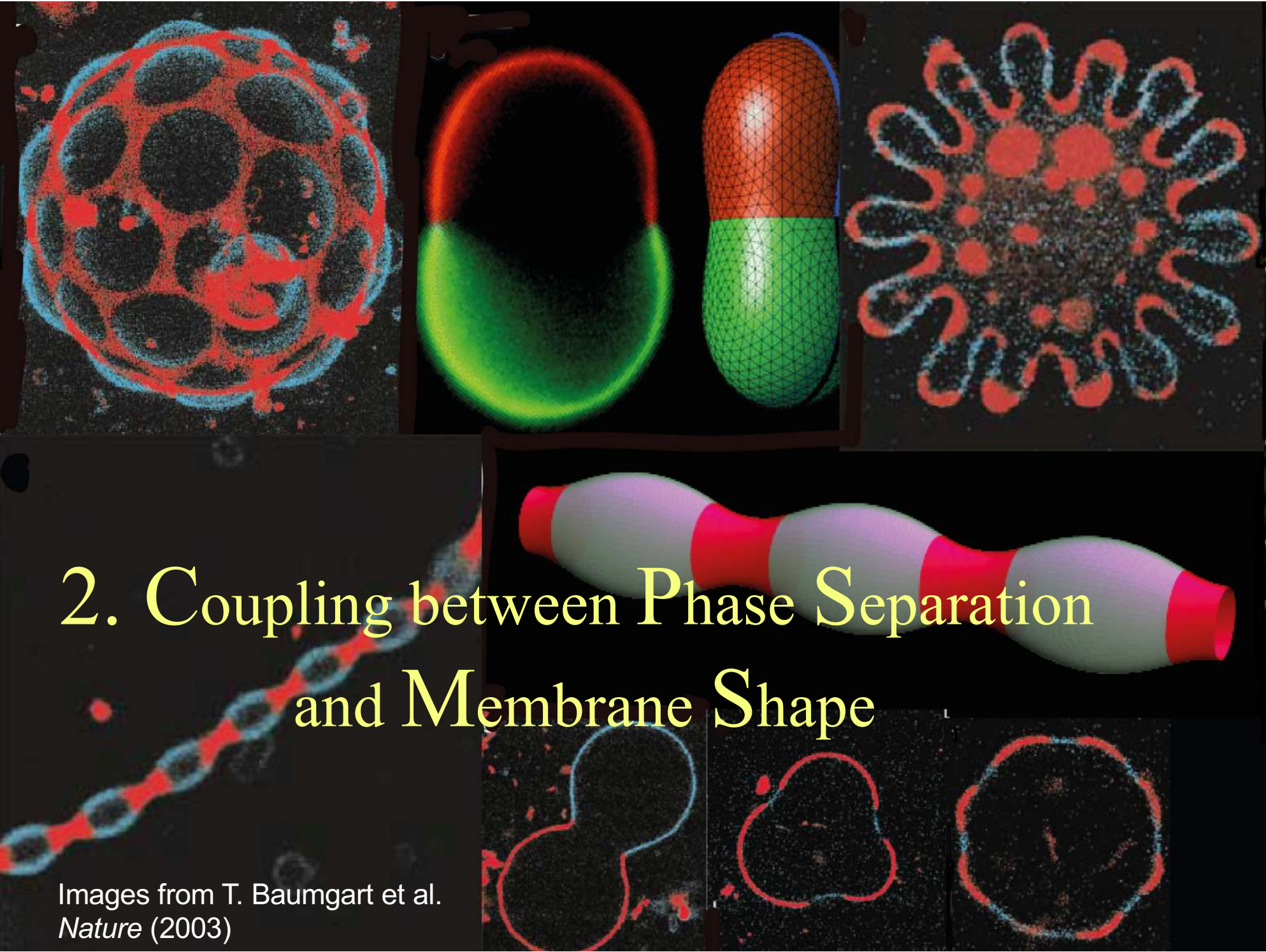




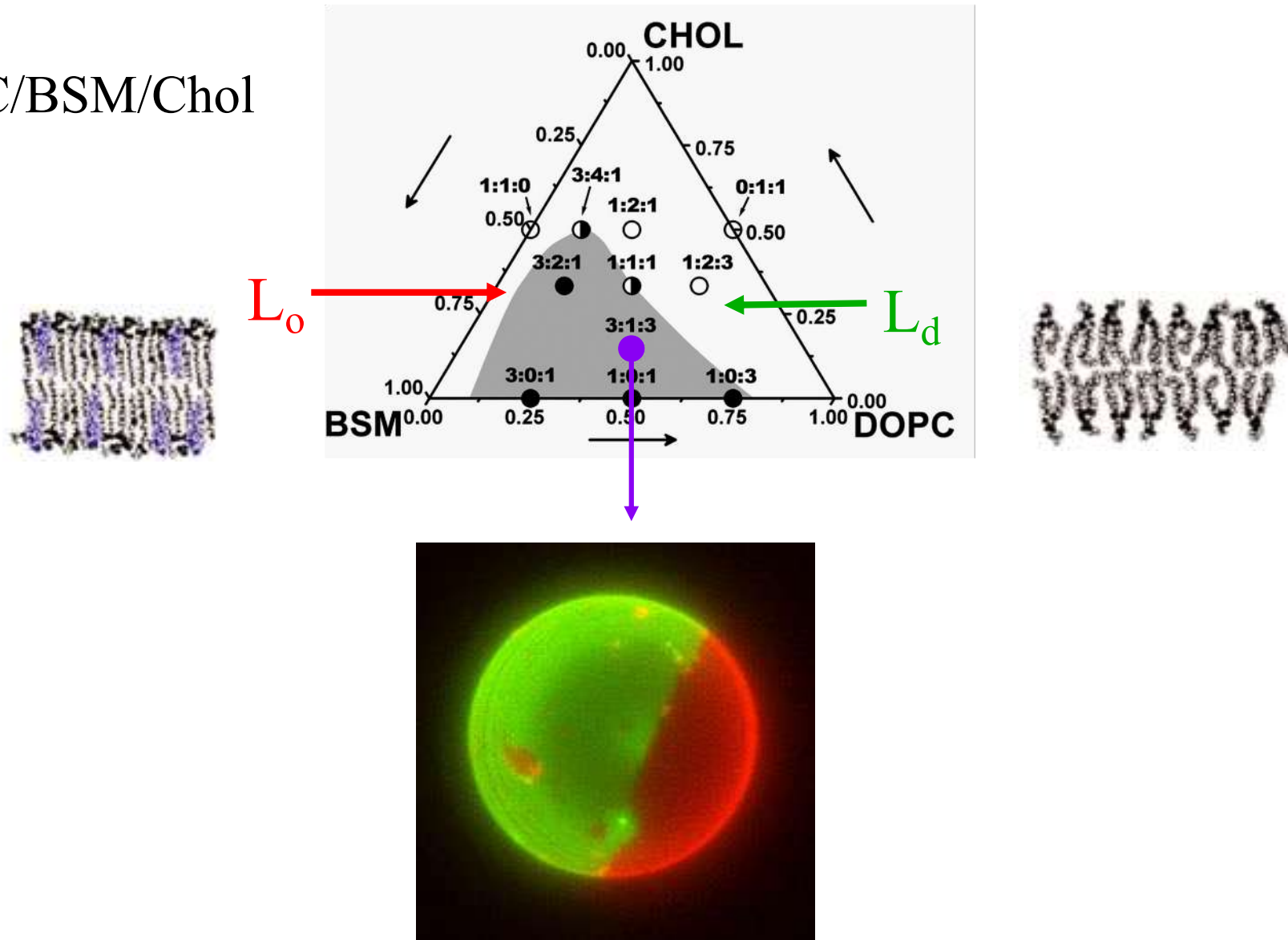
2. Coupling between Phase Separation and Membrane Shape

Images from T. Baumgart et al.
Nature (2003)

Membrane Deformation
due to Line Tension
between Lipid Domains

Fluid Phases Coexistence in Lipid Mixtures

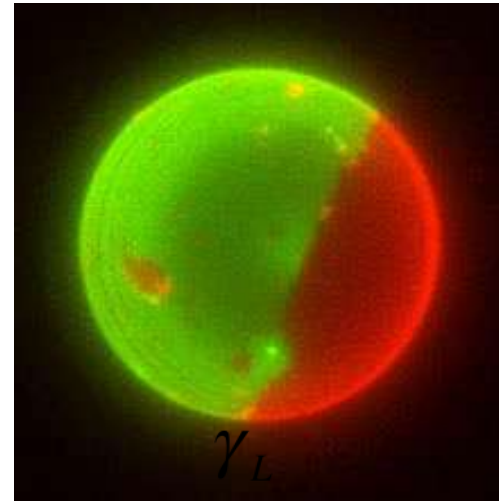
DOPC/BSM/Chol



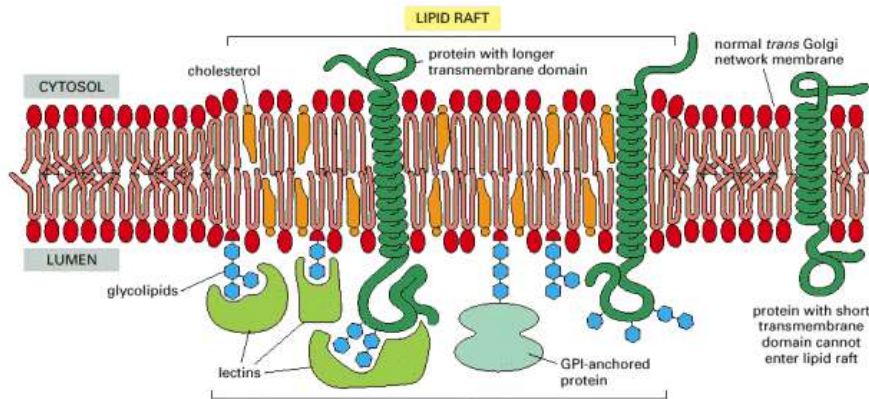
2D Phase Separation



At 3D :
Interfacial tension

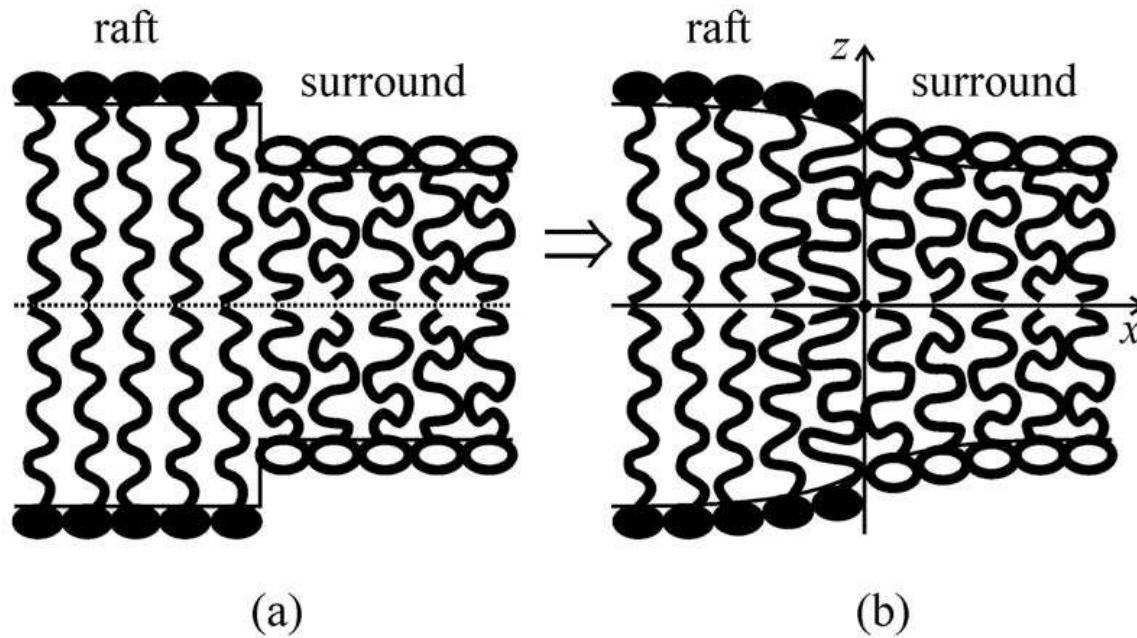


On a membrane (2D):
Line tension ≈ 1 to few pN
(depends on composition, temperature)

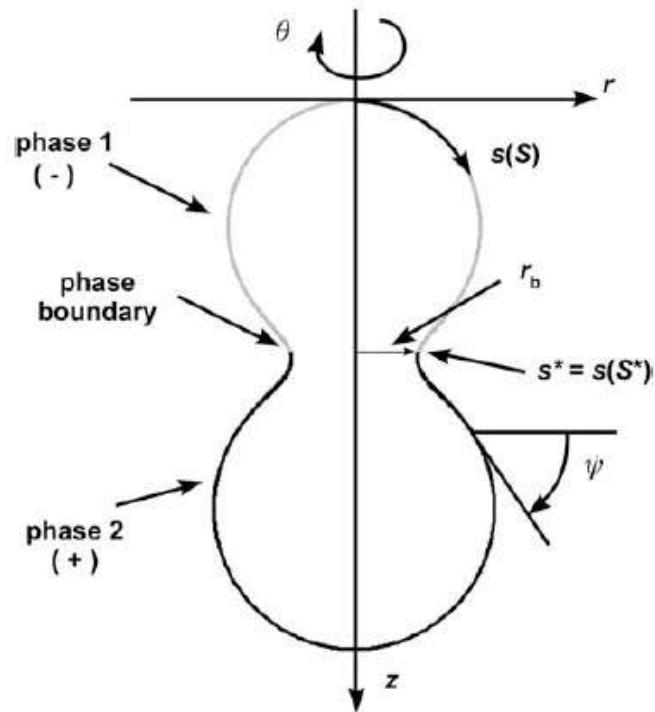


Coexistence of 2 phases

⇒ *Line tension*



Example of a Measure of Line Tension

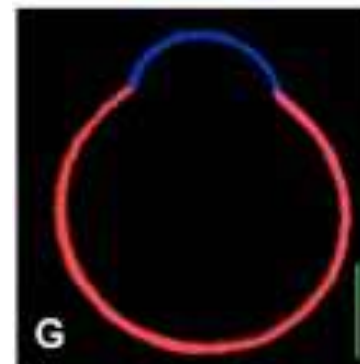
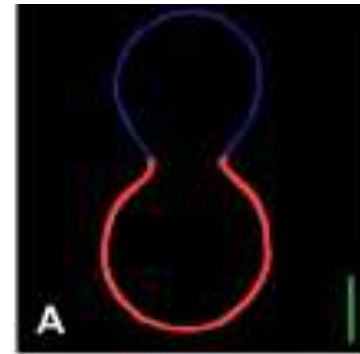


For symmetric doublets

$$\frac{\gamma_L}{p} = \frac{1}{2} r_b^2 (\cot \psi_1 - \cot \psi_2)$$

r_b : neck radius

p : pressure difference (Lapalce)



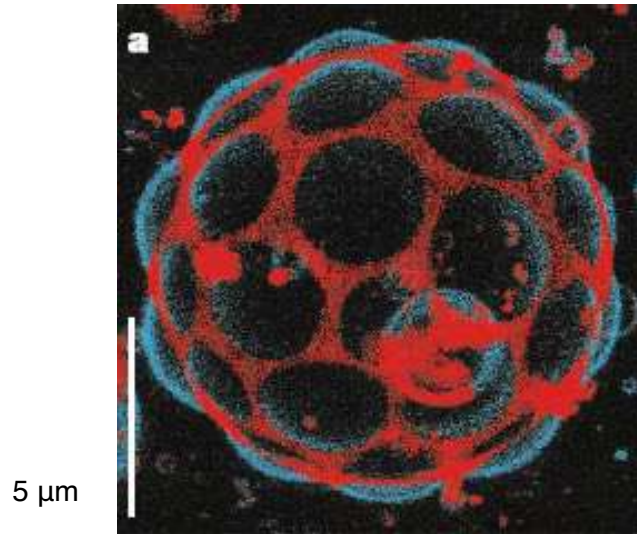
$\gamma_L \approx 1$ to few pN

T. Baumgart... J.Jenkins
Biophys. J. (2005)

5 μ m

Pure Lipid Domains can Form Vesicles

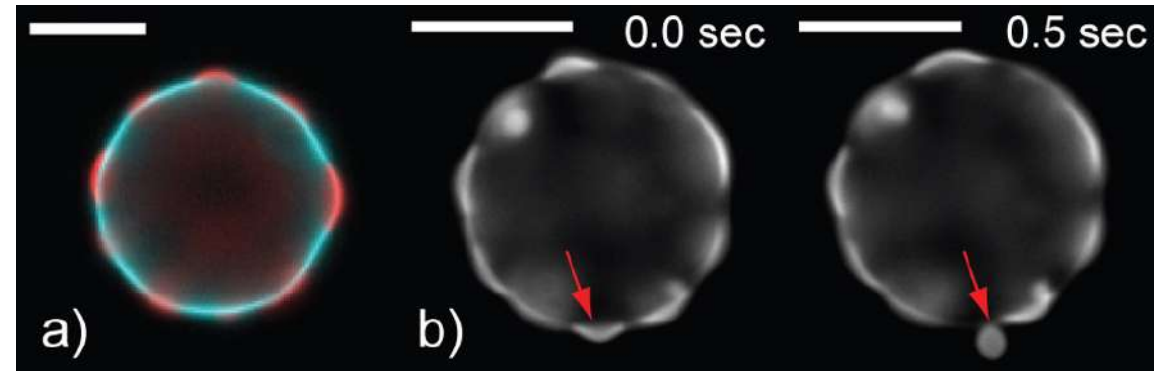
Tension lowered



5 μm

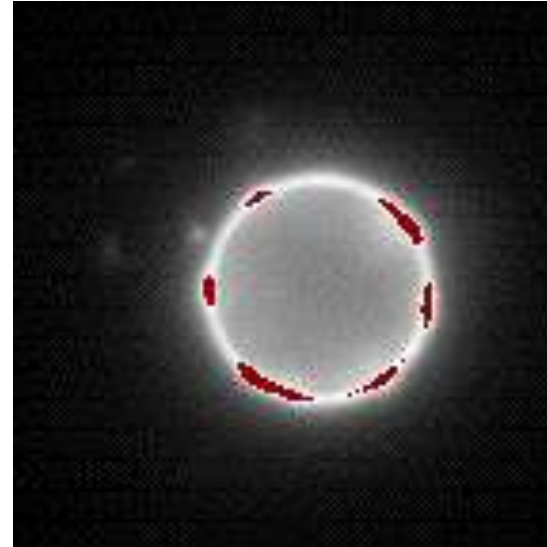
T. Baumgart et al *Nature* **425**, 821 (2003)

Ex: DOPC/Chol/SM



J. E. Rim, et al *PRL* **106**, 057801 (2011)

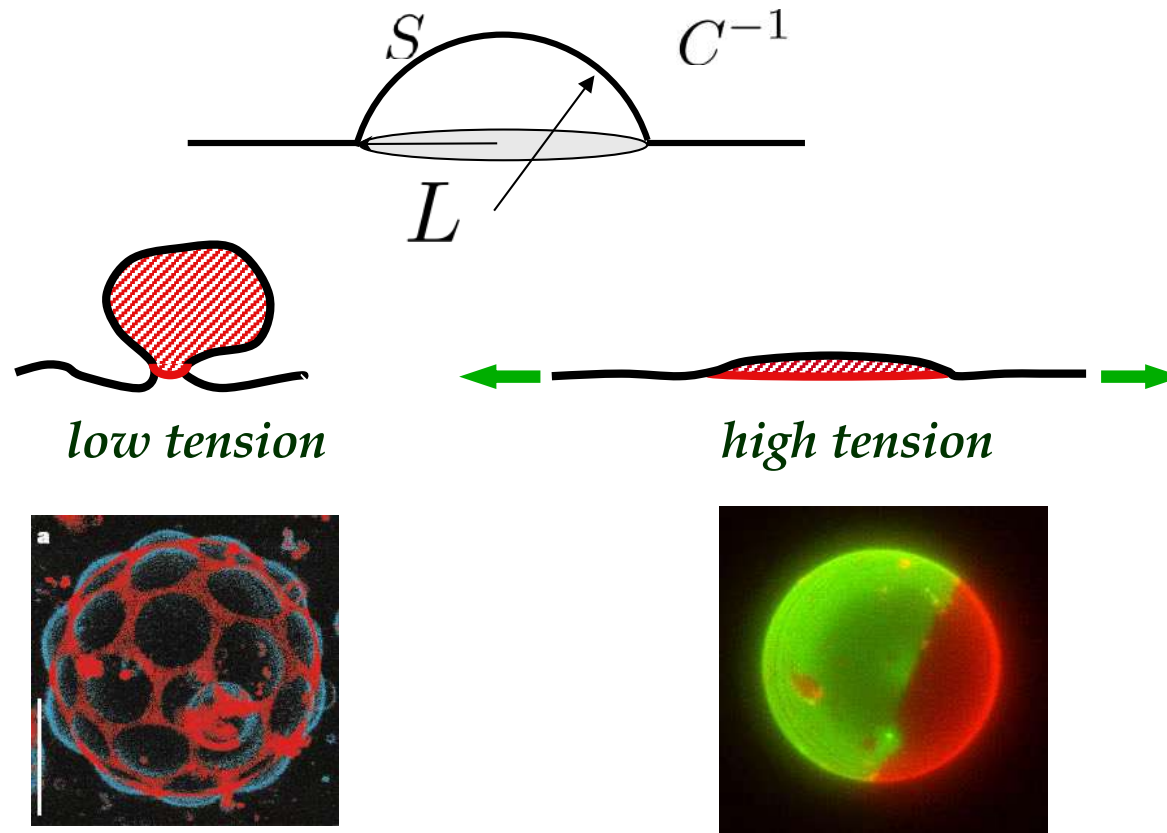
Bars, 10 μm



A. Roux et al.,
EMBO J. **24**, 1537 (2005)

With pure lipid domains: *micron-size vesicles*

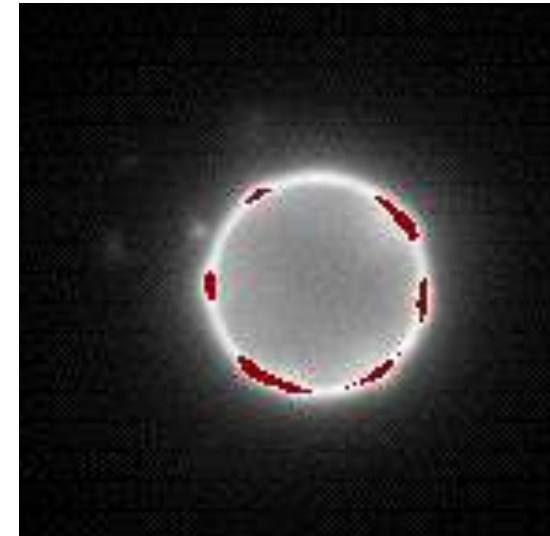
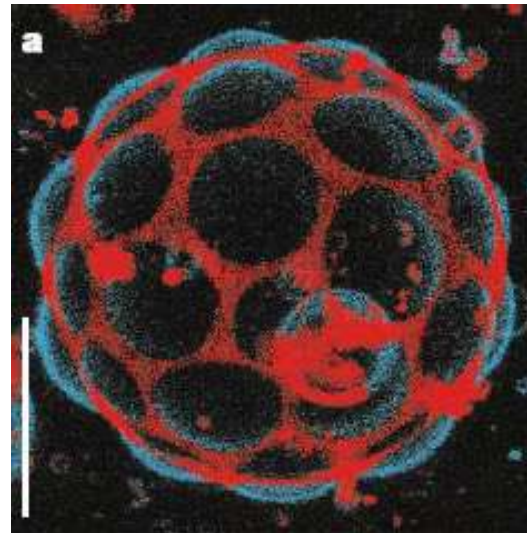
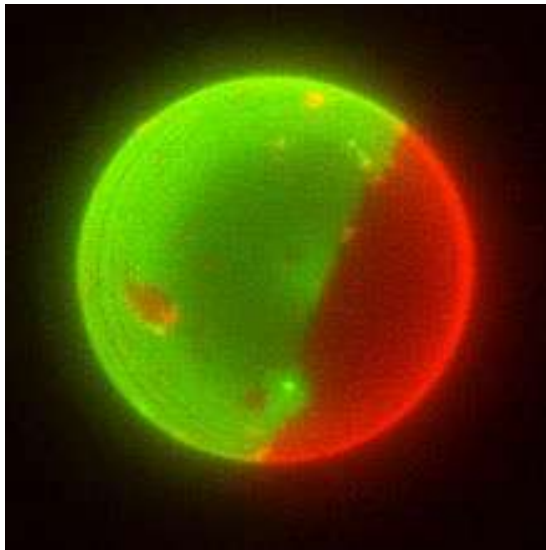
Competition Line Tension/Surface Tension



$$F = \underbrace{\gamma_L 2\pi L}_{\text{Line Tension}} + \underbrace{\frac{1}{2} \kappa S (C - C_0)^2}_{\text{Curvature}} + \underbrace{\sigma (S - \pi L^2)}_{\text{Surface Tension}}$$

Effect of Membrane Tension

Tension



Baumgart et al, Nature (2003)

Full separation

Budding

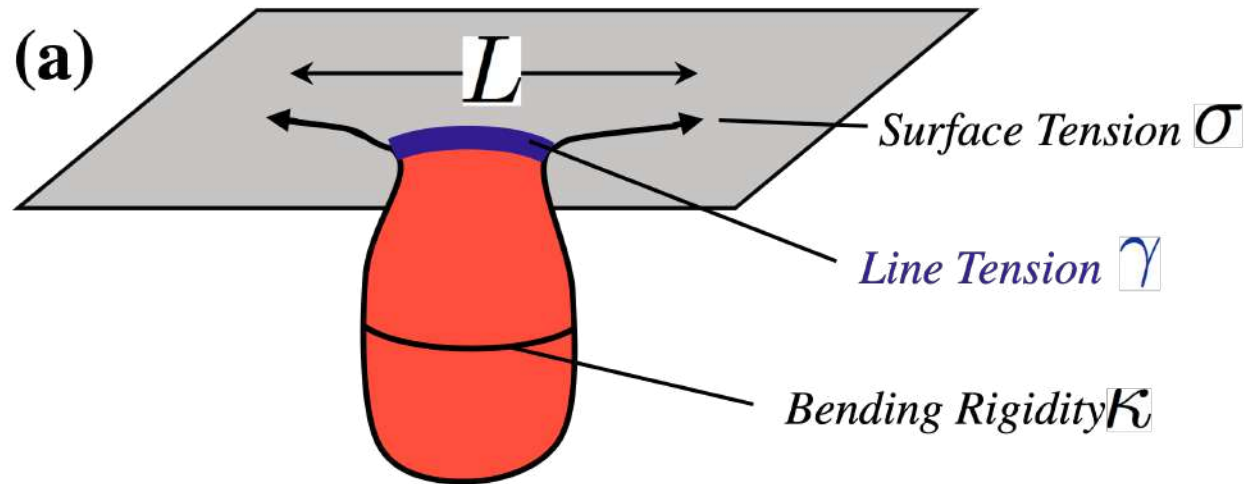
Constriction and **Fission**

$$F = \boxed{\gamma_L 2\pi L} + \frac{1}{2} \kappa S (C - C_0)^2 + \sigma (S - \pi L^2)$$

P. Sens, M.S. Turner *Phys. Rev. E* (2006)

Pure Lipid Domains Form *Large* Buds

Bud formation



Bending energy/ vesicle $F = 8\pi\kappa$ Size independent

Line tension γ

$F = 2\pi L\gamma$ $\gamma \approx 1$ to a few pN

If pure lipids and zero tension, **smallest domain size**:

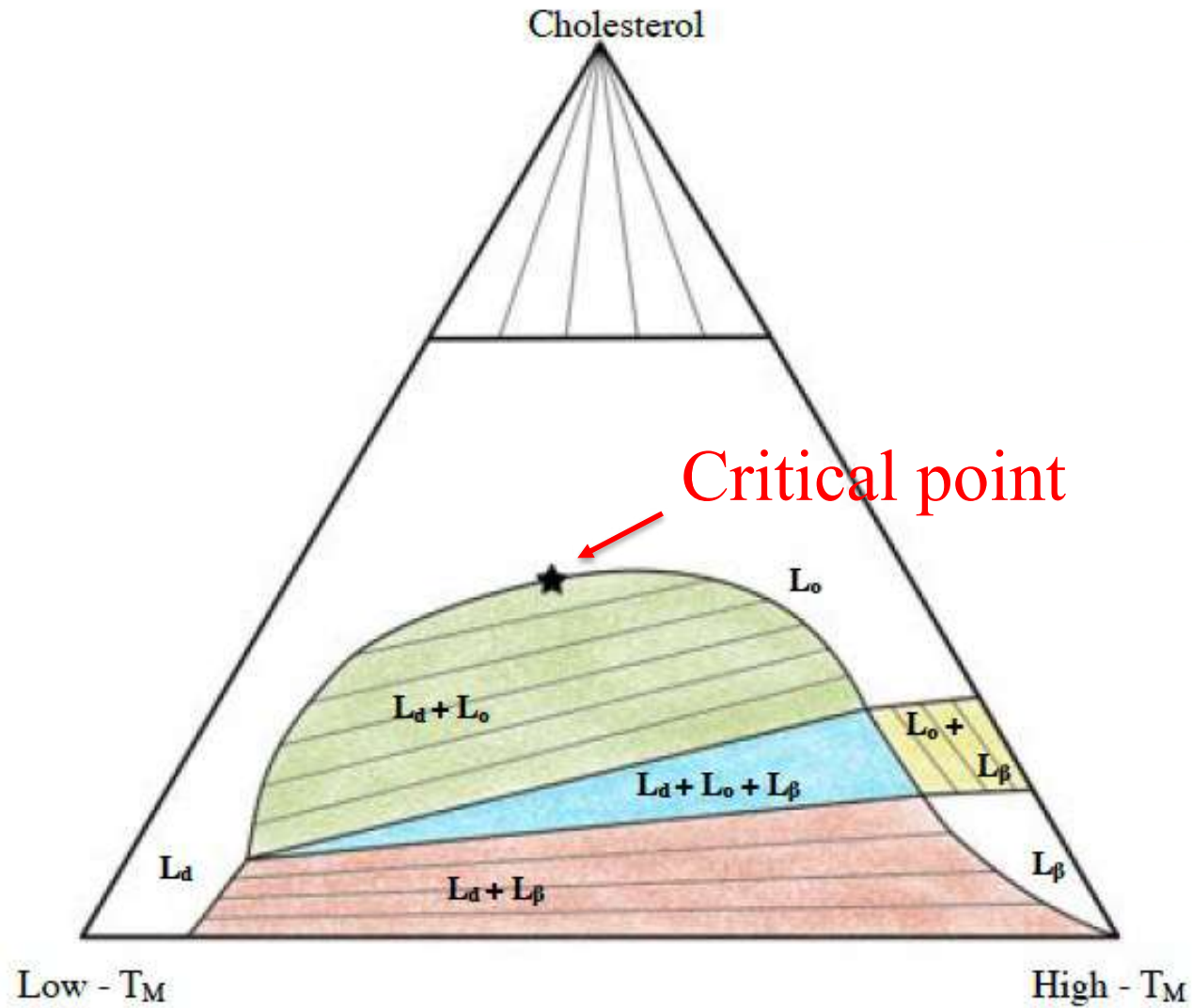
$$L \approx \frac{4\kappa}{\gamma} \approx 300nm$$

Increases with membrane tension

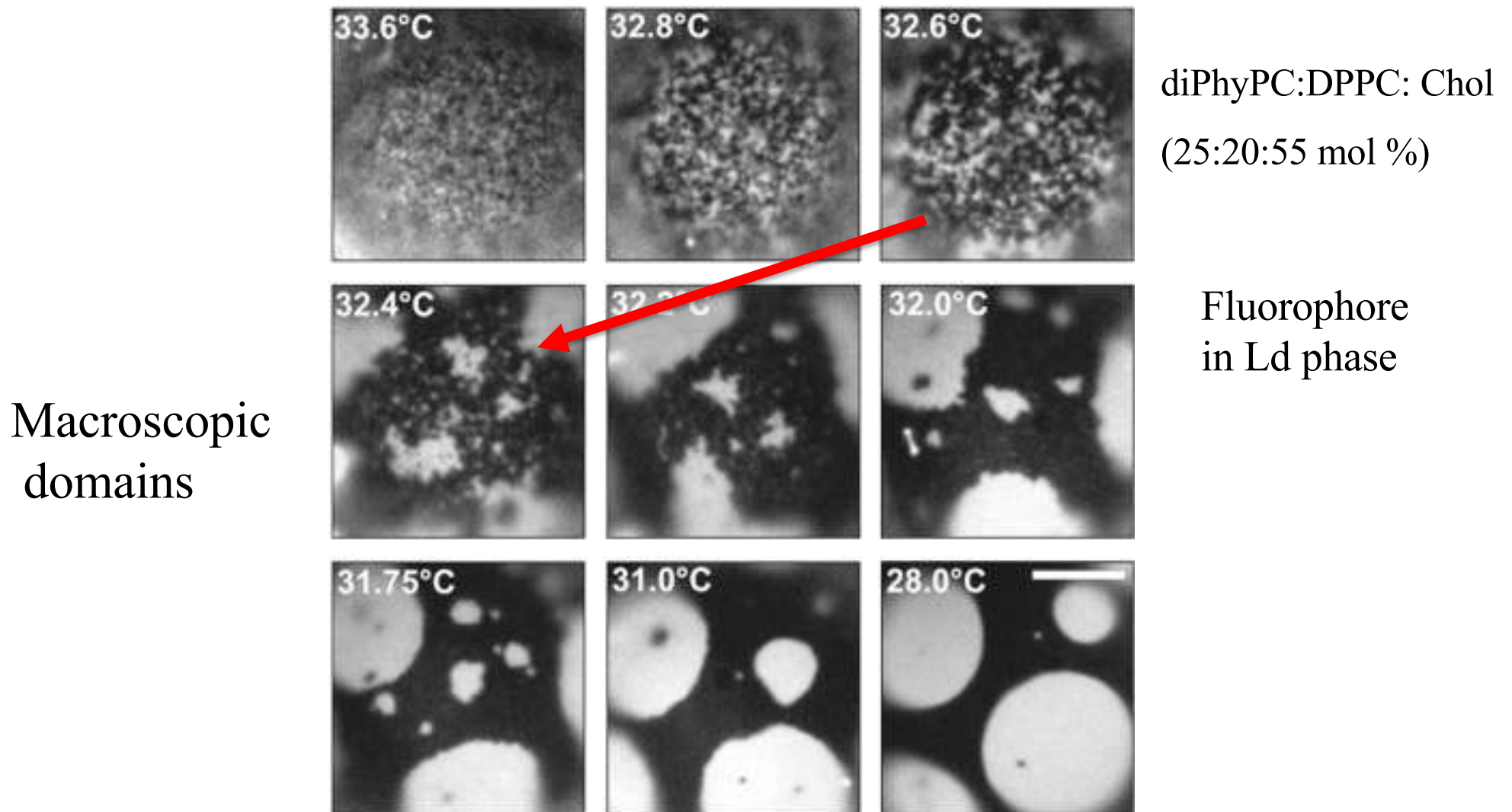
P. Sens et al *Curr. Opin. Cell Biol.*
(2008)

Proteins required to form small vesicles (or narrow tubes)

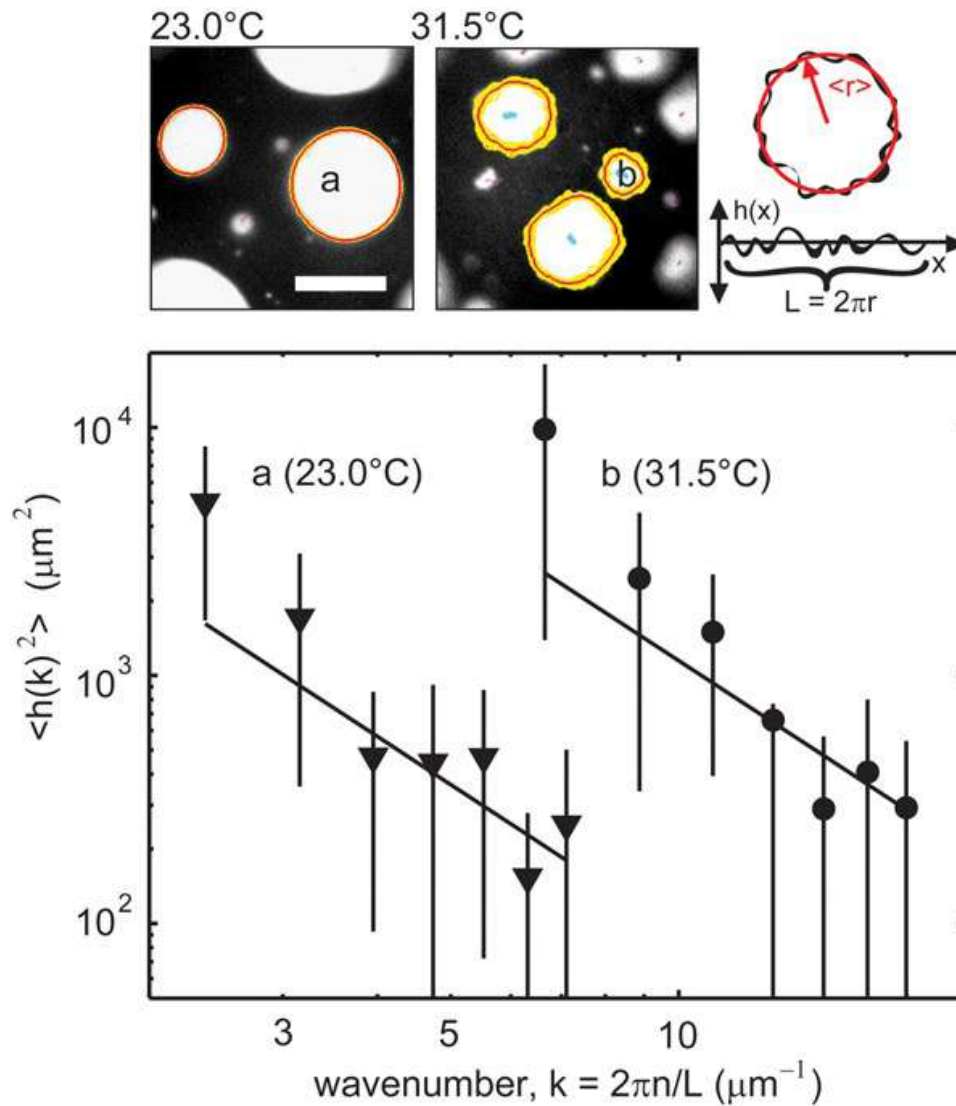
Phases Coexistence in Ternary Lipid Mixtures



Cooling a Mixture near a Critical Point



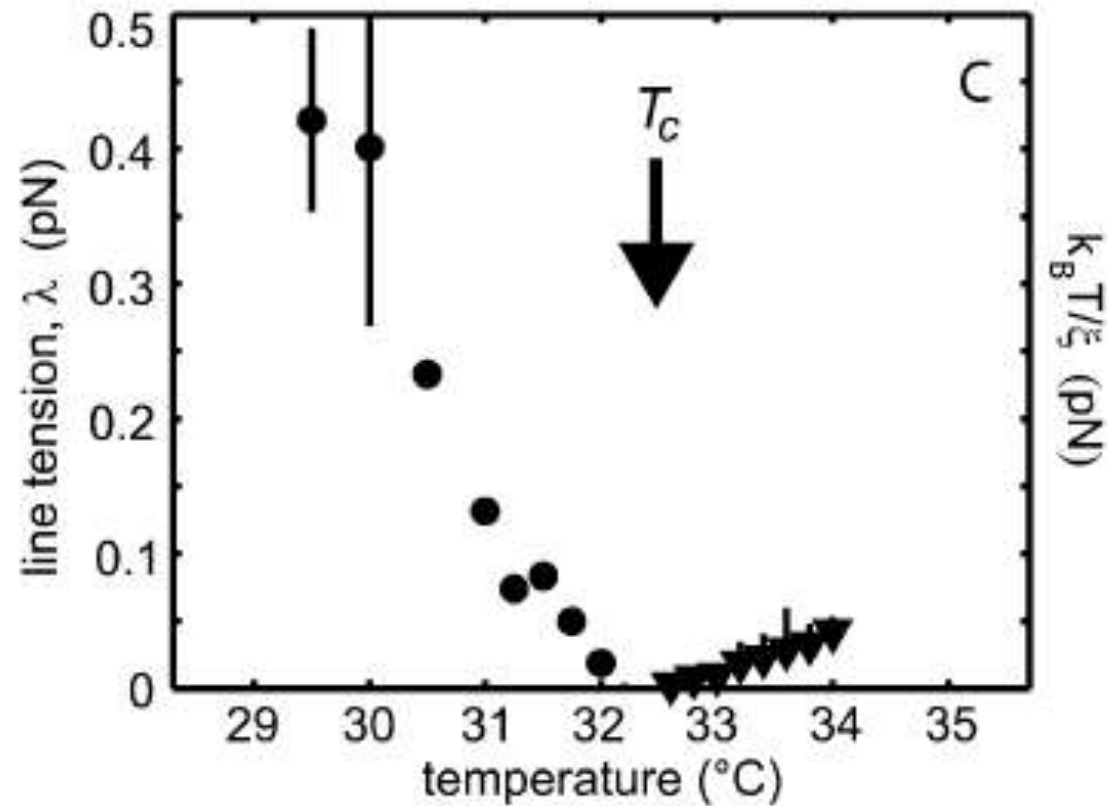
Measuring Line Tension from Fluctuations



L : smooth perimeter

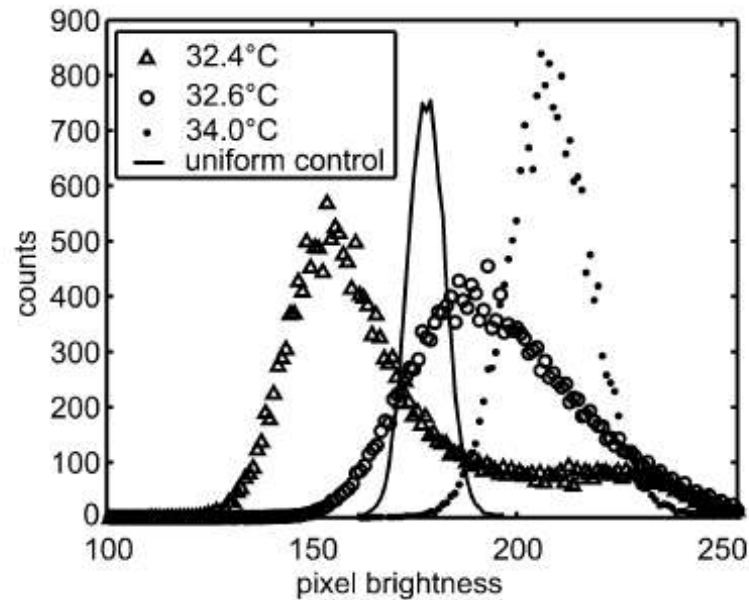
$$\langle h(k) \rangle^2 = \frac{k_B T}{\lambda L k^2}$$

Line Tension versus Temperature



λ vanishes at $T=T_c$

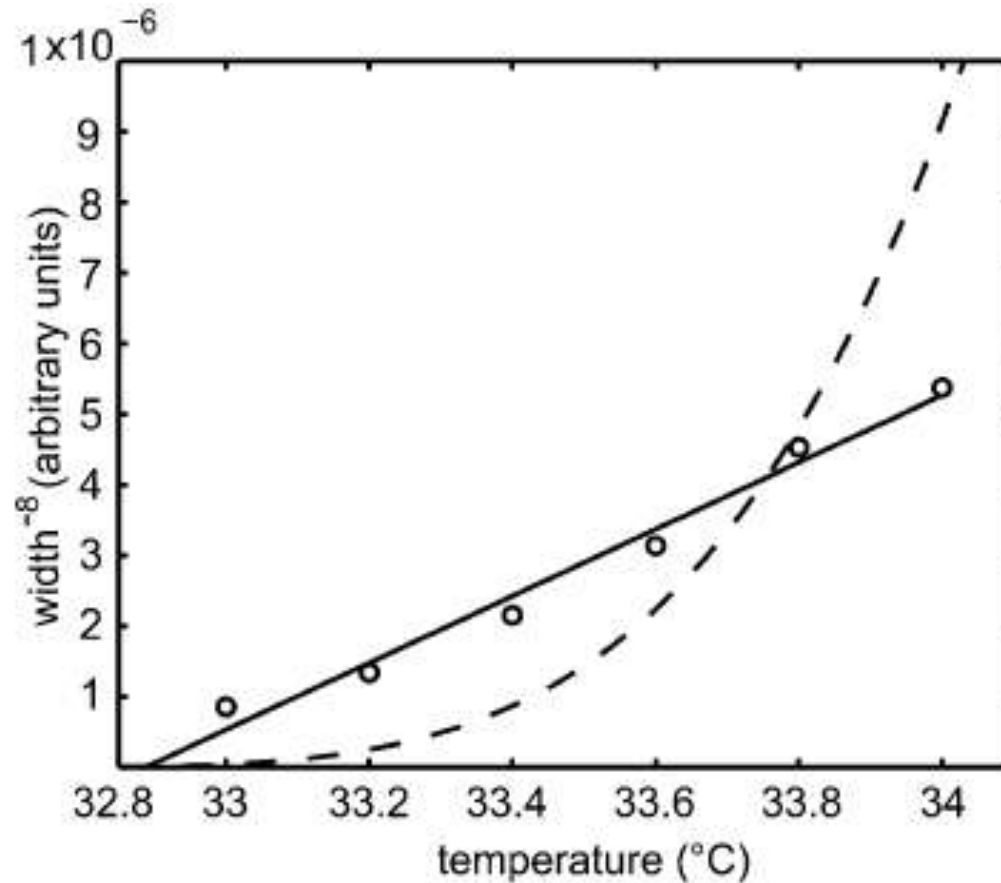
Critical Exponent



Mean Brightness $\langle m^2 \rangle^{1/2} \approx$ order parameter

$$W = \langle m^2 \rangle^{1/2} \propto \left(\frac{T - T_c}{T_c} \right)^{-\beta}$$

Critical Exponent

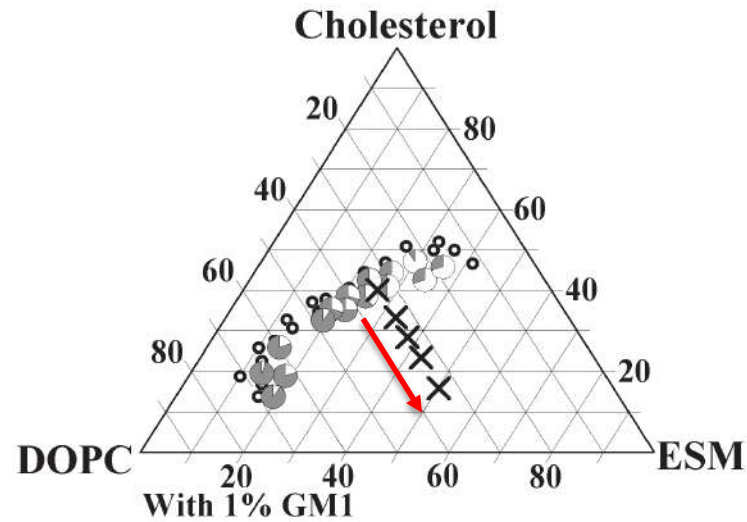


$$W^{-8} \propto T - T_c$$

➡ $\beta = \frac{1}{8} = 0.125$

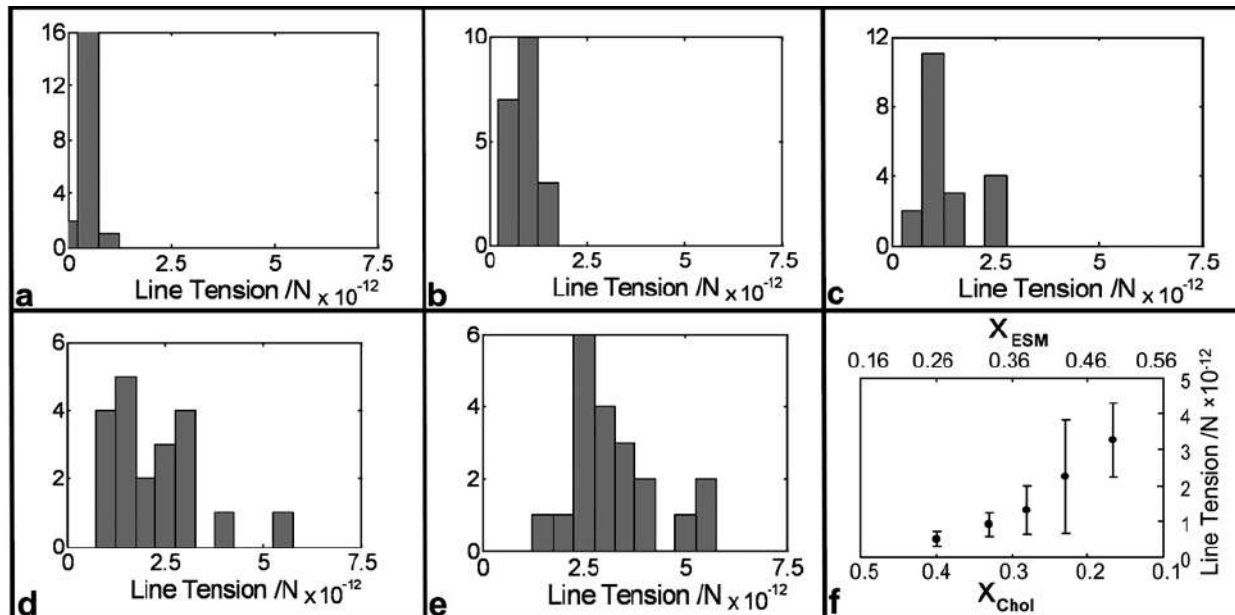
OK with a 2D-Ising model

Line Tension versus Lipid Composition



Reducing cholesterol

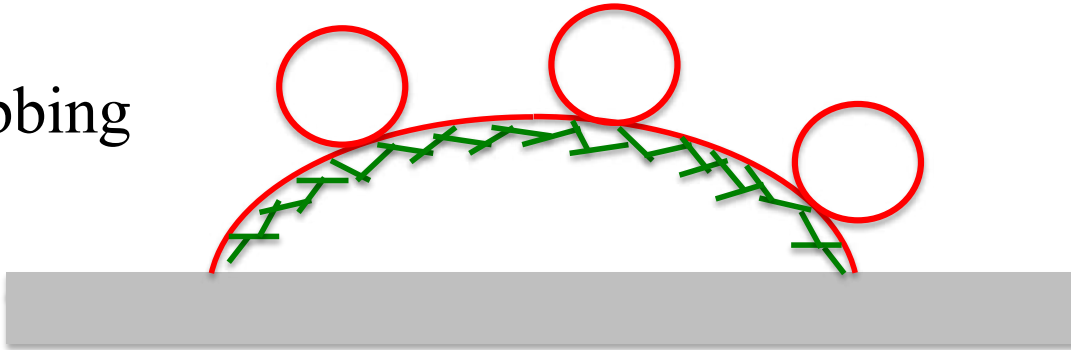
→ λ increases



A. Tian... T. Baumgart *Phys. Rev. Lett.* (2007)

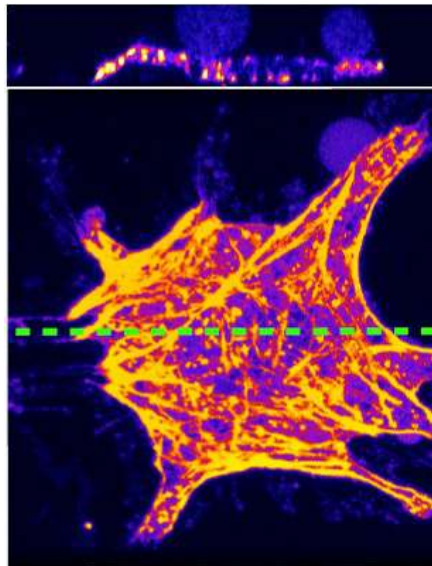
Plasma Membrane: Composition close to a Critical Point

Cell blebbing



Review:

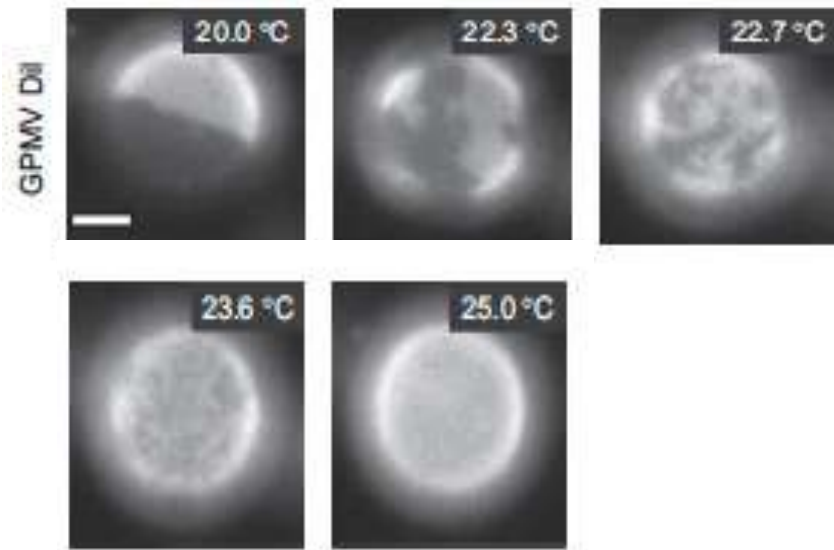
E. Sezgin et al *Nature Protocols* (2012)



GPMVs

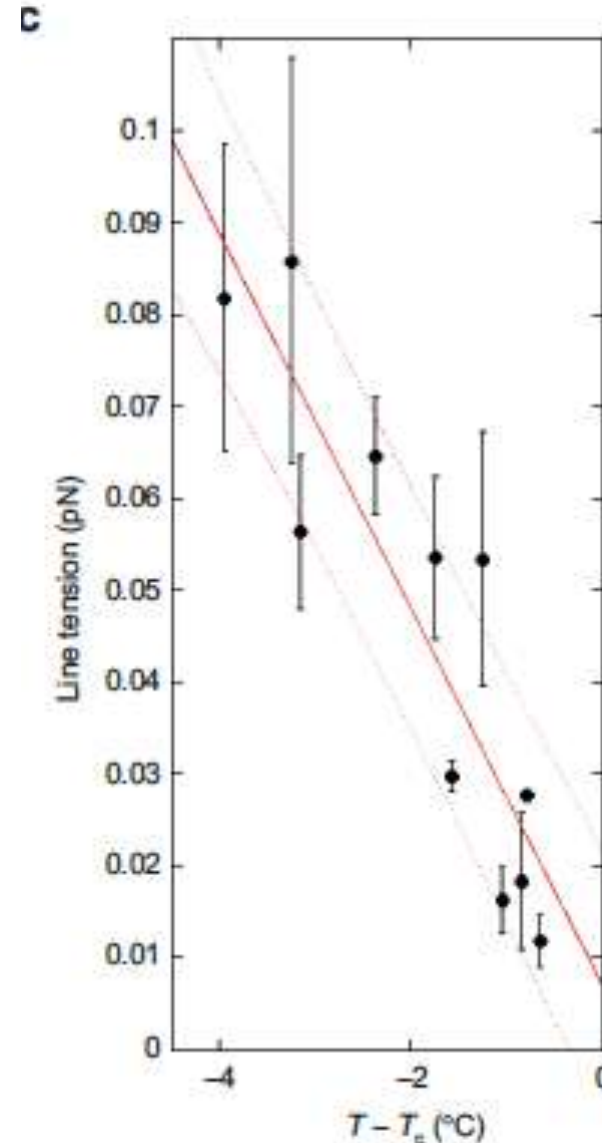
B. Sinha et al., *Cell* (2011)

Plasma Membrane: Composition close to a Critical Point



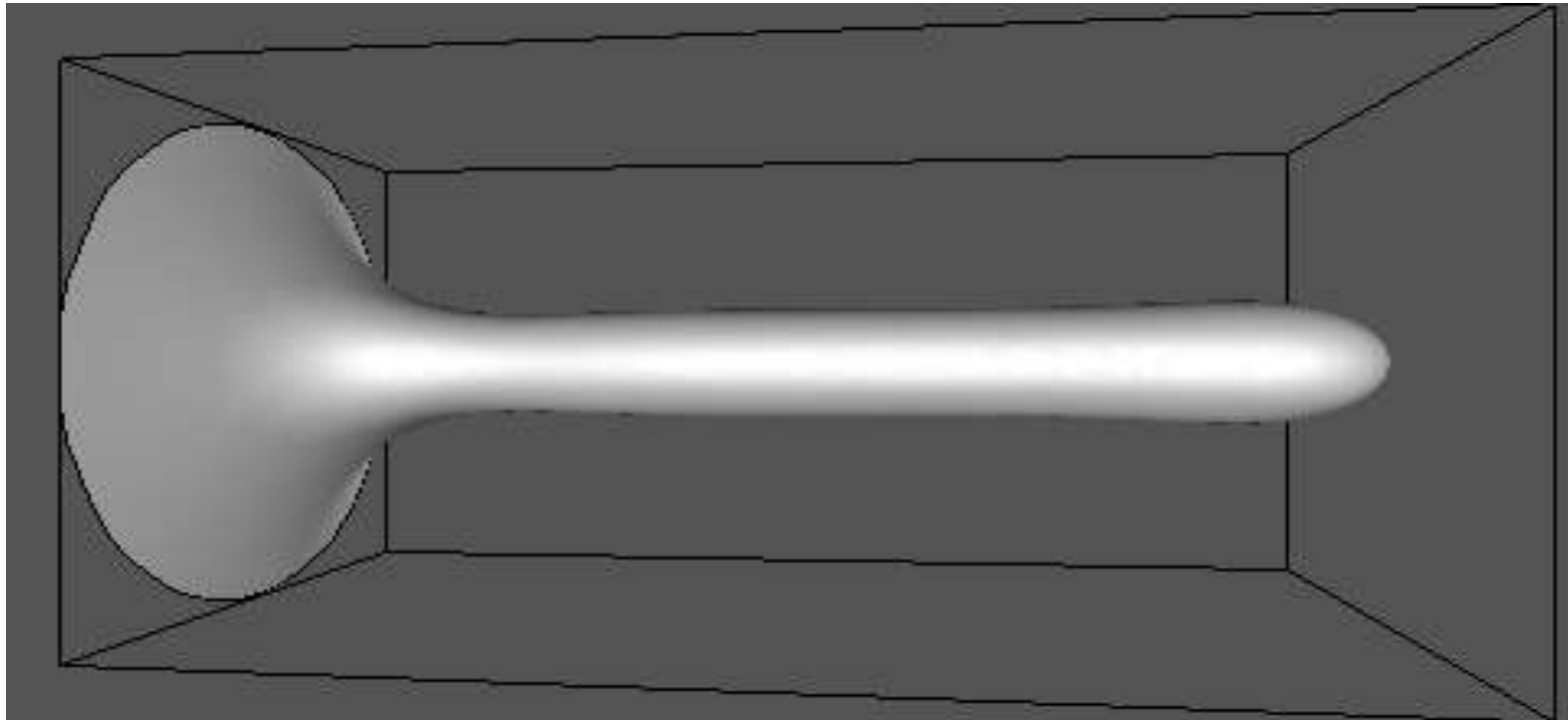
Cf Raft: size of domains determined by the proximity to some underlying critical point.

Sensitive to various stimuli



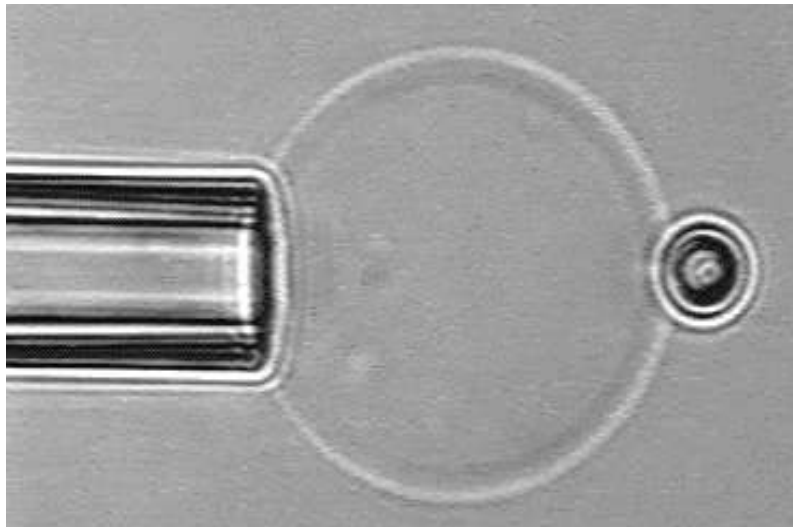
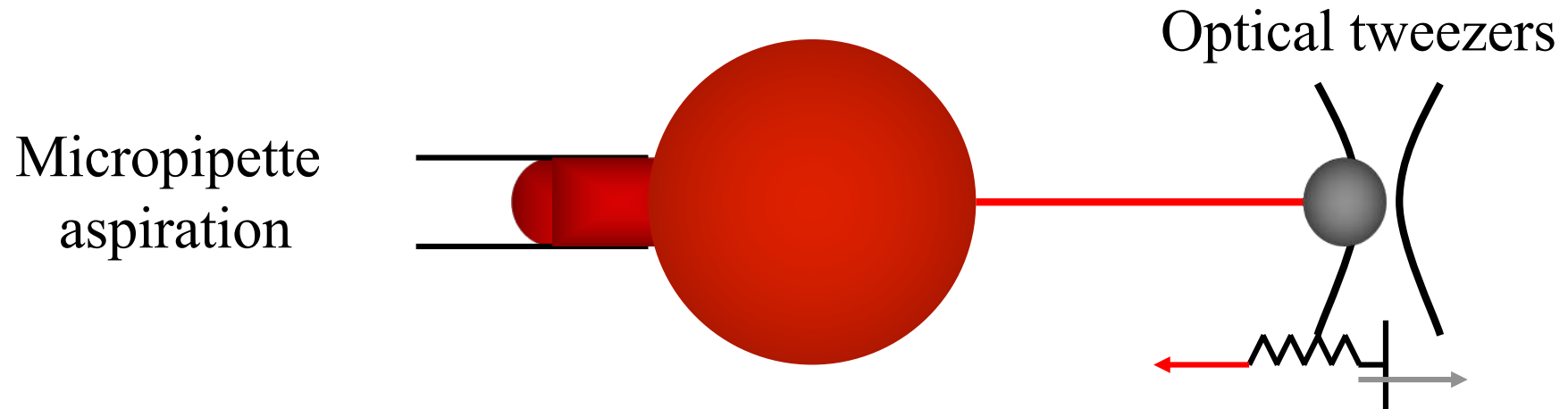
What Happens in a Highly Curved Membrane?

Membrane Nanotubes

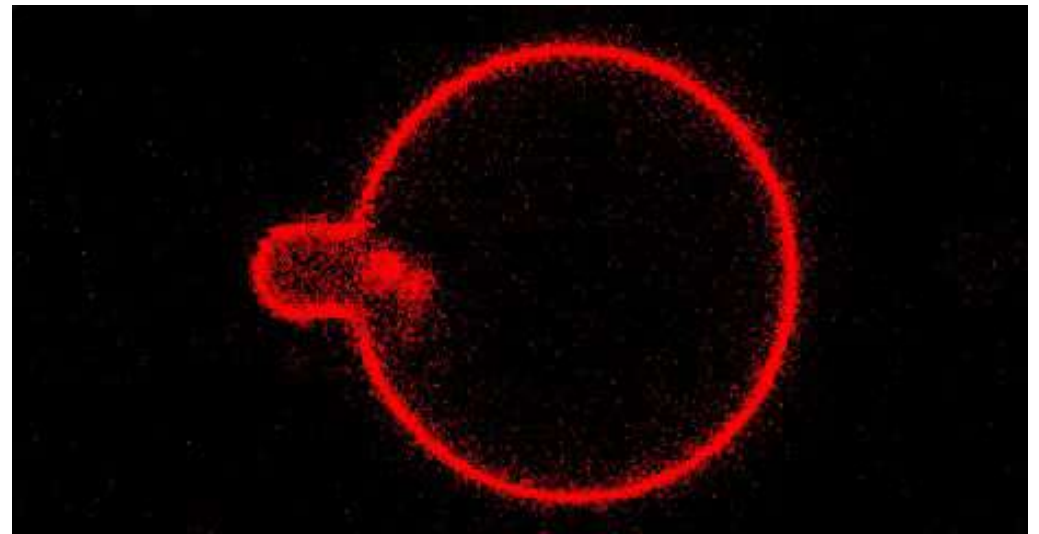


From Derényi et al, *PRL* (2002)

Membrane Nanotubes : Highly Curved

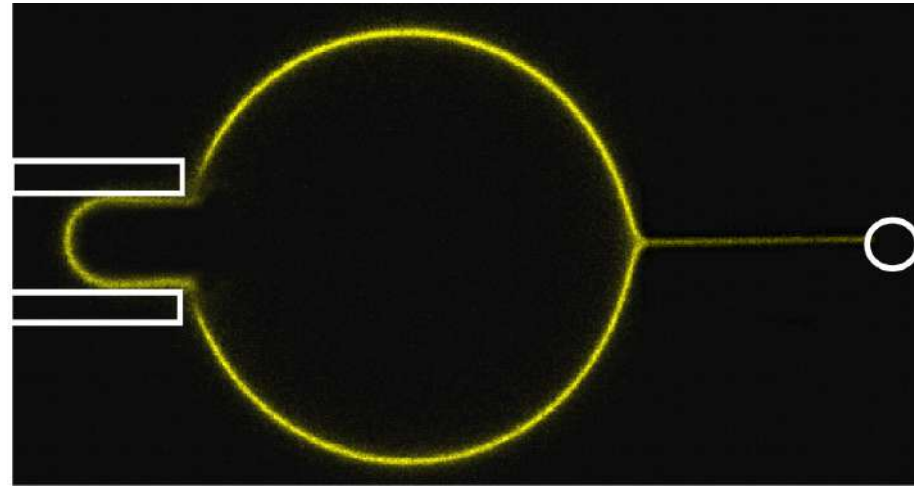


D. Cuvelier ... P. Nassoy *Biophys. J* (2005)

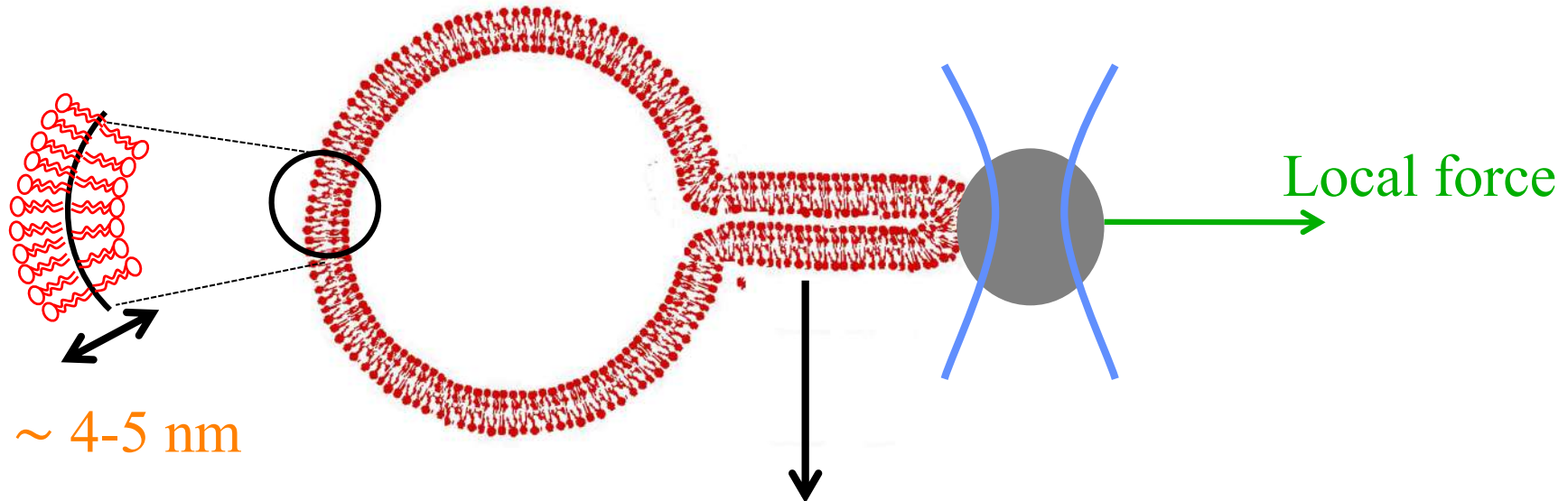


B. Surre et al, *PNAS* (2009)

Membrane Nanotubes : Highly Curved



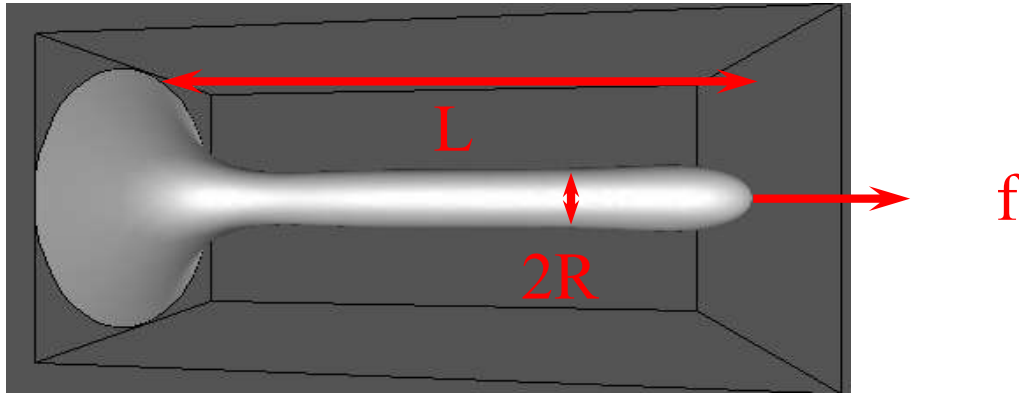
~ 10-50 μm



~ 4-5 nm

$R \sim 7 \text{ nm to few } 100 \text{ nm}$

Physics of Membrane Nanotubes



Some references

- Evans, Yeung *Chem. Phys. Lipids* (1994)
- Bozic, Svetina, Zeks *PRL* (1997)
- Svetina, Zeks, Waugh, Raphael *Eur. B. J* (1998)
- Derényi, Jülicher, Prost *PRL* (2002)
- Powers, Huber, Goldstein *PRL* (2002)

$$H = \iint_A dA \left[\frac{\kappa}{2} C^2 + \sigma \right] - fL$$

$$E_{tube} = 2\pi L \frac{\kappa}{2R} + 2\pi R \sigma L - fL$$

$$\frac{\partial E}{\partial R} = 0$$

Radius:

$$R_0 = \sqrt{\frac{\kappa}{2\sigma}}$$

$\approx 10 \text{ nm} - \text{few } 100 \text{ nm}$

Set by tension

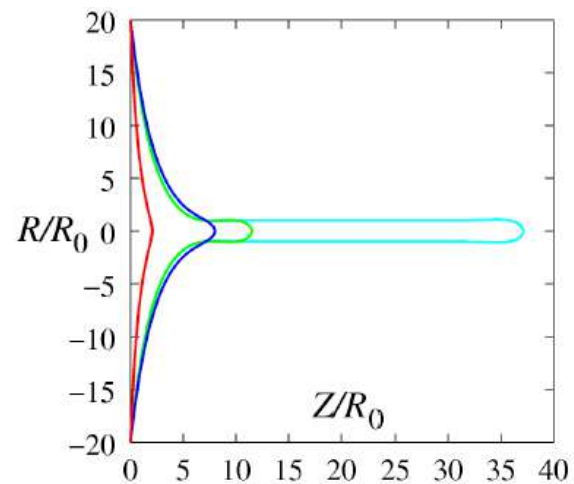
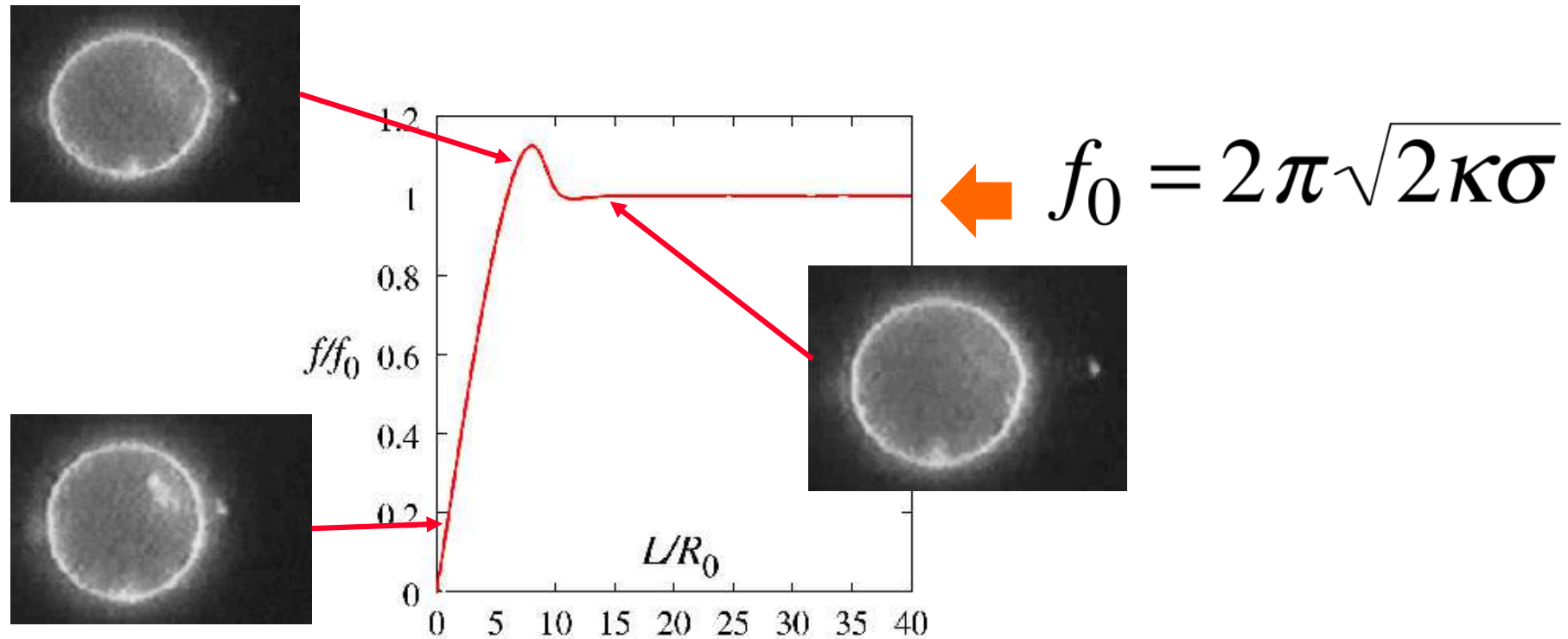
$$\frac{\partial E}{\partial L} = 0$$

Force:

$$f_0 = 2\pi \sqrt{2\kappa\sigma}$$

few pN - few 10 pN

Theory: Force/Extension at Constant Tension



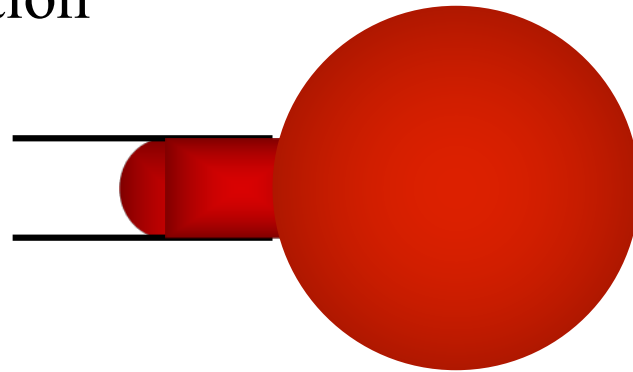
Dérényi et al, PRL **88** (2002) 238101

Powers et al PRE **65** (2002) 041901.

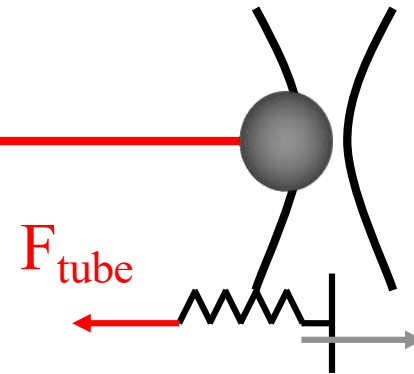
Nanotube - Controlled Curvature

Controlling curvature:
Micropipette aspiration

$$R_0 = \sqrt{\frac{\kappa}{2\sigma}}$$



Force:
Optical tweezers



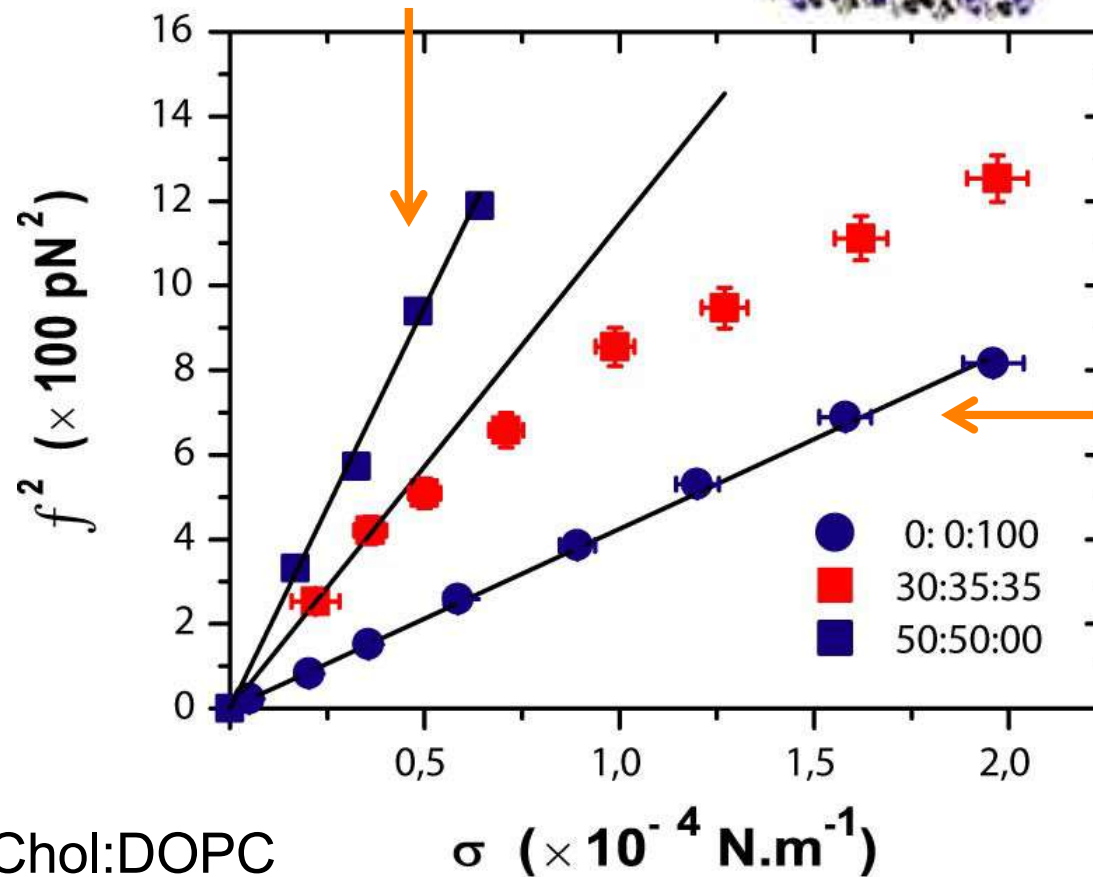
$$f_0 = 2\pi\sqrt{2\kappa\sigma}$$

Force ↔ Membrane tension
Radius ↔ Lipid composition

Experiments

$$f_0^2 = 8\pi^2 \kappa \cdot \sigma$$

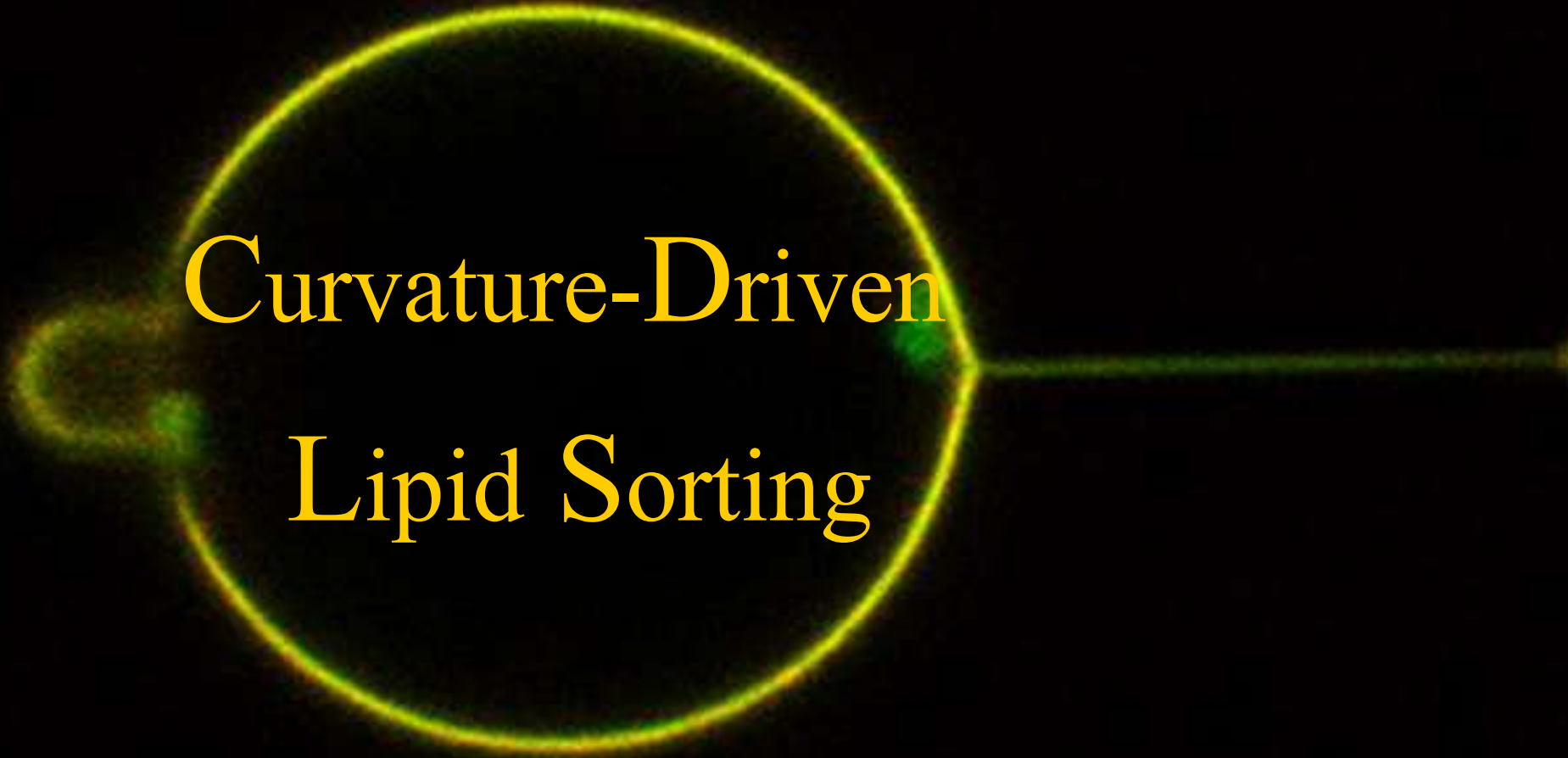
$$\kappa_{\text{SM:Chol}} = 65 \pm 10 \text{ k}_B\text{T}$$



$$\kappa_{\text{DOPC}} = 16 \pm 2 \text{ k}_B\text{T}$$



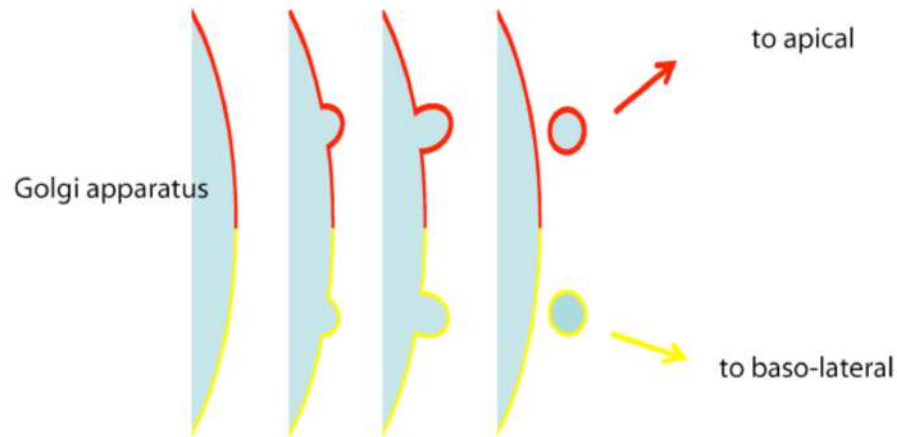
SM:Chol:DOPC

A glowing green circular structure, possibly representing a lipid bilayer or a vesicle, is centered on a black background. A horizontal line of green light passes through the center of the circle. The text "Curvature-Driven Lipid Sorting" is overlaid on the circle in a yellow, serif font.

Curvature-Driven Lipid Sorting

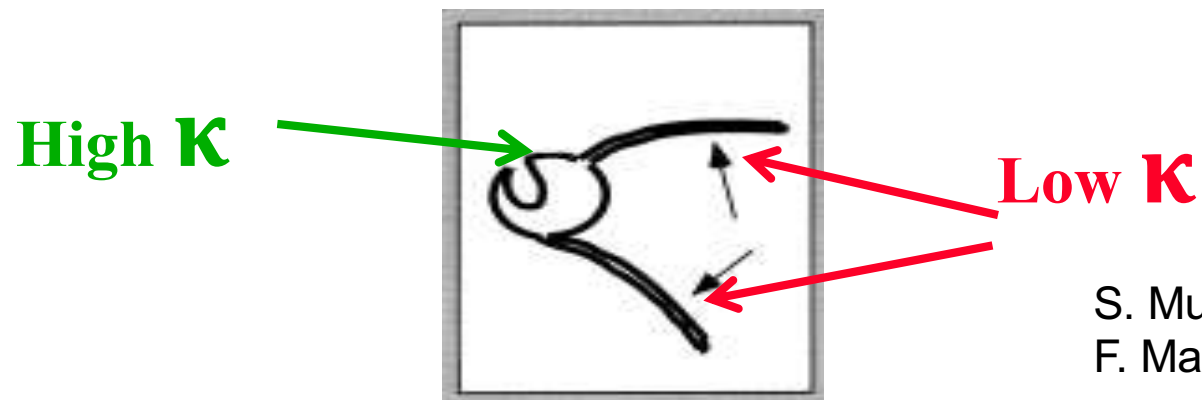
Sorting Lipids ?

- *Lipid rafts*



K. Simons, G. van Meer
Biochemistry (1988)

- *Global: Membrane mechanical properties*

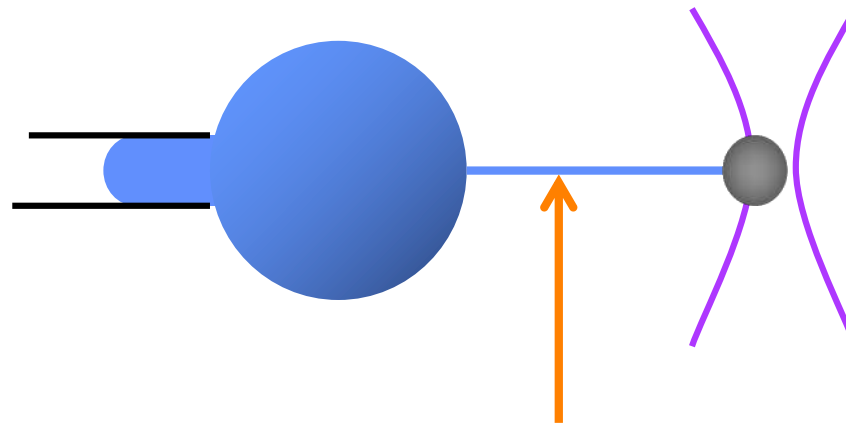


S. Mukherjee, T. Soe
F. Maxfield, *J. Cell Biol* (1999)

Expected:

Sphingomyelin/Cholesterol/DOPC

Composition	$K (\kappa_B T)$
DOPC	16
DOPC+50% Chol	30
SM+50% Chol	65



Low κ

PC rich

Sphingo. depleted

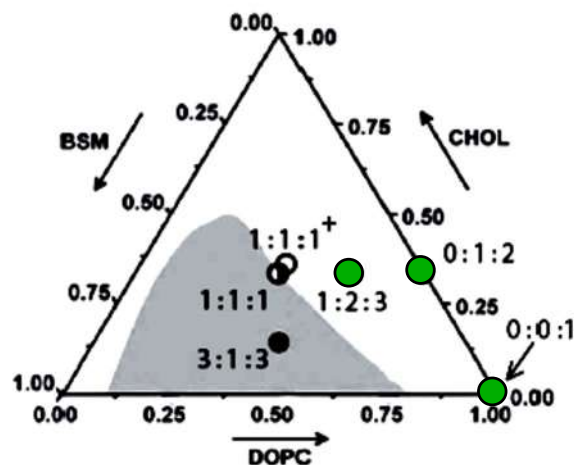


Rhodamine PE (Ld)

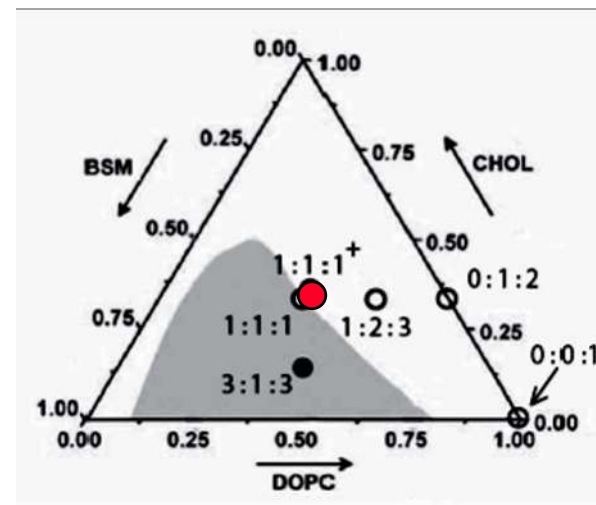
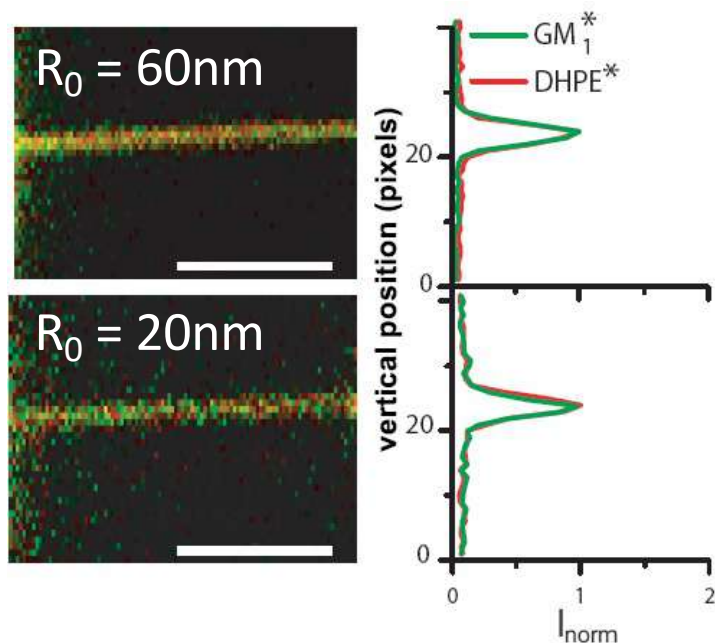
GM1* (no preference)

Sorting coefft:
$$S = \frac{I_{PE}^t}{I_{GM_1^*}^t} \bigg/ \frac{I_{PE}^v}{I_{GM_1^*}^v}$$

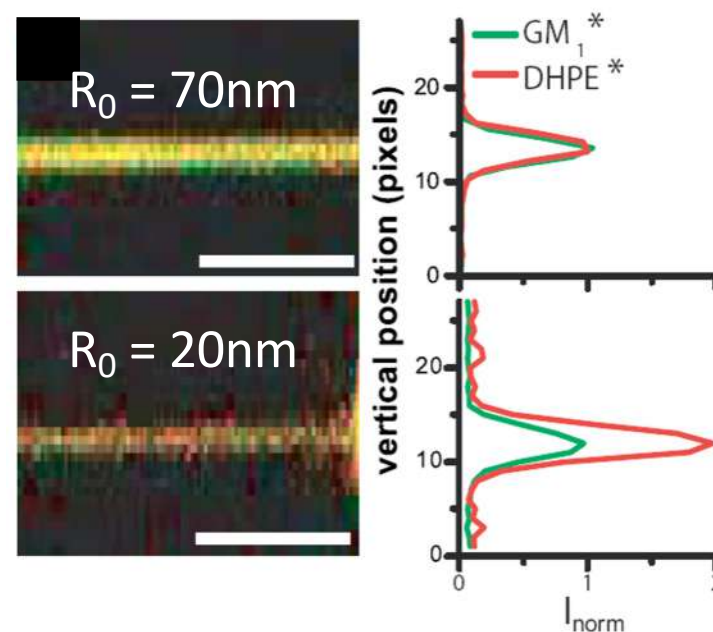
Sorting : Only in the Vicinity of Phase Boundary



Far from phase boundary



Close to phase boundary



— Sorre, Callan-Jones *et al.* PNAS (2009)

Theoretical Modeling

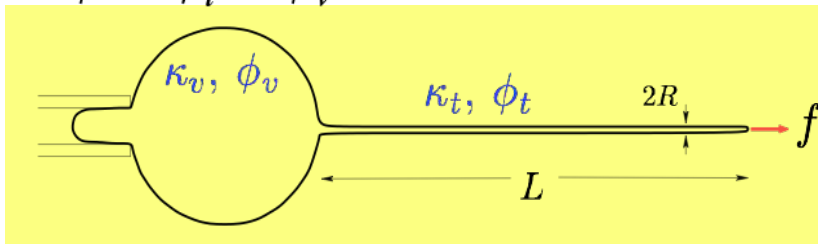
A. Callan-Jones, J.F. Joanny, J. Prost

$$F_t = 2\pi R L_{tube} \left[\frac{\kappa(\phi)}{2R^2} + \sigma + \frac{1}{2} \lambda(\phi_v) \Delta\phi^2 \right] - f_{tube} \times L_{tube}$$

Gain

Entropic Penalty

$$\Delta\phi = \phi_t - \phi_v$$



$$\lambda = \frac{kT}{a} \left(\frac{1}{\phi_v(1-\phi_v)} - 2\chi \right)$$

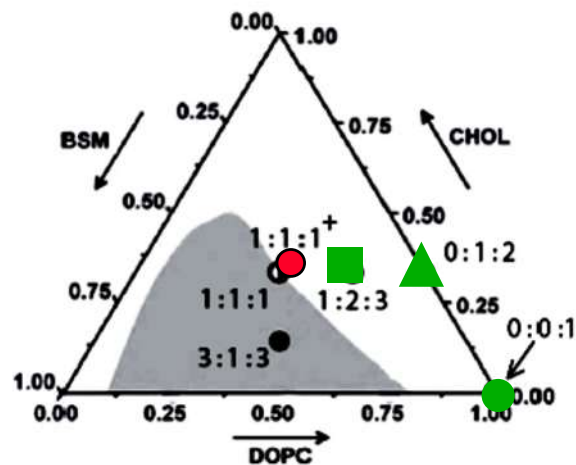
a: size of sorted specie
 χ : lipid-lipid interaction

$\lambda = 0$ at the demixing limit (spinodal)

If not close to demixing, mixing entropy dominant

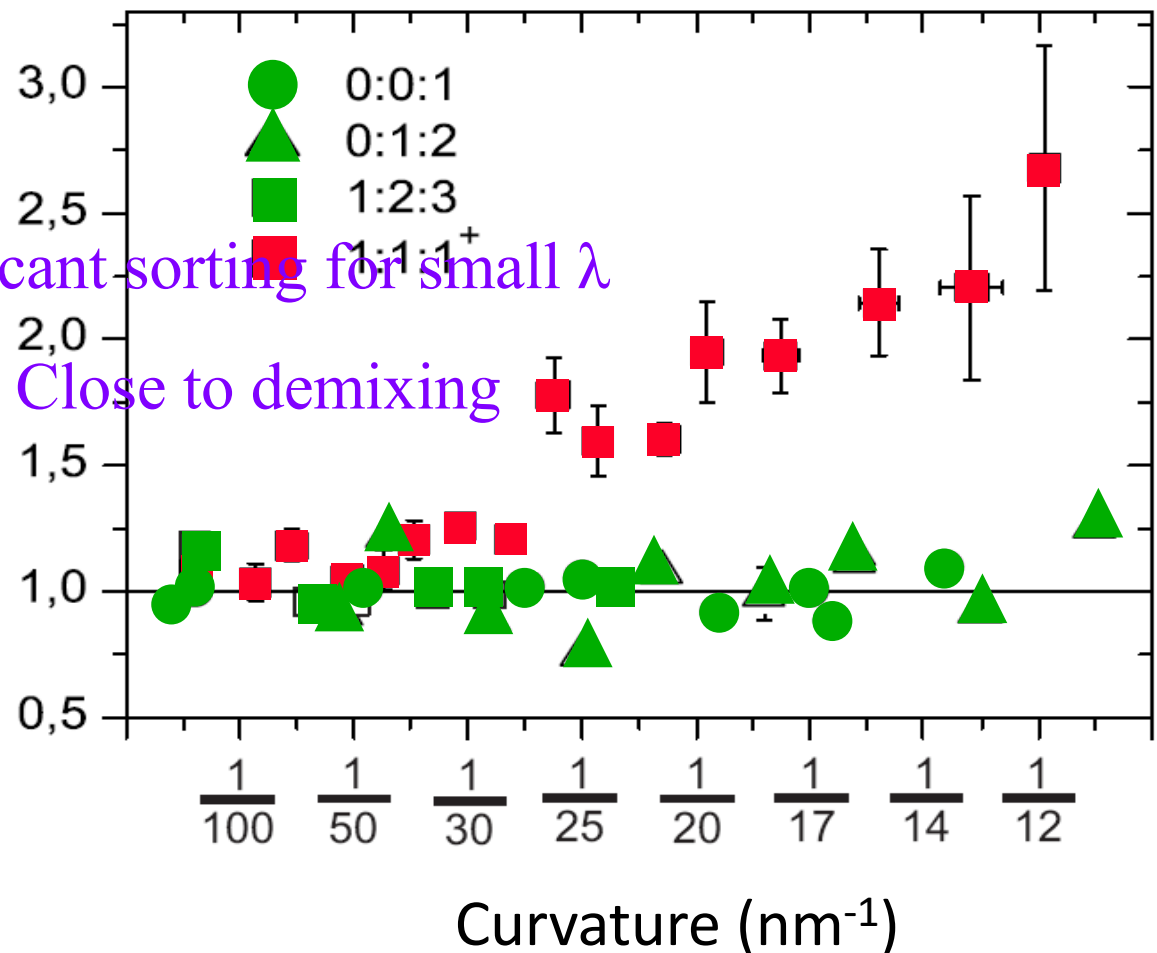
Curvature and Lipid Sorting

Sorting: $\phi_t - \phi_v \propto \frac{1}{\Phi_t} \frac{1}{\lambda R_t^2} \propto \frac{1}{\lambda} \frac{1}{\sigma}$



Significant sorting for small λ
Close to demixing

Near phase boundary:
Sorting increases with
curvature



Tube enriched in DOPC, depleted in SM

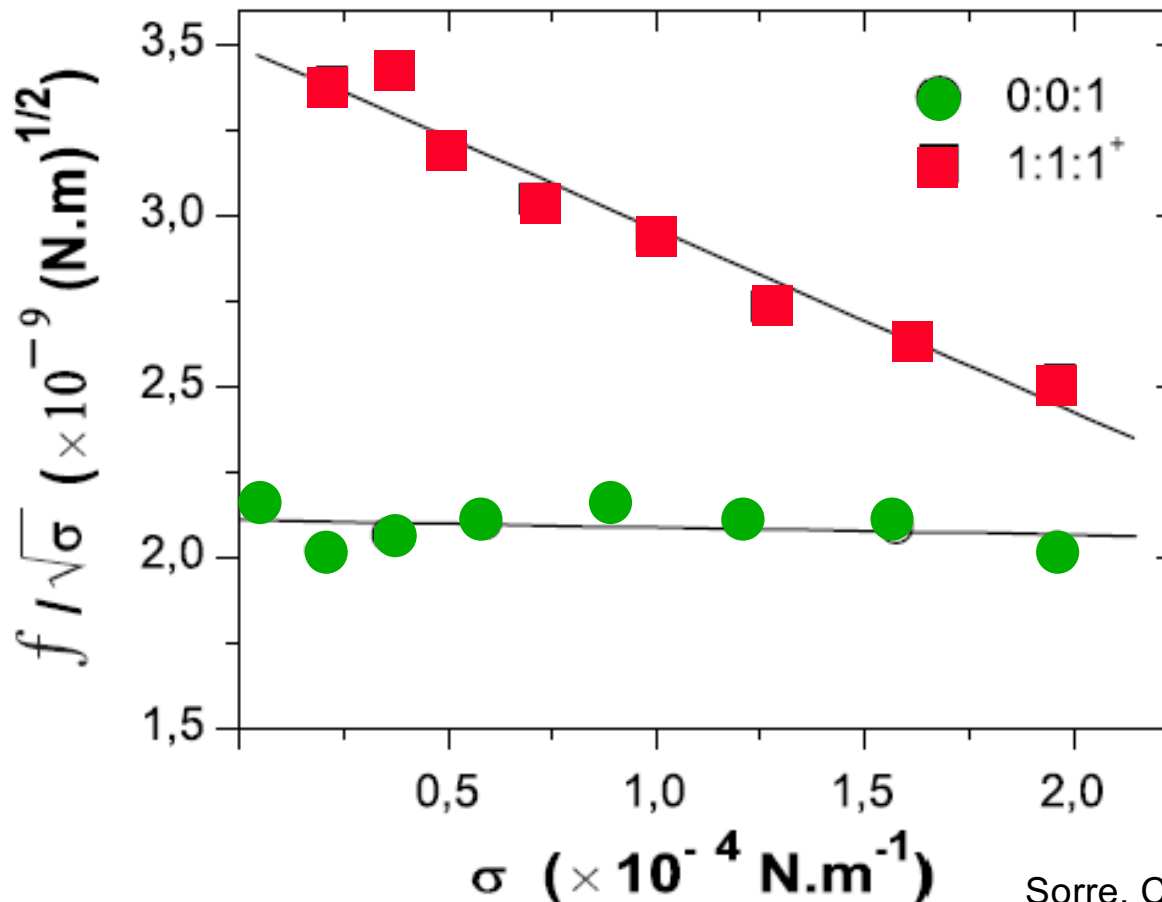
Sorre, Callan-Jones *et al.* PNAS (2009)

Effect on the Force

$$f(\Phi_t) \approx 2\pi\sqrt{2\kappa_t\sigma} \left[1 - \frac{\sigma}{\lambda} \right]$$

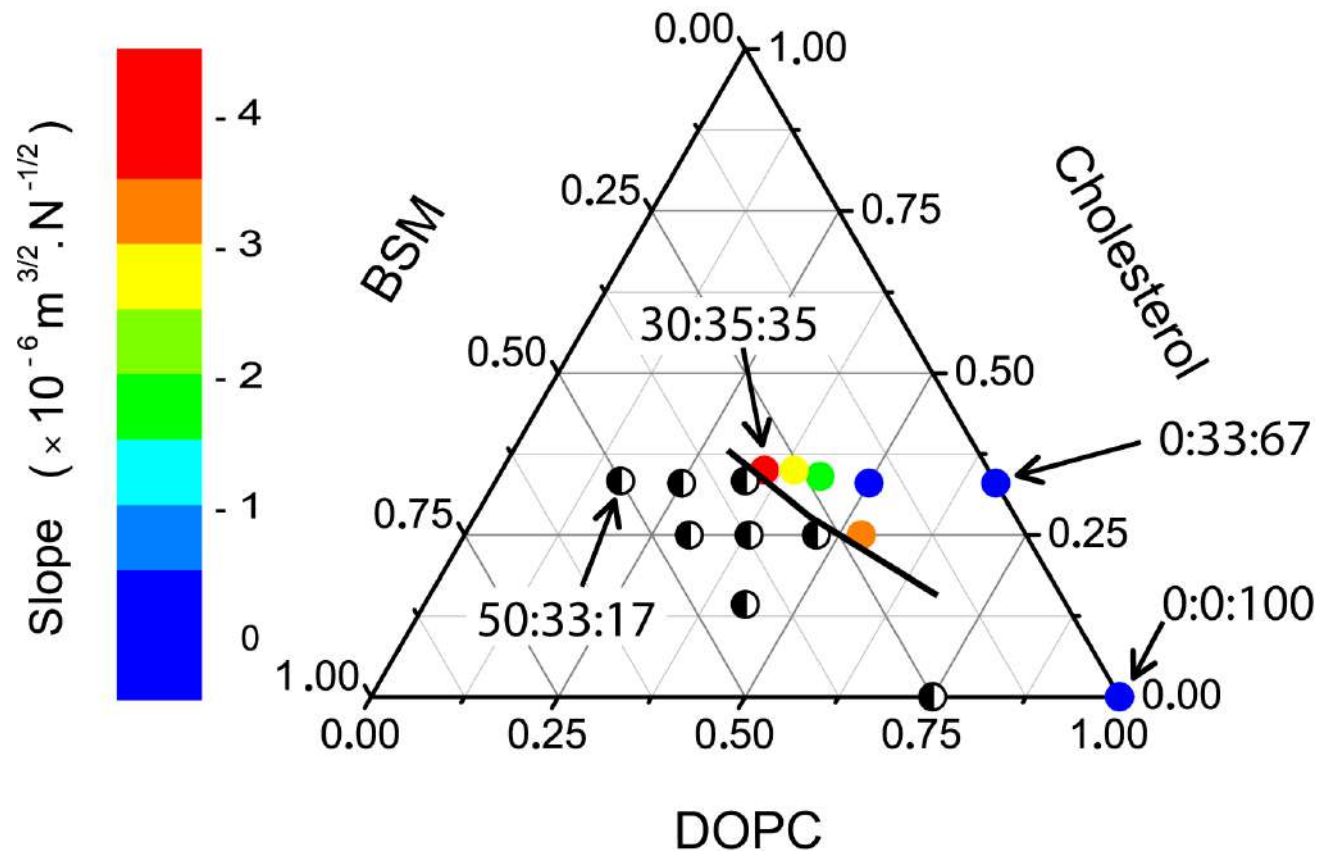
λ  when approaching demixing
 Larger deviation of the force

Sorting Strength



Curvature-driven Sorting Needs Proximity to a Demixing Point

Sorting Strength



Sorre, Callan-Jones *et al.* PNAS (2009)

(Also Tian *et al.*, B.J. (2009)).

Conclusions

- Lipid sorting only close to demixing: tube enriched in Ld phase (lowest bending rigidity)

Biological membranes: PM close to demixing

"Plasma membranes are poised for activation of raft phase coalescence at physiological temperature"

D. Lingwood et al. PNAS (2008)

- If lipids only, requires tight regulation of lipid composition

How to reduce the entropic penalty due to lipid mixing?

$$\Delta f_m = \frac{1}{2} \lambda \Delta \phi^2$$

➡ Minimize λ

$$\lambda = \frac{kT}{a} \left(\frac{1}{\phi_v(1-\phi_v)} - 2\chi \right)$$

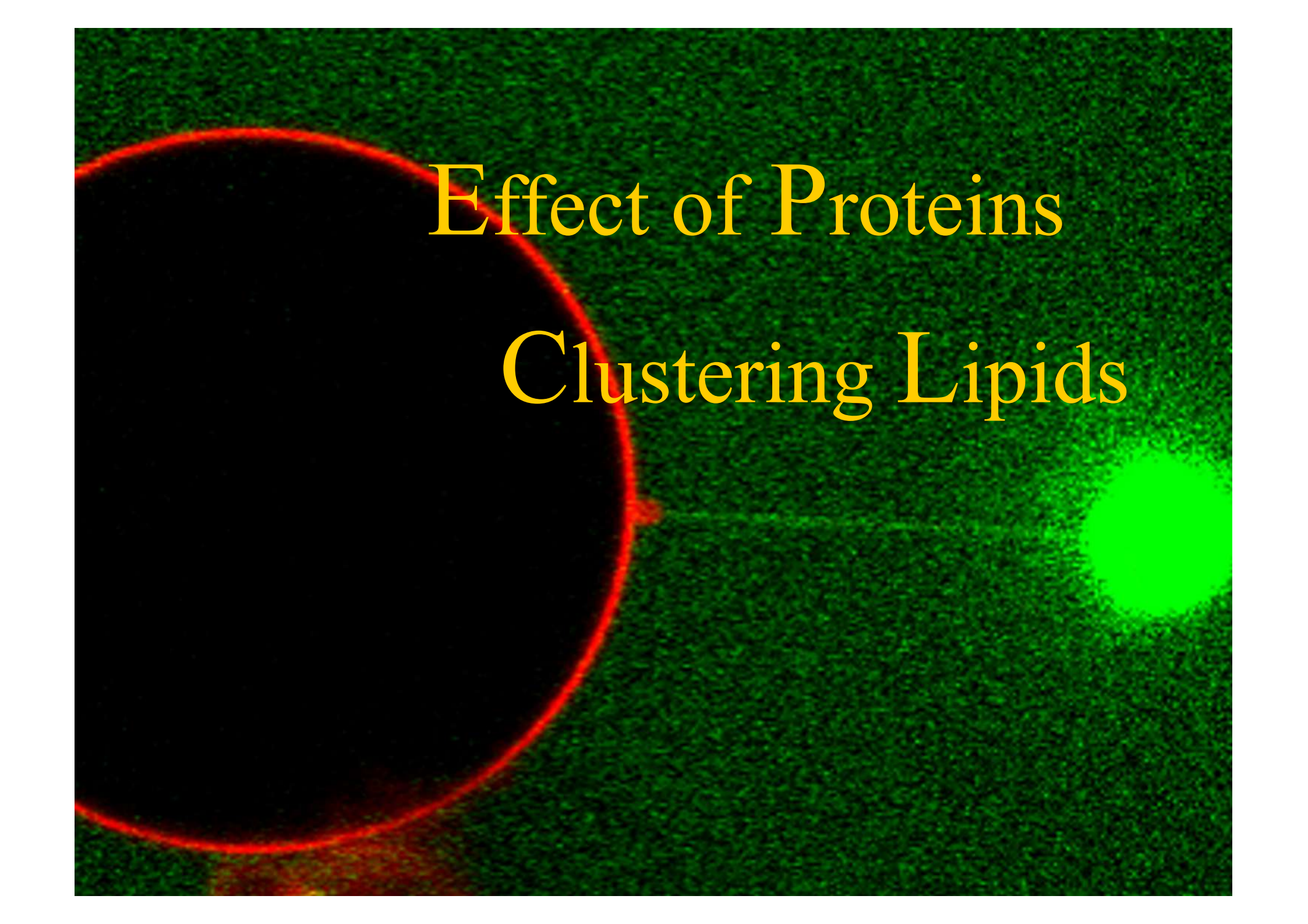
a: size of sorted specie
 χ : lipid-lipid interaction

➡ a: Increase lipid size ➡ clustering

Stronger sorting for proteins

➡ χ : Increase short range interaction

➡ Add a protein with preferential interaction



The background of the slide is a fluorescence microscopy image of a cell membrane. A large, dark, circular region on the left is outlined by a bright red ring, representing a lipid cluster. To the right of this, there is a bright green, circular cluster, representing a protein cluster. The text is overlaid on the right side of the image.

Effect of Proteins Clustering Lipids

AB₅ Toxins Cluster Glycolipids

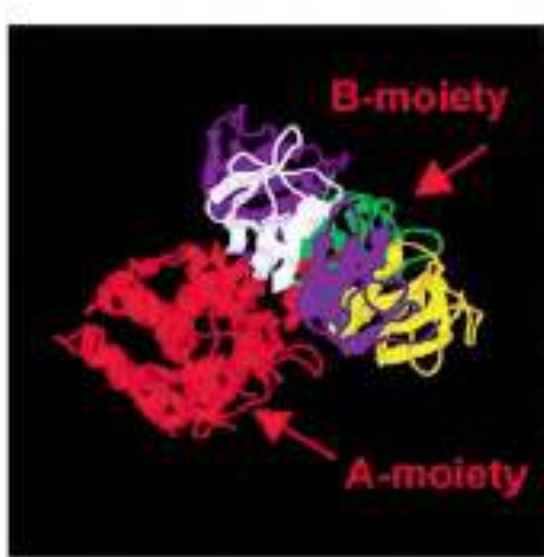


From J. Sanchez and J. Holmgren
Cell. Mol. Life Sci. (2008)

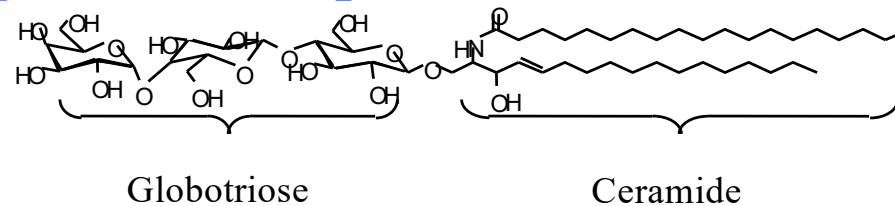
Cholera Toxin (CTxB)

- CtxB : pentamer
- Receptor = ganglioside GM1 lipid
- 1 CTxB pentamer $\longleftrightarrow$ **5** GM1

Shiga Toxin (STxB)



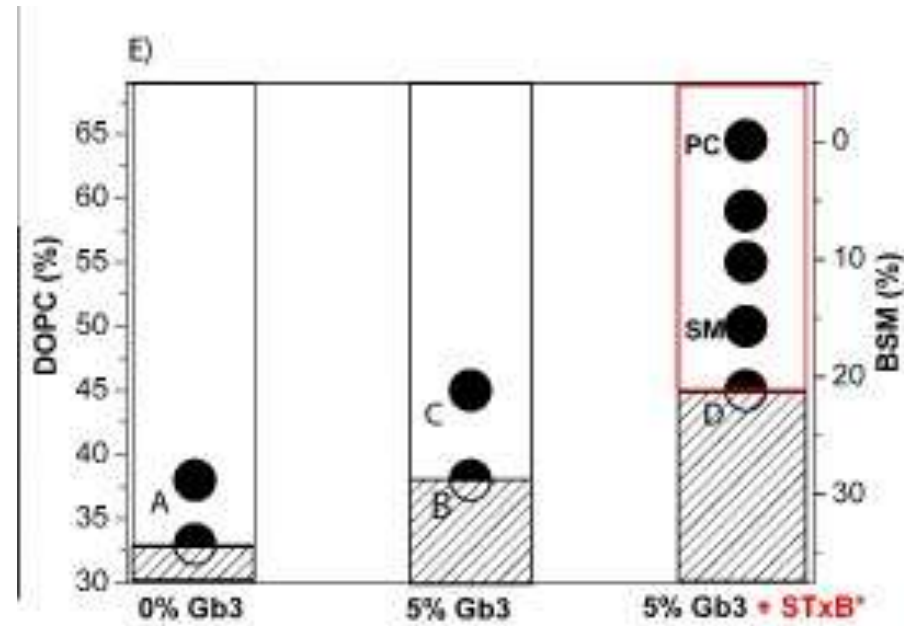
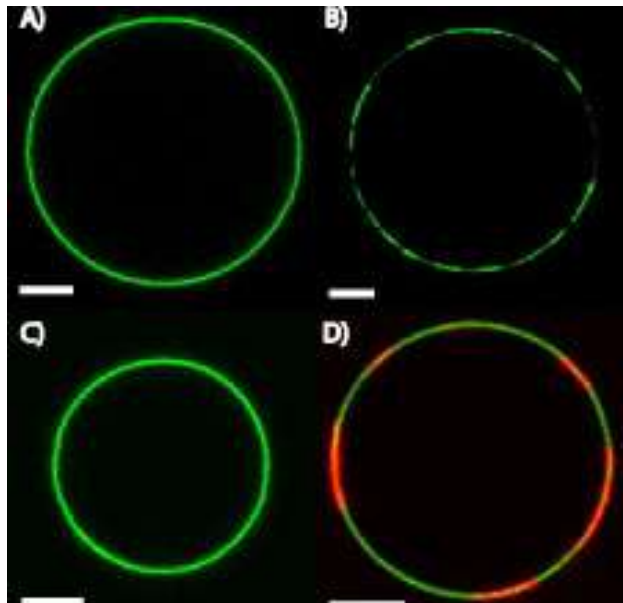
- StxB : pentamer
- Receptor = Gb3 lipid



- 1 STxB pentamer $\longleftrightarrow$ **15** Gb3

AB₅ Toxin Binding Facilitates Lipid Demixing

STxB

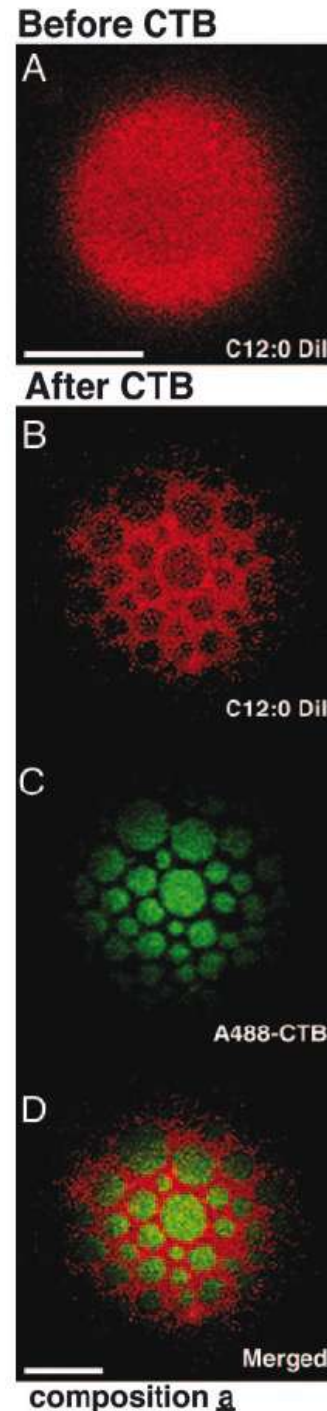


Safouane, Berland et al *Traffic*(2010)

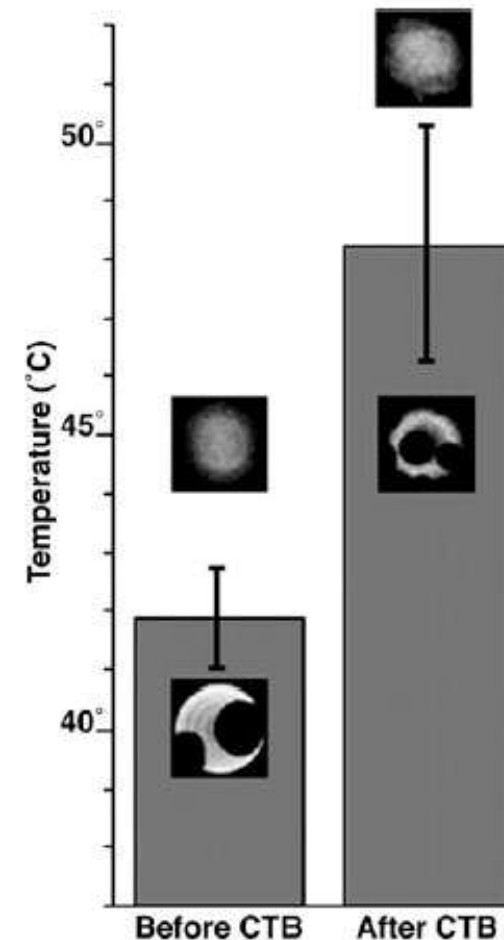
Clustering: reduces entropy cost

Vesicle close to
phase separation
NO toxin

With
CTxB



CTxB clusters lipids
Reduces mixing entropy
Phase separation facilitated
(at higher T°C)

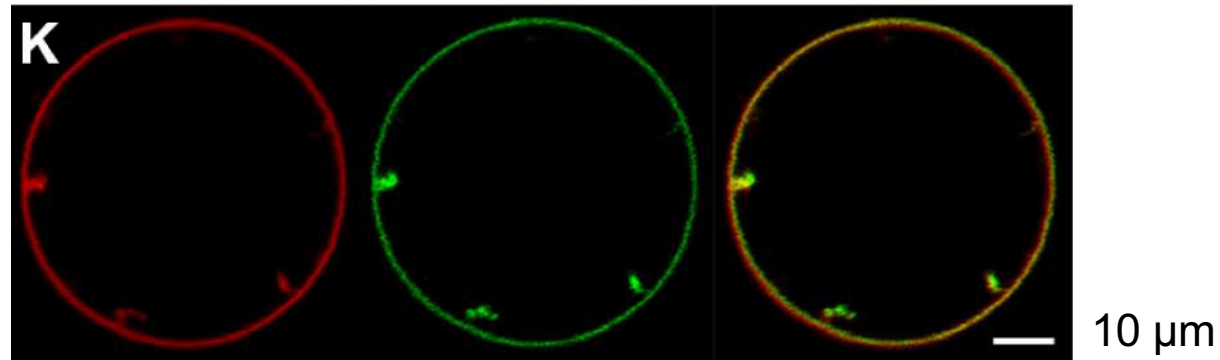


A. Hammond ... G. W. Feigenson (*PNAS*) 2005

CTxB and STxB Induce Membrane Invaginations

CTxB

DOPC+30% Chol
+5%GM1



10 μ m

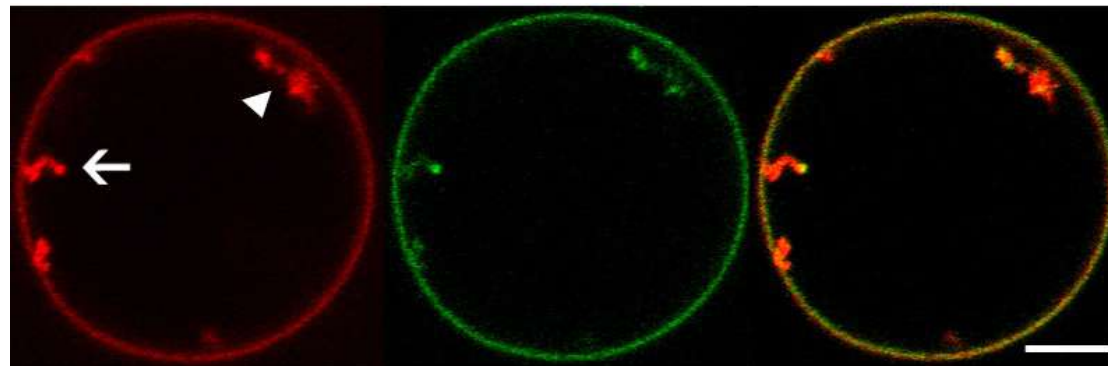
CTxB

Membrane

L. Berland, W. Römer

STxB

DOPC+30% Chol
+5% Gb3



10 μ m

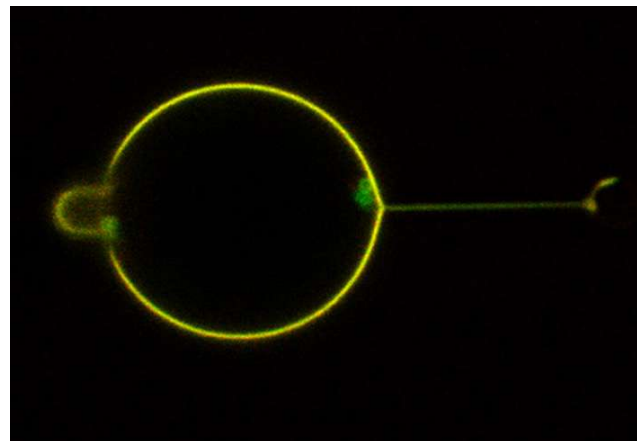
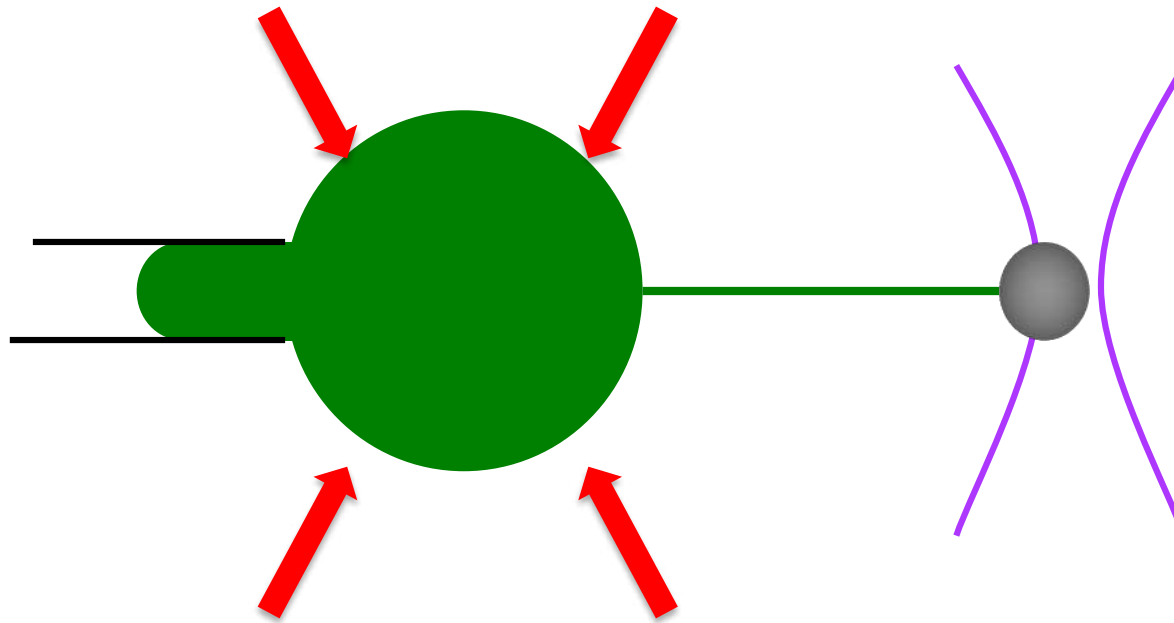
STxB

Membrane

W. Römer, L. Berland, et al., *Nature* (2007)

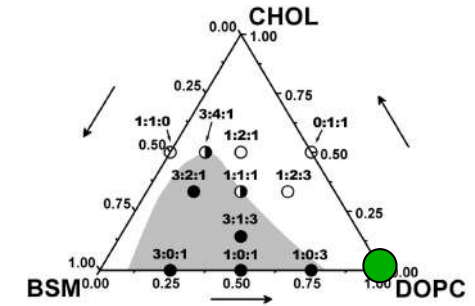
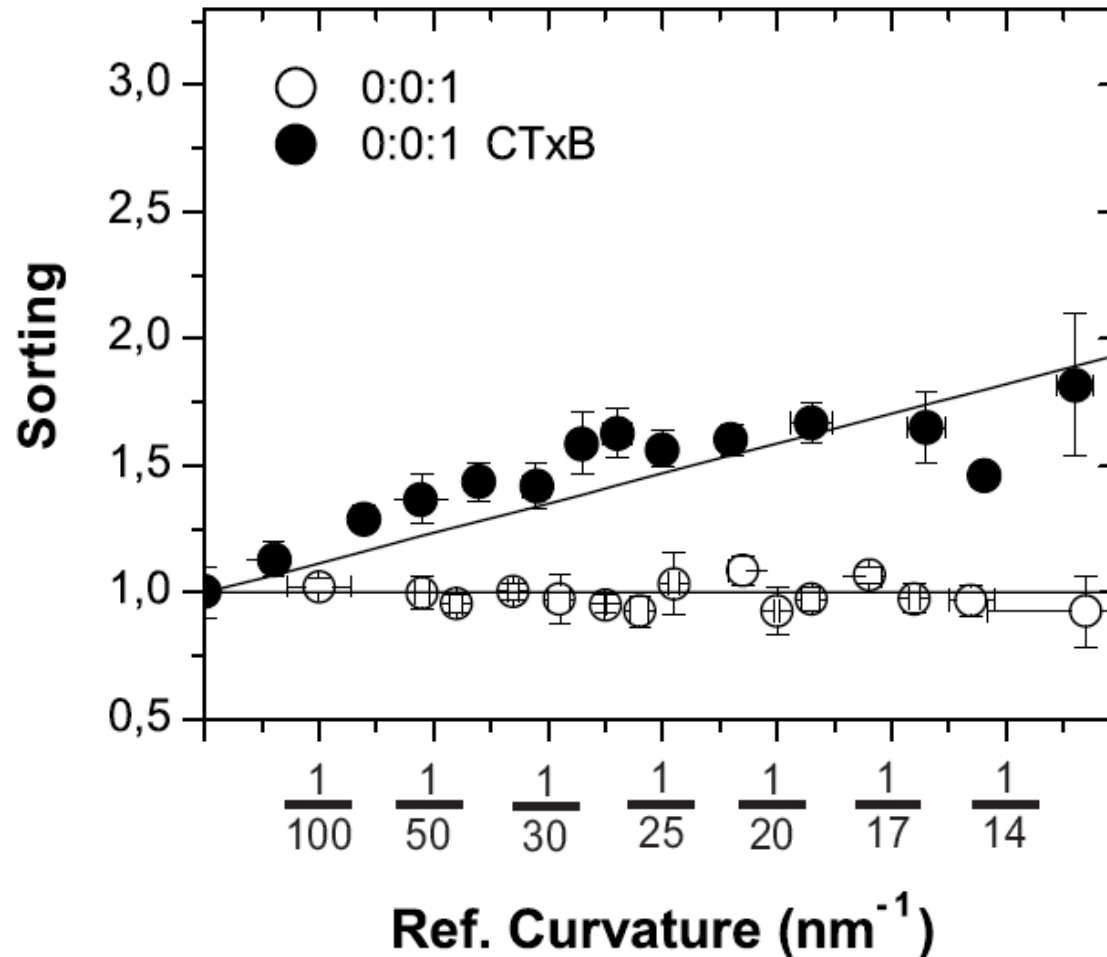
Cluster lipids + induce negative spontaneous curvature

CTxB added from outside : *depleted from the tube*



Cholera Toxin Sorting in DOPC Tubes

DOPC
(0:0:1)
PE*
1% GM1*
+ CTxB



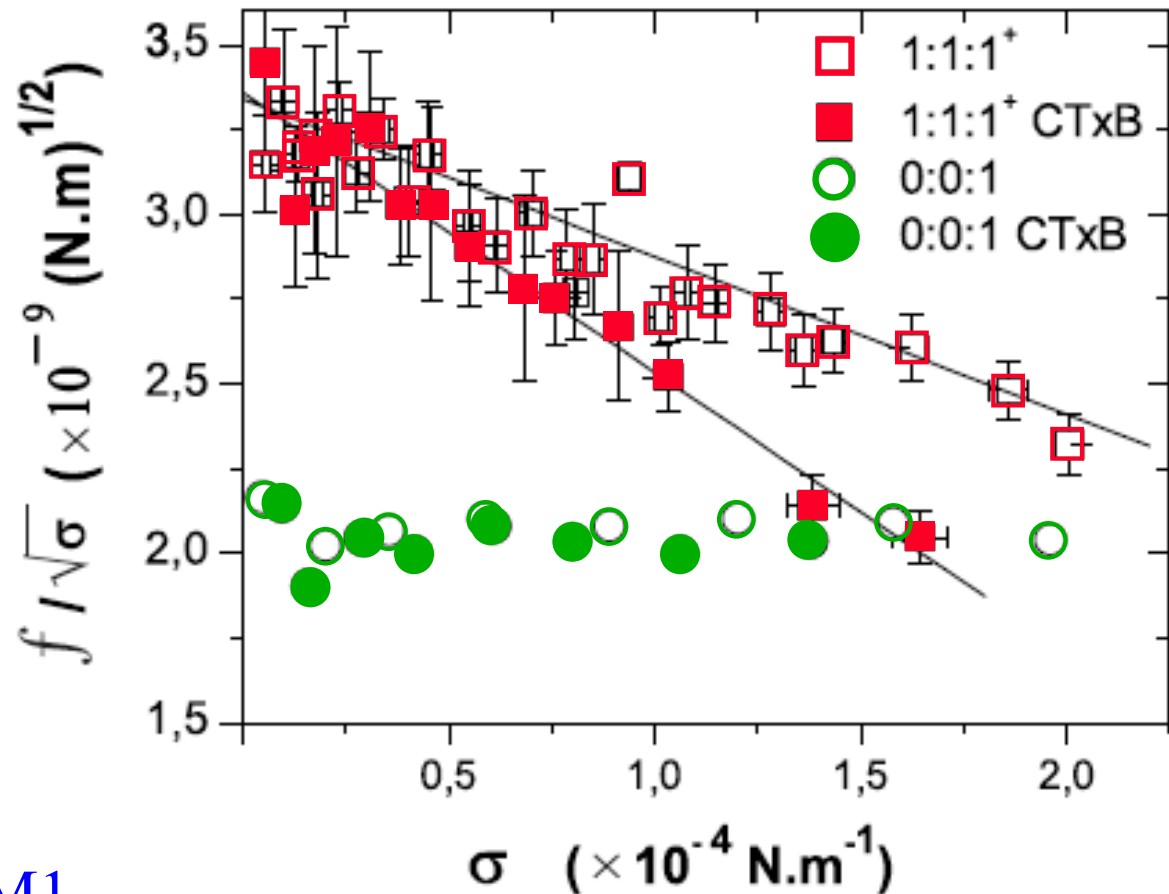
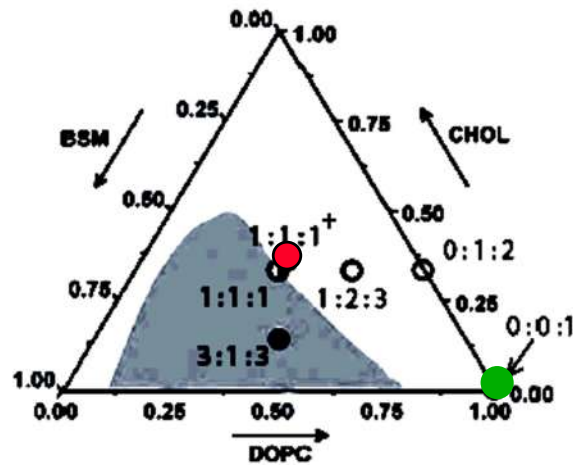
GM1* depleted from DOPC tubes

CTxB depleted from DOPC tubes

Consistent with CTxB-induced negative curvature

Sorre et al. *PNAS* (2009)
Tian, Baumgart, *B.J.* (2009)

Cholera Toxin Enhances Lipid Sorting Close to Demixing

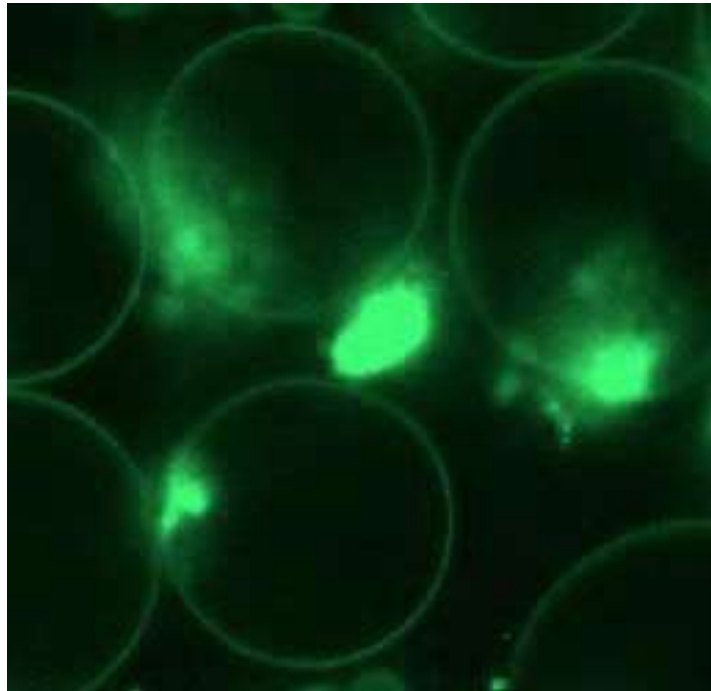


CTxB addition: clusters GM1
 λ decreases

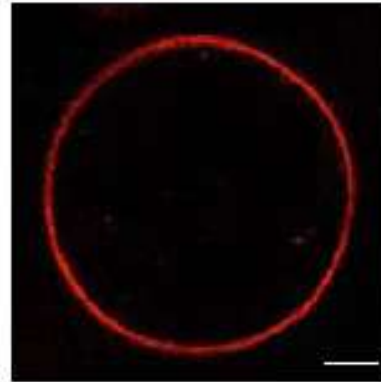


enhances BSM depletion from tube

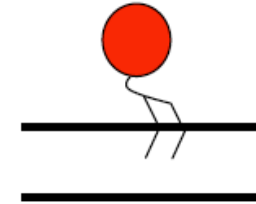
"Plasma membranes are poised for activation of raft phase coalescence at physiological temperature"



Cell blebs
(PMS or GPMV)

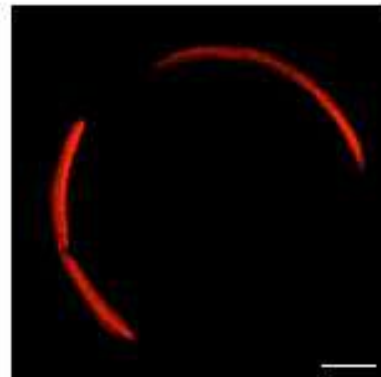


GL-GPI-mRFP



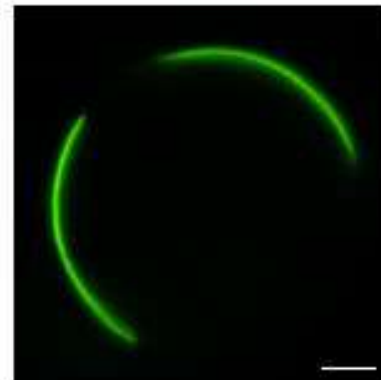
37°C

No toxin



37°C

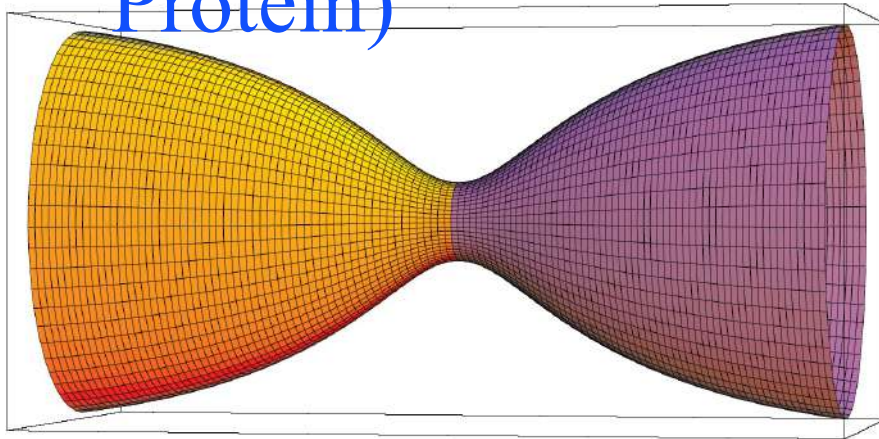
With CTxB



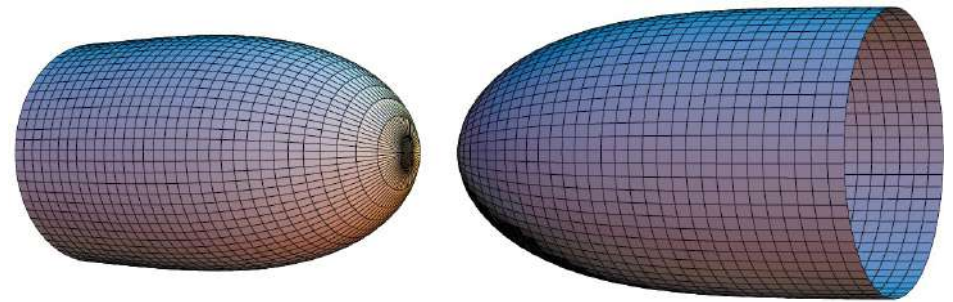
Induces phase separation

Tube Scission due to Line Tension

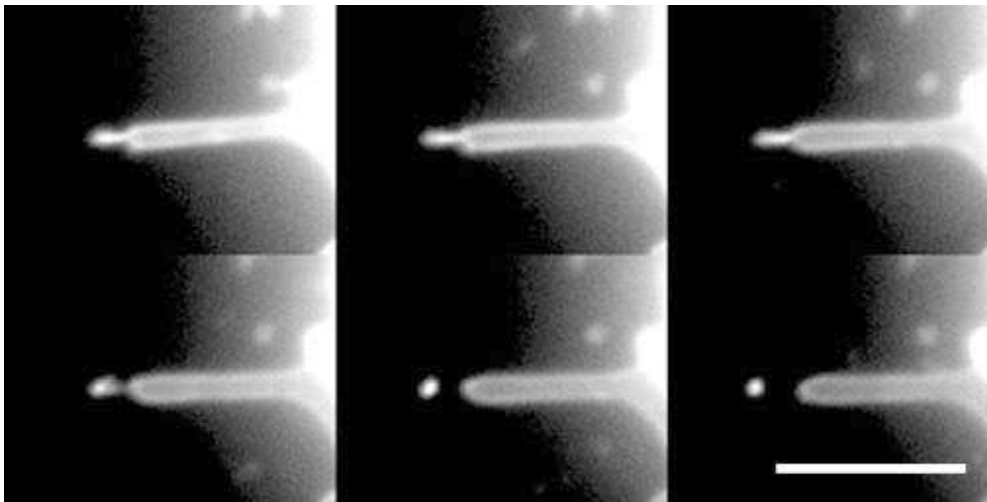
Line Tension Induces Tube Fission (NO Protein)



Line tension



J.-M. Allain, K Storm et al *PRL*.(2004) **93**, 158104

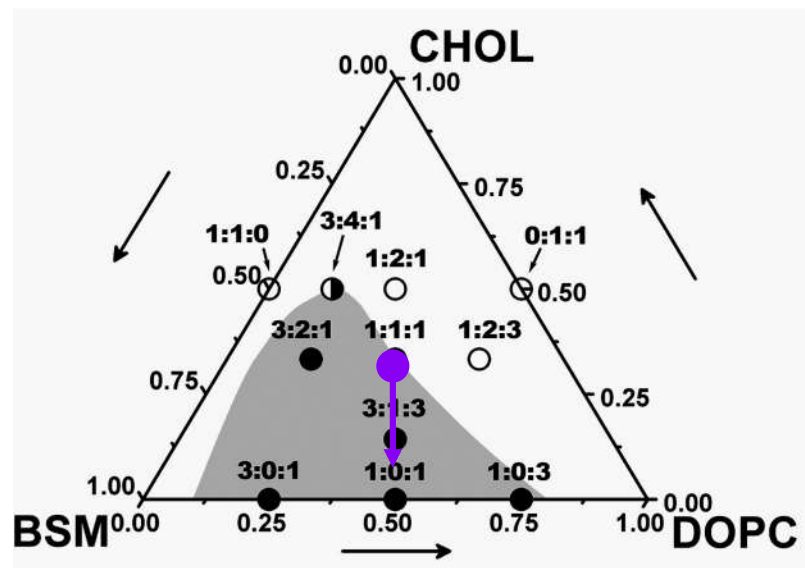


DOPC:SM:Chol 1:1:1

Cholesterol decreased

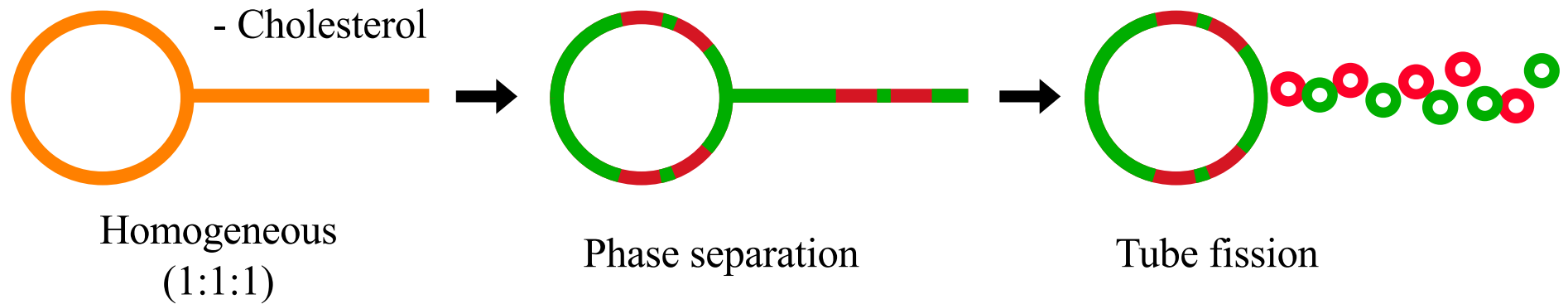
A. Roux et al *EMBO J.* (2005)

DOPC/BSM/Chol



Cholesterol decreased ➡ Phase separation

Line Tension Induces Tube Fission



Induced by photo-activation



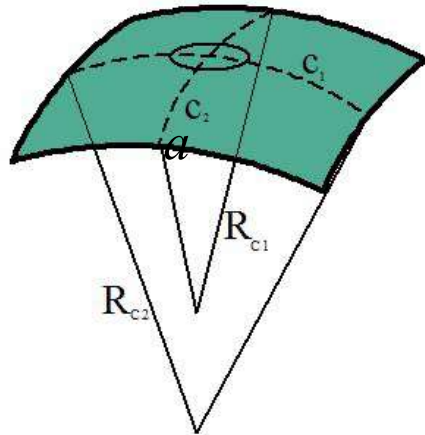
By M β CD

Canham-Helfrich free energy:

$$H = \int dA \left[\frac{1}{2} \kappa (2J)^2 + \bar{\kappa} K + \sigma \right]$$

J: Mean curvature

K: Gaussian curvature



$$J = \frac{1}{R_{c1}} + \frac{1}{R_{c2}}$$

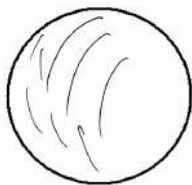
MEAN CURVATURE

$$K = \frac{1}{R_1} \cdot \frac{1}{R_2}$$

GAUSSIAN CURVATURE

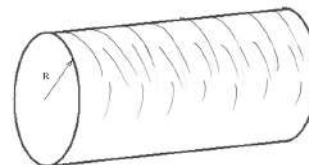
TYPICAL SHAPES

SPHERE



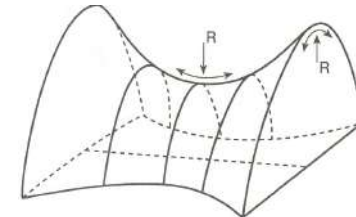
$$J = \frac{2}{R} \quad K = \frac{1}{R^2}$$

CYLINDER



$$J = \frac{1}{R} \quad K = 0$$

SADDLE



$$J = 0 \quad K = -\frac{1}{R^2}$$

Canham-Helfrich free energy:

$$H = \int dA \left[\frac{1}{2} \kappa (2J)^2 + \bar{\kappa} K + \sigma \right]$$

J: Mean curvature

K: Gaussian curvature

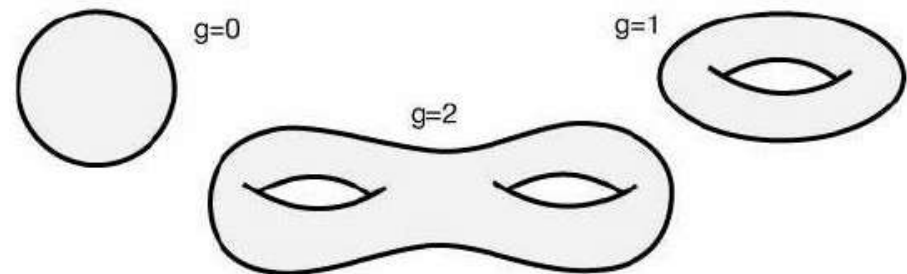
Gauss-Bonnet theorem for closed surfaces:

$$\oint K dA = 4\pi(1 - g)$$

g = number of "handles"
of the surface (genus)

sphere : g=0

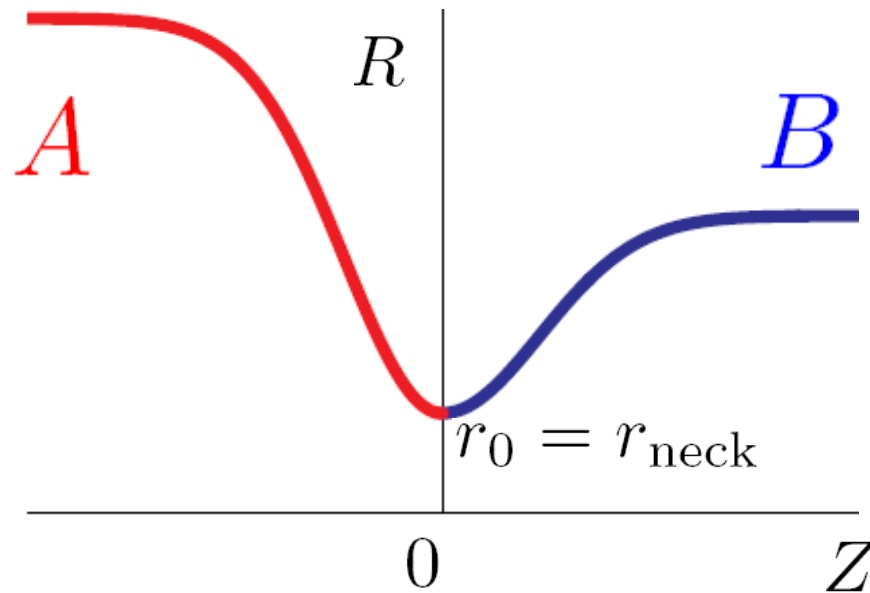
donut : g=1



total integrated Gaussian curvature = a *topological invariant*

For a tube with 2 phases A and B:

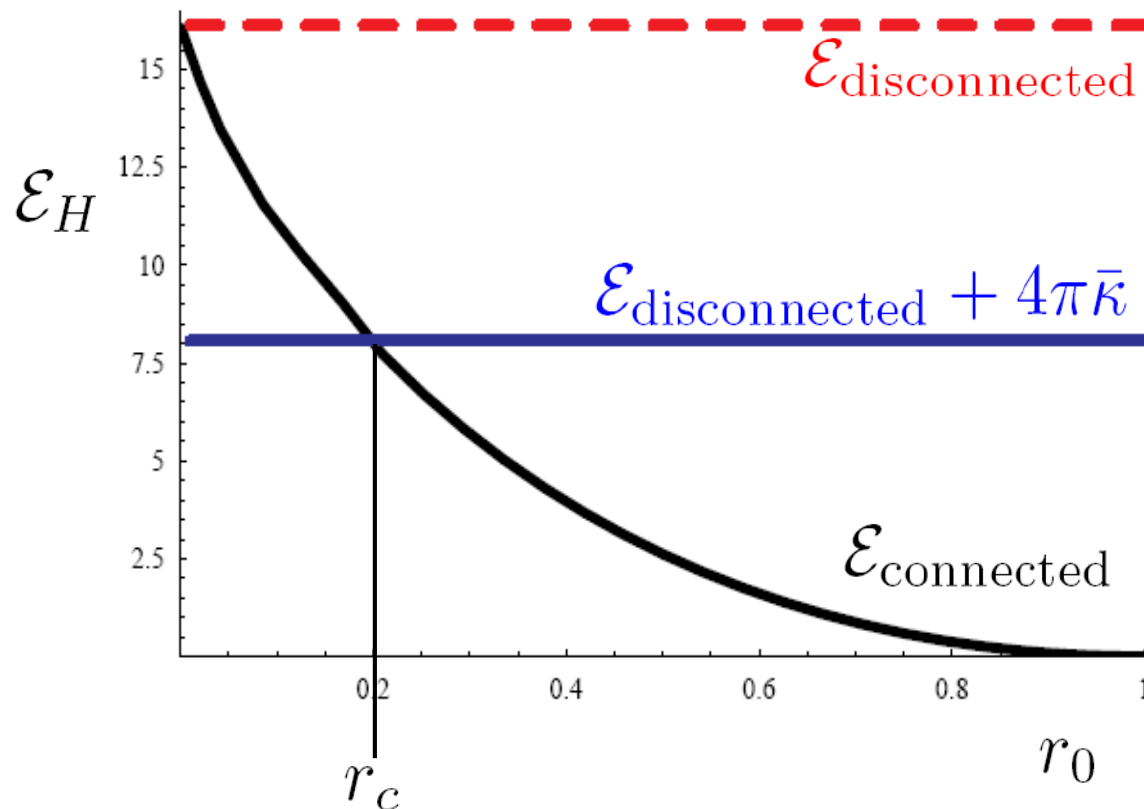
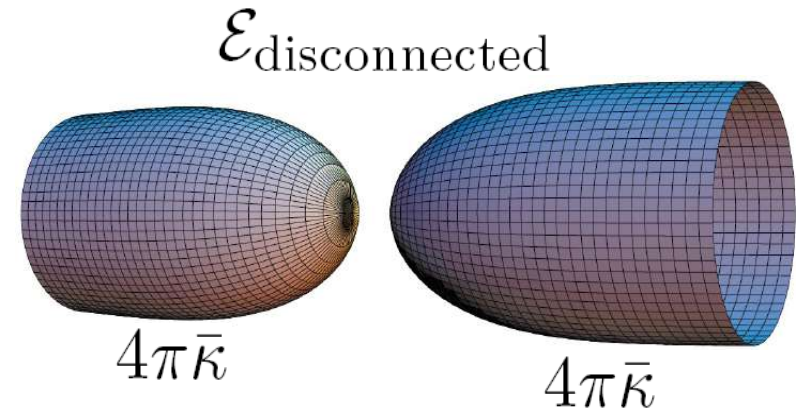
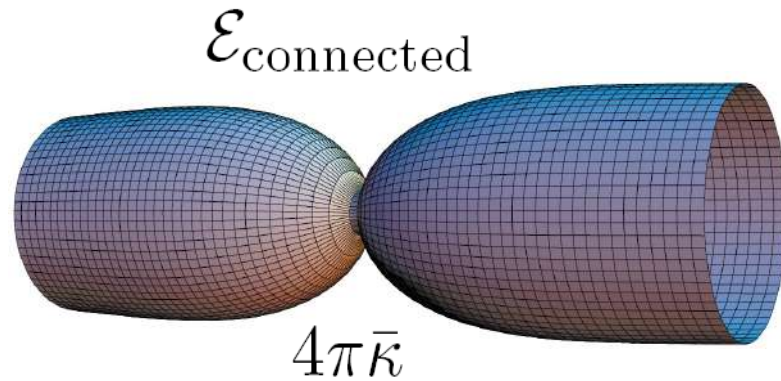
$$H = \int dA \left[\frac{1}{2} \kappa(z) (2J)^2 + \sigma(z) \right] - fL + 2\pi\gamma r_0 + F(\Delta\bar{\kappa})$$



Line tension induces
constriction (neck)

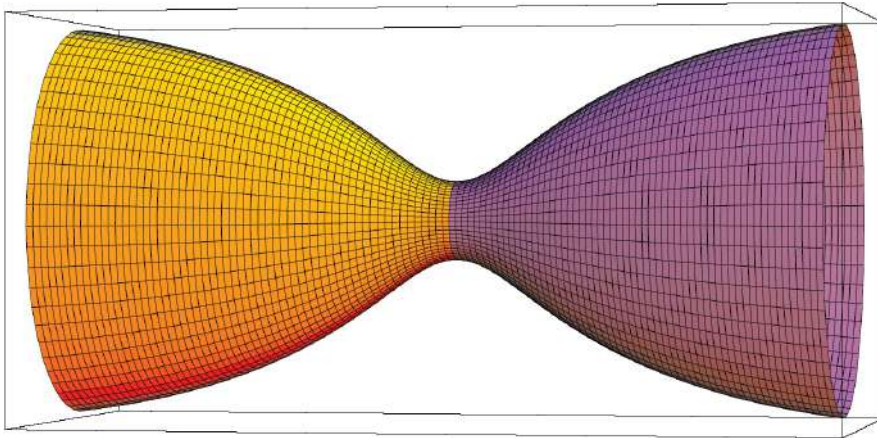
Fission Energy Barrier

Topology changes upon fission

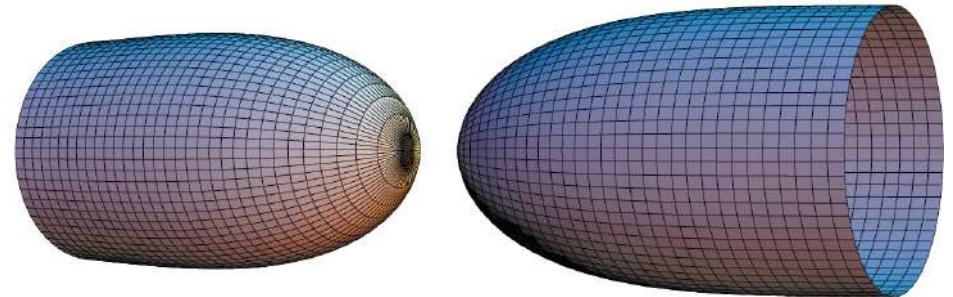
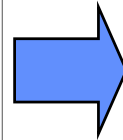


Fission at a *finite tube radius*
 $r_c \neq 0$

Line Tension and Fission



Line tension



J.-M. Allain et al *PRL*.(2004)

On a **tube** ($\neq$ vesicle): **tension** decreases tube radius
decreases energy barrier
thus **facilitates** fission

Thanks!

