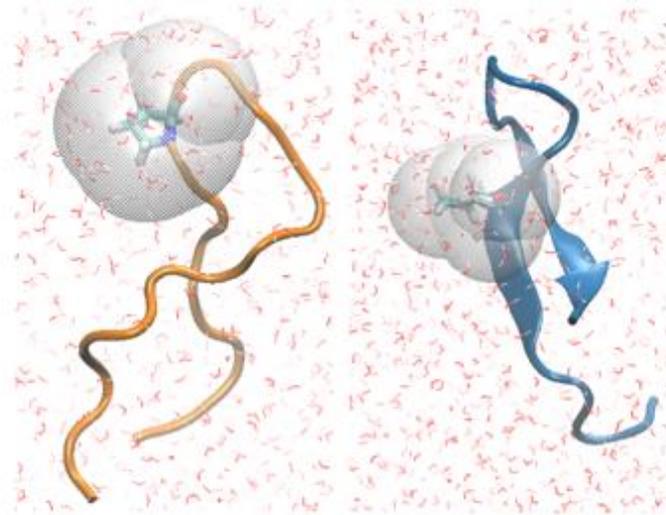


$$\mu_{\text{ex}}^{\text{phil}} > \mu_{\text{ex}}^{\text{bulk}} > \mu_{\text{ex}}^{\text{phob}}$$



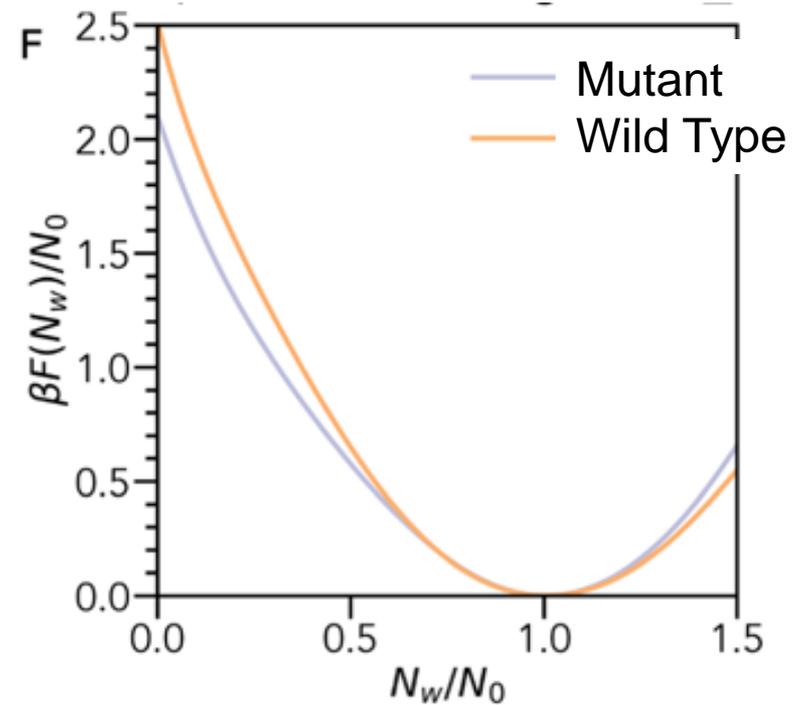
$$\mu_{\text{ex}} = \mathbf{F}(0) = -\ln P_v(0)$$

# Free energy of dewetting lower for jR2R3-P301L: an additional factor favoring association of jR2R3-P301L

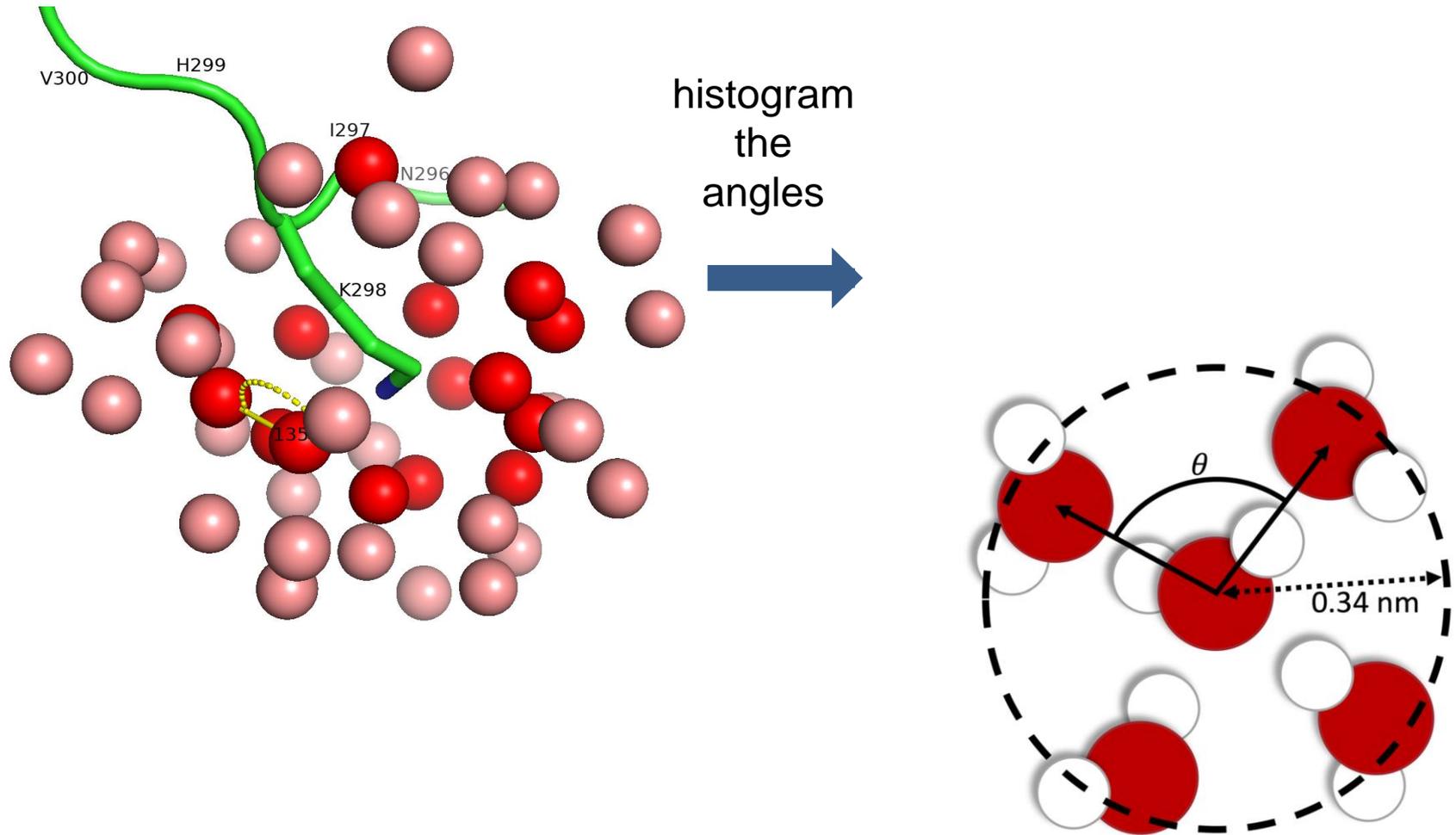


Wild Type

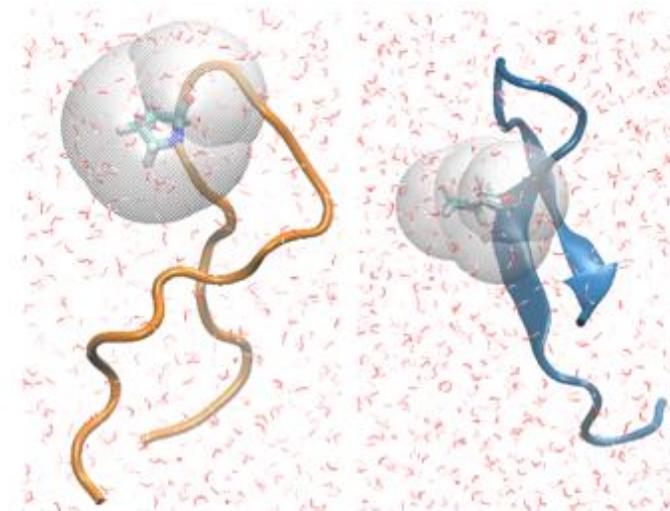
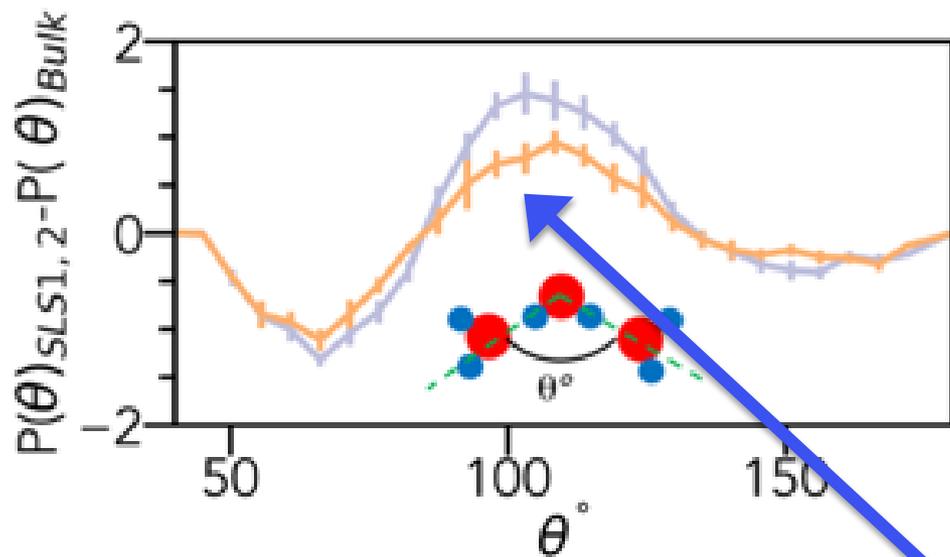
Mutant



# Quantifying Water Structure: *water triplet distribution*



Increased tetrahedral ordering of hydration waters near the L301 (mutant) compared to P301 (wild type)



Wild Type

Mutant

TETRAHEDRAL  
“hydrophobic”

# Dimer Simulations of jR2R3 and JR2R3-P301L



jR2R3

6.4%



6.1%



5.6%

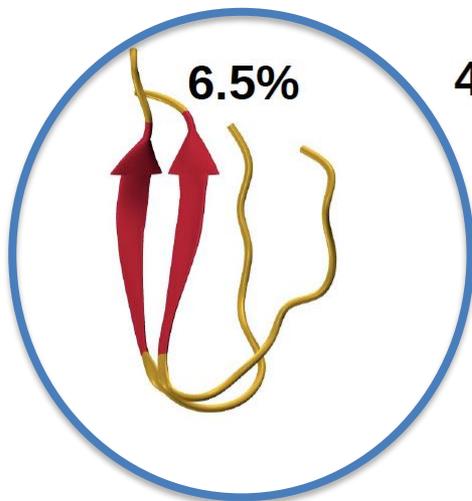


2.9%



jR2R3-  
P301L

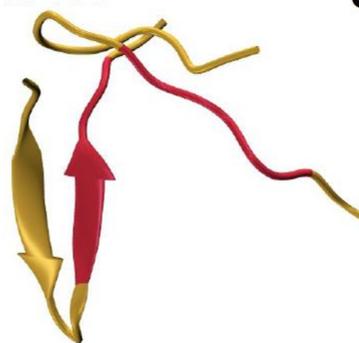
6.5%



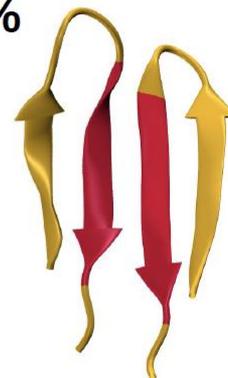
4.8%



4.4%

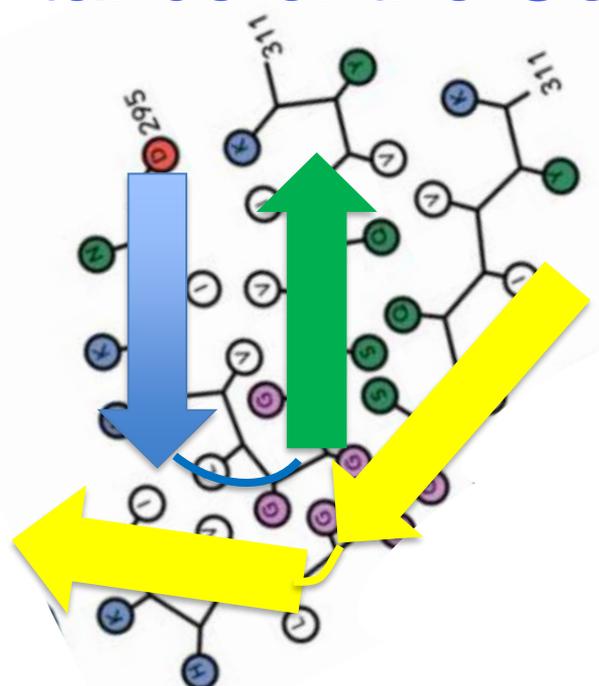
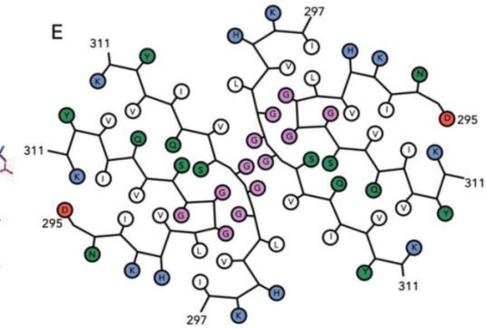
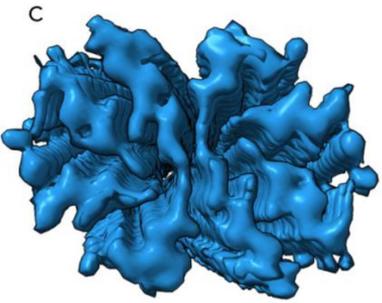


3.5%

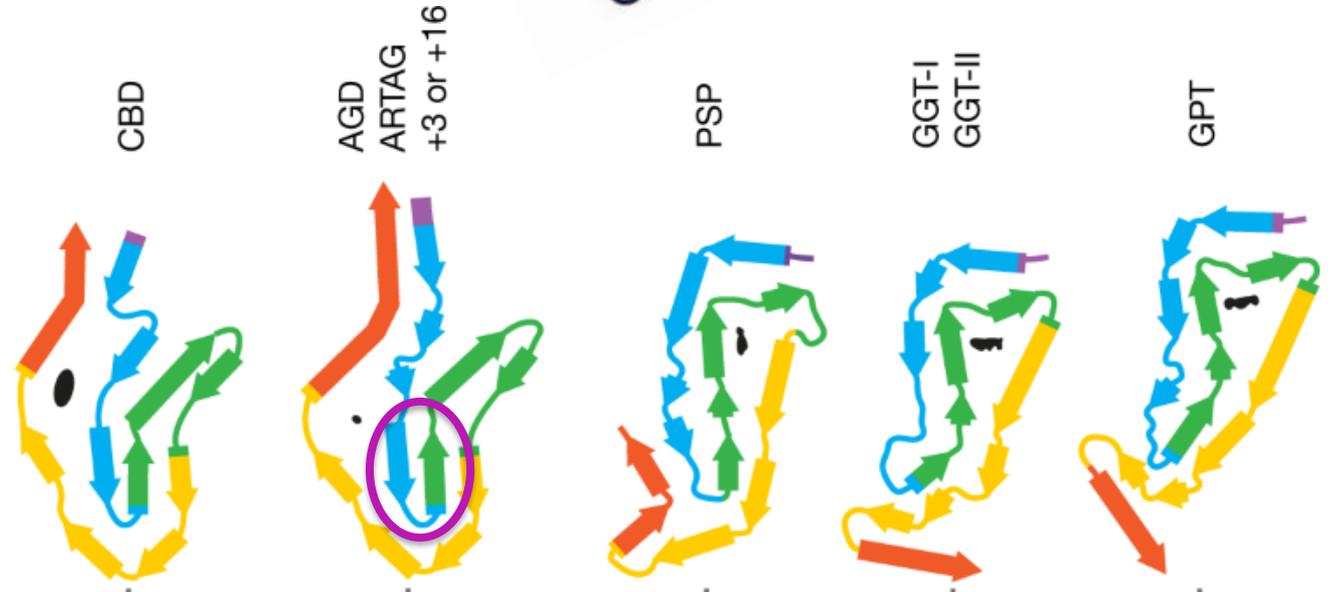




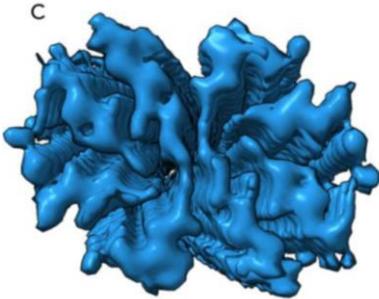
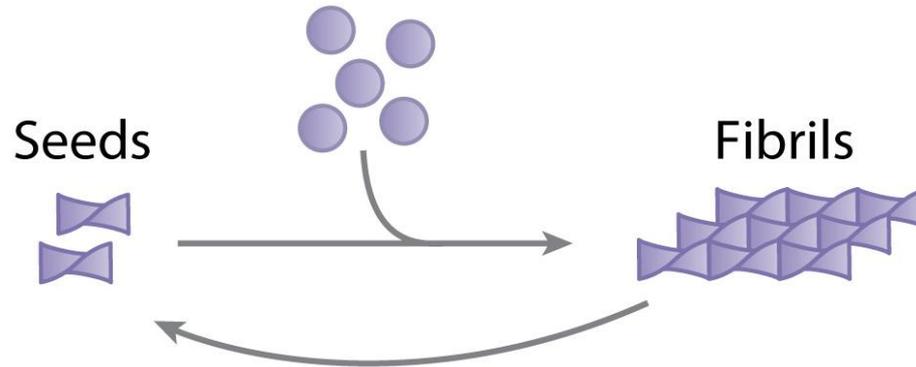
# Importance of the Counter-Strand



jR2R3-P301L

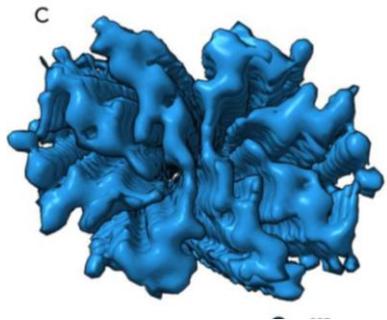


# Seeding



Can jR2R3-P301L can seed the fibrillization of full length Tau?

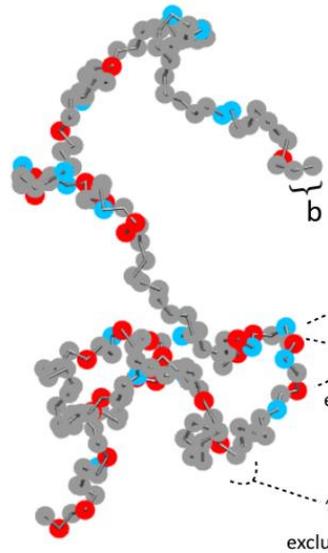
# jR2R3-P301L can seed the fibrillization of full length Tau in Vitro



Fibril of Tau  
Fragment  
jR2R3-P301L

DNIKHV**L**GGS  
VQIVY**K**

+



Full length Tau

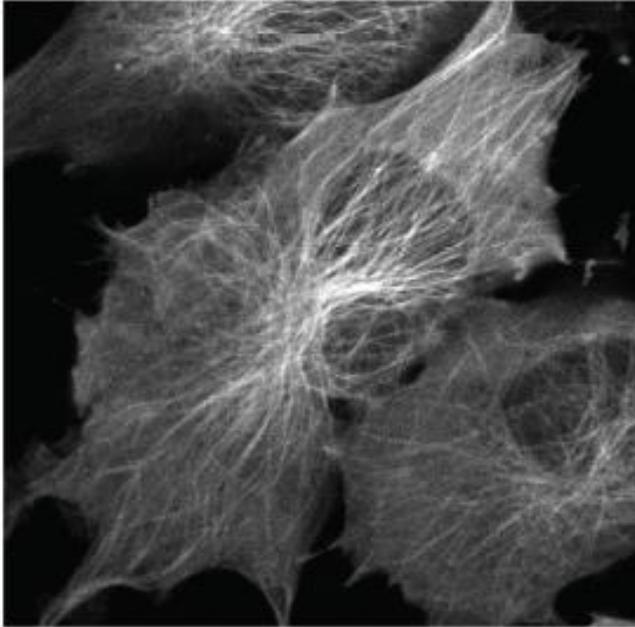


Full Length Tau  
Fibrils

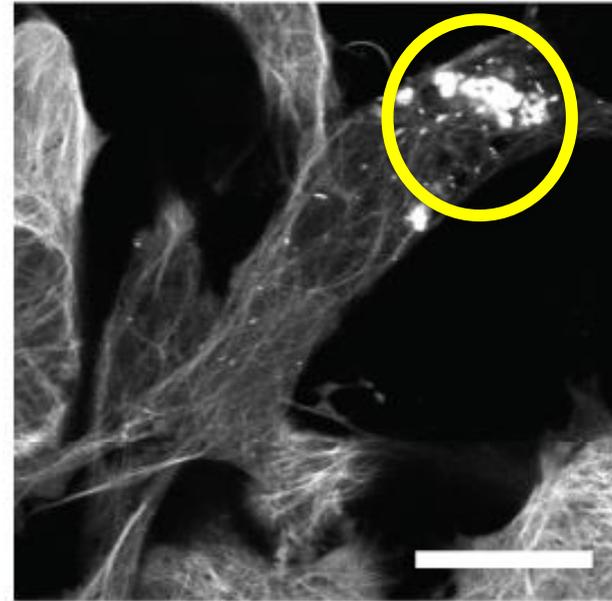
	272	GGK	274	R1	
275	VQI	IINKKLDLS-NVQSKCGSKDNIKHVPGGGS	305	R2	
306	VQIVYK	PVDLS-KVTSKCGSLGNIHHKPGGGQ	336	R3	
337	VEVKSEK	LDLDFKDRVQSKIGSLDNITHVPGGGN	368	R4	
369	KKIETHK	LTFR	NAKAKTD	387	C

# jR2R3-P301L can seed the fibrillization of full length Tau in Vivo

Before jR2R3 P301L

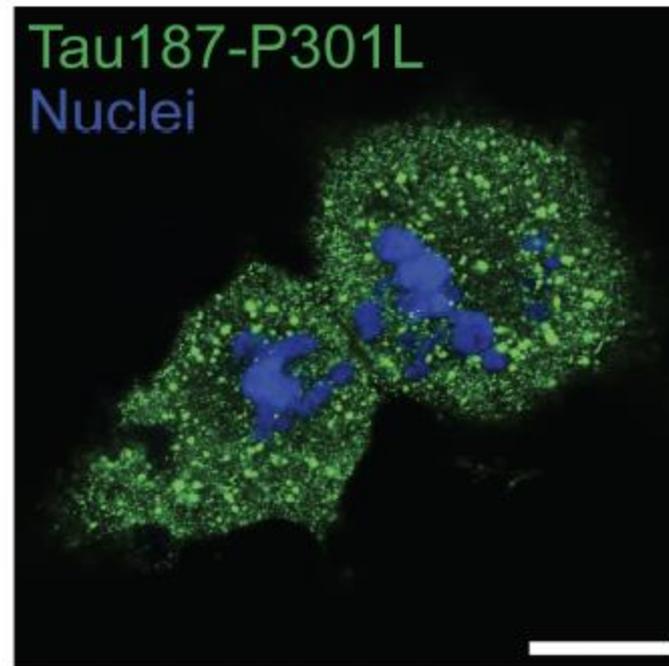


After jR2R3 P301L



Cells expressing mClover3-Tau187-P301L seeded with jR2R3-P301L fibrils

# jR2R3-P301L acts as a prion: propagates the strain

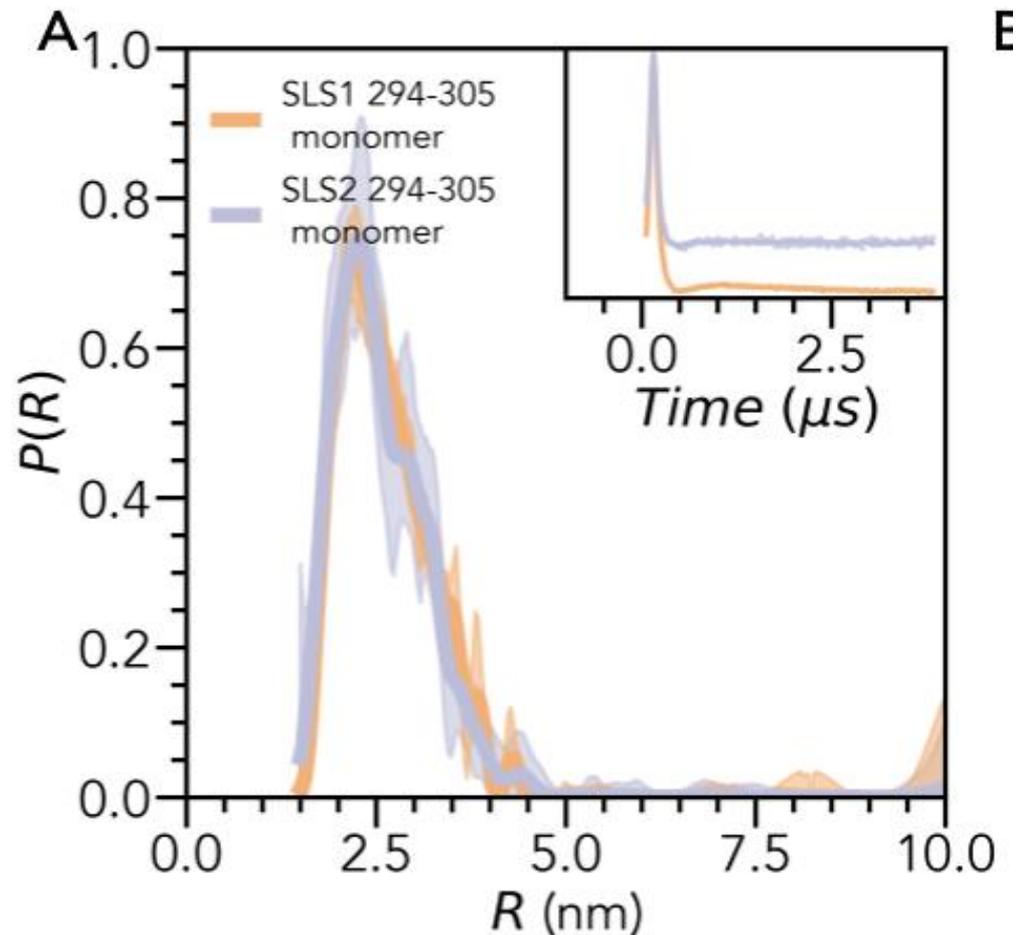


**Prof. Ken Kosik, UCSB**

Cells seeded with jR2R3-P301L fibrils undergo division and propagate aggregates to daughter cells

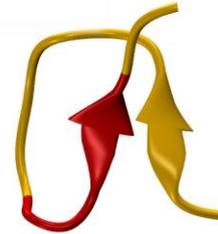
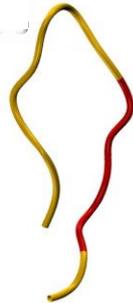


# Double Electron-Electron Resonance (DEER) Spectroscopy cannot distinguish between wild type and mutant



# Simulations of jR2R3 and JR2R3-P301L (Replica Exchange; Charmm36m)

jR2R3



jR2R3-P301L

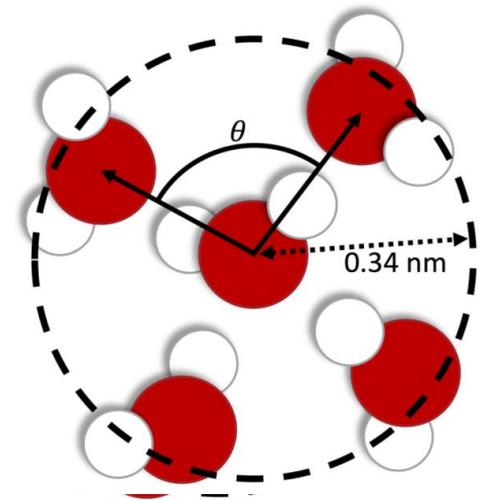
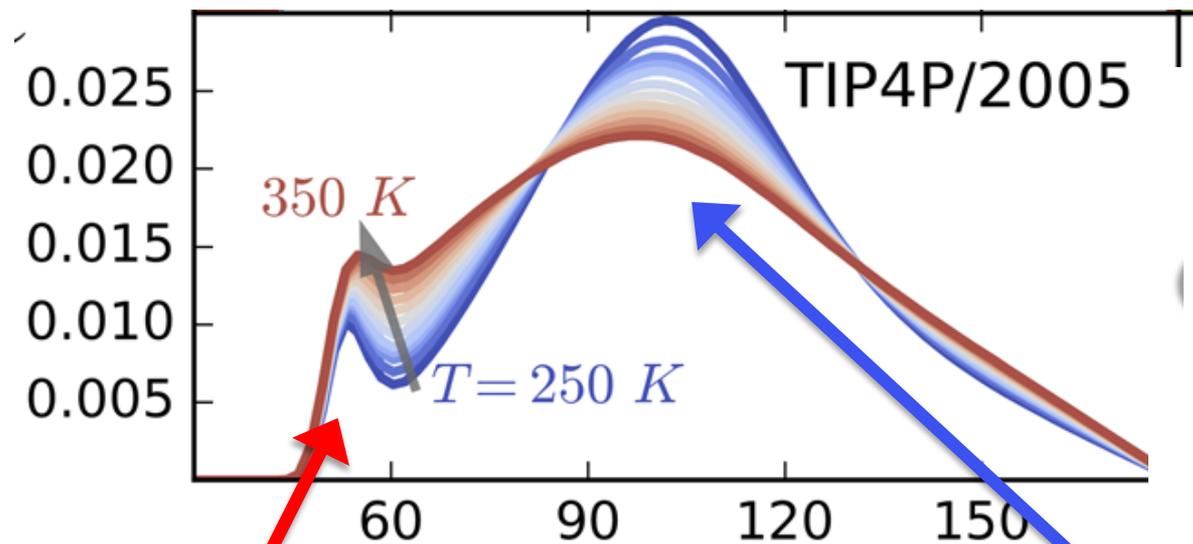


**LITTLE DIFFERENCE**



**Pritam Ganguly**

# Water Triplet Distribution



$P(\theta)$

$350\text{ K}$

$T = 250\text{ K}$

TIP4P/2005

3-body angle  $\theta$

“hydrophilic”

TETRAHEDRAL  
“hydrophobic”